

,		



A Stereo-Atlas of Ostracod Shells

edited by R. H. Bate, D. J. Horne, J. W. Neale, and David J. Siveter

Volume 14, 1987

Part 1 (pp.1–72); 30th May, 1987 Part 2 (pp. 73–151); 30th November, 1987

Published by the British Micropalaeontological Society, London

Contents

1	On Cathaycythere reticulata Whatley & Zhao gen. et sp. nov.; by R. C. Whatley & Zhao Quanhong	1
2	On Sinocythere sinensis Hou; by R. C. Whatley & Zhao Quanhong	5
3	On Albileberis sinensis Hou; by Zhao Quanhong & R. C. Whatley	9
4	On Sinocytheridea impressa (Brady); by Zhao Quanhong & R. C. Whatley	13
5	On Pterygocythereis vannieuwenhuisei Brouwers sp. nov.; by E. M. Brouwers	17
6	On Muellerina hazeli Coles & Cronin sp. nov.; by G. P. Coles & T. M. Cronin	21
7	On Healdianella? aremorica Crasquin sp. nov.; by S. Crasquin	25
8	On Maghrebeis tuberculata Majoran gen. et sp. nov.; by S. Majoran	29
9	On Howeina camptocytheroidea Hanai; by N. Ikeya & E. Compton-Gooding	33
10	On Spinoleberis eximia (Bosquet); by J. F. Babinot & J. P. Colin	37
11	On Kovalevskiella caudata (Lutz); by P. Carbonel, J. P. Colin & L. Londeix	41
12	On Calocaria maurae Vannier gen. et sp. nov.; by J. Vannier	45
13	On Spinohippula esurialis Vannier, Krůta & Marek gen. et sp. nov.; by J. Vannier, M. Krůta &	
	L. Marek	49
14	On Beyrichia (Sagenabeyrichia) siveteri Pollicott subgen. et sp. nov.; by P. D. Pollicott	57
15	On Bythocythere intermedia Elofson; by D. J. Horne	65
16	On Bythocythere zetlandica Athersuch, Horne & Whittaker; by D. J. Horne	69
17	On Kuiperiana robusta Whatley & Maybury sp. nov.; by R. C. Whatley & C. Maybury	73
18	On Loxocauda subquadrata Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley	77
19	On Sagmatocythere minuta Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley	81
20	On Sagmatocythere alaefortis alaefortis Whatley & Maybury sp. nov.; by R. C. Whatley &	
	C. Maybury	85
21	On Sagmatocythere alaefortis gallica Whatley & Maybury subsp. nov.; by R. C. Whatley &	
	C. Maybury	89
22	On Sagmatocythere wyatti Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley	93
23	On Carinocythereis carinata (Roemer); by J. Athersuch & J. E. Whittaker	97
24	On Carinocythereis whitei (Baird); by J. Athersuch & J. E. Whittaker	103
25	On Abrotocythere quinquicornis Zhao gen. et sp. nov.; by Zhao Yuhong	111
26	On Abrotocythere ovata Zhao sp. nov.; by Zhao Yuhong	115
27	On Leucocythere weiningensis Zhao sp. nov.; by Zhao Yuhong	119
28	On Leucocythere plena Zhao sp. nov.; by Zhao Yuhong	123
29	On Limnocythere xinanensis Zhao sp. nov.; by Zhao Yuhong	127
30	On Metacypris aphthosa Zhao sp. nov.; by Zhao Yuhong	131
31	On Beninea ibecetenensis Apostolescu gen. et sp. nov.; by V. Apostolescu	135
32	On Glyptolichvinella spiralis (Jones & Kirkby); by R. F. Lundin	139
33	On Glyptolichvinella ovicella Lundin & Visintainer sp. nov.; by R. F. Lundin & L. M. Visintainer	143
34	Index for Volume 14, 1987	149

A Stereo-Atlas of Ostracod Shells

edited by R. H. Bate, D. J. Horne, J. W. Neale,

and David J. Siveter Volume 14, Part 1; 30th May, 1987

Published by the British Micropalaeontological Society, London

Editors

Dr R.H. Bate, SSI (UK) Ltd., Tannery House, Tannery Lane, Send, Woking, Surrey GU23 7EF. Dr D.J. Horne, Department of Geology, City of London Polytechnic, Walburgh House, Bigland Street, London E1 2NG.

Prof. J.W. Neale, Department of Geology, The University, Hull HU6 7RH.

Dr David J. Siveter, Department of Geology, The University, Leicester LE1 7RH.

Editorial Board

Dr G. Bonaduce, Stazione Zoologica, 80121 Napoli, Italy.

Dr J.-P. Colin, Esso Production Research - European, 213 Cours Victor Hugo, 33321 Bègles, France.

Dr P. De Deckker, Research School of Pacific Studies, Australian National University, PO Box 4, Canberra ACT 2600, Australia.

Dr D. van Harten, Universiteit van Amsterdam, Geologisch Instituut, Nieuwe Prinsengracht 130, Amsterdam, The Netherlands.

Dr I. Purper, Departamento de Paleontologia e Estratigrafia, UFRGS, 90 000 Porto Alegre RS, Brazil. Dr R.E.L. Schallreuter, Universität Hamburg, Geologisch-Paläontologisches Institut, Bundesstrasse 55, D 2000 Hamburg 13, West Germany.

Officers of the British Micropalaeontological Society

Chairman Dr A.C. Higgins, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN.

Secretary Dr P.P.E. Weaver, Institute of Oceanographic Sciences, Brook Road, Wormley, Godalming, Surrey GU8 5UB. Tel: 0428-79 4141.

Treasurer Dr J.E. Whittaker, Department of Palaeontology, British Museum (Natural History), Cromwell Road, London SW7 5BD. Tel: 01-589 6323.

Journal Editor Dr. L.M. Sheppard, SSI (U.K.) Limited, Chancellor Court, 20 Priestly Road, Guildford, Surrey GU2 5YL. Tel: (0483) 506605.

Newsletter Editor Dr R.L. Austin, Department of Geology, University of Southampton, Southampton SO9 5NH. Tel: (0703) 559122/557941

Conodont Group Chairman Dr R.J. Aldridge, Department of Geology, University of Nottingham, University Park, Nottingham NG7 2RD.

Secretary Dr P.M. Smith, Department of Earth Sciences, University of Cambridge, Downing Street, Cambridge CB2 3EQ. Tel: (0223) 355463 (or 276121).

Foraminifera Group Chairman Dr P. Copestake, Britoil, 150 St. Vincent Street, Glasgow G2 5LJ. Secretary Dr D.J. Shipp, Robertson Research Int. Limited, Ty'n-y-Coed, Llanrhos, Llandudno LL30 1SA. Tel: (0492) 81811.

Microplankton Group Chairman Dr G.L. Eaton, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN.

Secretary Dr A.J. Powell, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN. Tel: (09327) 62818.

Ostracod Group Chairman Dr D.J. Horne, Geology Department, City of London Polytechnic, Walburgh House, Bigland Street, London E1 2NG.

Secretary Dr C. Maybury, Department of Geology, University College of Wales, Aberystwyth, Dyfed SY23 3DB. Tel: (0970) 3111.

Palynology Group Chairman Dr M.C. Boulter, N.E. London Polytechnic, Romford Road, London E15 4LZ.

Secretary Dr J.E.A. Marshall, Department of Geology, The University, Southampton SO9 5NH. Tel: (0703) 559122.

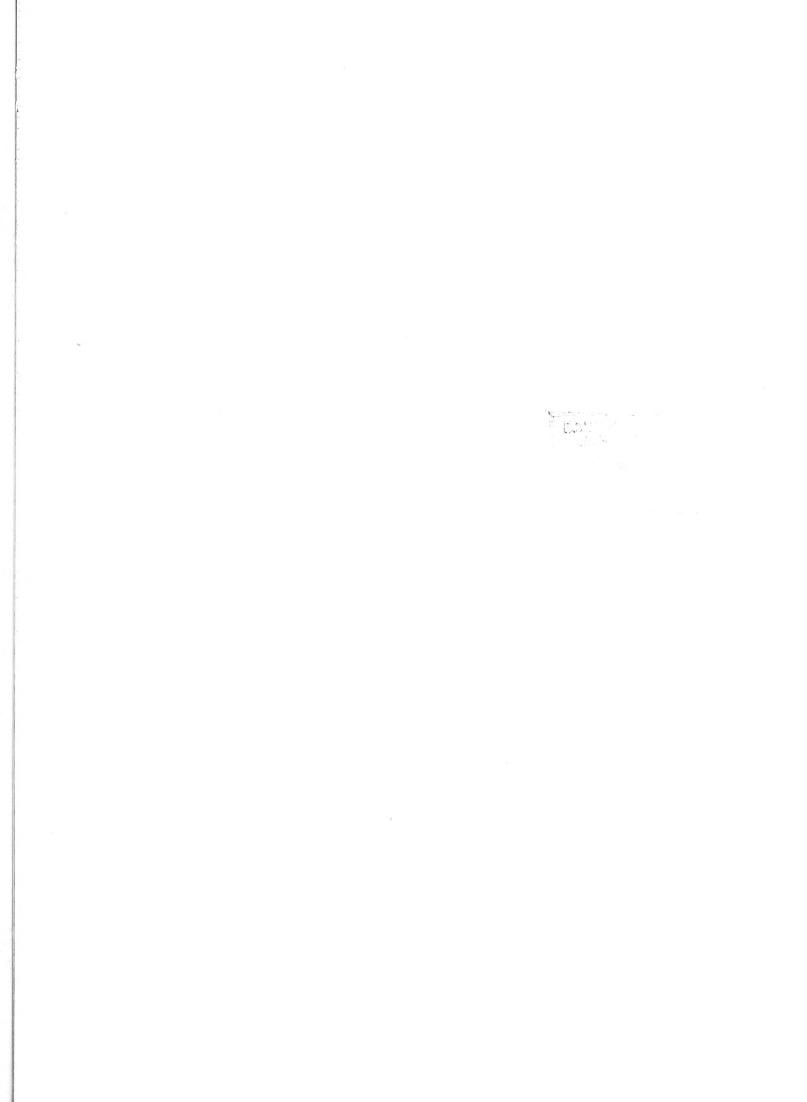
Calcareous Nannofossil Group Chairman Mr M. Jakubowski, Robertson Research Int. Limited, Ty'n-y-Coed, Llanrhos, Llandudno, Gwynedd LL30 1SA.

Secretary Dr J. Crux, BP Research Centre, Chertsey Road, Sunbury on Thames, Middlesex TW16 7LN. Tel: (09327) 63062.

Instructions to Authors

Contributions illustrated by scanning electron micrographs of Ostracoda in stereo-pairs are invited. Format should follow the style set by the majority of papers in this issue. Descriptive matter apart from illustrations should be cut to a minimum; preferably each plate should be accompanied by one page of text only. Blanks to aid in mounting figures for plates may be obtained from any one of the Editors or Editorial Board. Completed papers should be sent to Dr David J. Siveter.

The front cover shows a left valve of *Neolimnocythere hexaceros* Delachaux, 1928, from Quaternary Deposits at Lago Junin, Peru. Photograph by Dr P. De Deckker, University of Monash, Victoria, Australia.



ON CATHAYCYTHERE RETICULATA WHATLEY & ZHAO gen. et sp. nov.

by Robin Whatley & Zhao Quanhong

(University College of Wales, Aberystwyth, UK & Tongji University, Shanghai, China)

Genus CATHAYCYTHERE gen. nov.

Type-species: Cathaycythere reticulata gen. et sp. nov.

Derivation of name: From Cathay (China) + cythere; with reference to the type locality of the type-species in the South

China Sea.

Diagnosis:

Medium sized; subrectangular in lateral view, highest anterodorsally and longest subventrally; anterior margin broadly rounded; posterior margin truncate above and bluntly pointed below. Carapace laterally compressed at both end margins. Eye-tubercle and internal ocular sinus absent. Surface coarsely and irregularly reticulate with a prominent subcentral node surrounded by a subcircular sulcus. Hinge holamphidont with high conical anterior tooth, bulbous posterior tooth and locellate median element in right valve. Inner lamella wide with a marked oval excavation posteriorly aligned parallel to the postero-dorsal slope of the posterior margin; very narrow vestibulae at each end; radial pore canals few, long, thin and simple. Adductor scars relatively small, consisting of a vertical row of four; frontal scar single, oval.

Explanation of Plate 14, 2

Fig. 1, RV, ext. lat. (holotype, 1986.404, 710 μ m long); fig. 2, LV, ext. lat. (paratype, 1986.405, 665 μ m long); fig. 3, LV, ext. lat. (paratype, 1986.406, 665 μ m long). Scale A (100 μ m; ×90), figs. 1-3.

Stereo-Atlas of Ostracod Shells 14, 3

Cathaycythere reticulata (3 of 4)

Remarks:

The excavation on the wide posterior inner lamella is not known to occur in any other genus and can be used to distinguish it from *Sinocythere* Hou, 1978. The latter is closest in ornament and muscle scar pattern to the present genus, but differs in hingement (hemiamphidont with clearly crenulate posterior tooth) and in lacking the posterior excavation on the inner lamella. *Krithe* exhibits a similar excavation posteriorly but it occurs distal to the selvage whereas in *Cathaycythere* it is proximal. The familial status of *Cathaycythere* is uncertain and possibly a new family is required to accommodate this genus and *Sinocythere* (see Whatley & Zhao, *Stereo-Atlas Ostracod Shells* 14, 5-8, 1987).

Cathavcythere reticulata Whatley & Zhao gen. et sp. nov.

Holotype:

British Museum (Nat. Hist.) no. 1986.404; RV.

[Paratypes: British Museum (Nat. Hist.) nos. **1986.405-409.**]

Type locality:

Off Guangsi Province of China, Gulf of Tonkin, South China Sea; lat. 21° 29′ 09"N, long. 108° 44′

46"E. Recent, water depth: 14m.

Derivation of name:

With reference to its reticulate surface.

Figured specimens:

British Museum (Nat. Hist.) nos. **1986.404** (holotype, RV: Pl. **14**, 2, fig. 1), **1986.405** (paratype, LV: Pl. **14**, 2, fig. 2), **1986.406** (paratype, LV: Pl. **14**, 2, fig. 3), **1986.407** (paratype, car.: Pl. **14**, 4, fig. 1), **1986.408** (paratype, LV: Pl. **14**, 4, fig. 2), **1986.409** (paratype, RV: Pl. **14**, 4, fig. 3). Nos. **1986.404-406** are from the type locality. Nos. **1986.407-409** are from lat. 21° 15′ 46″N, long. 109°

24' 57"E; Recent, water depth: 14m.

Diagnosis:

As for the genus. Monotypic.

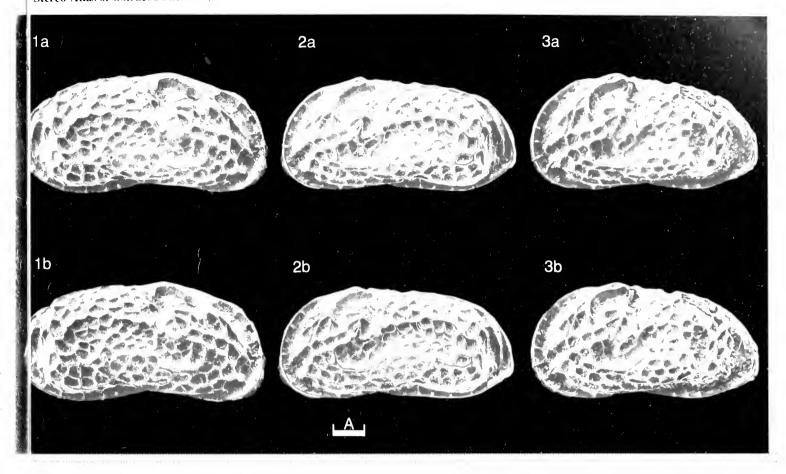
Distribution:

In four bottom samples off Guangsi Province, China, northern Gulf of Tonkin, South China Sea.

Water depth: 10 - 16m; substrate: mud to fine sand.

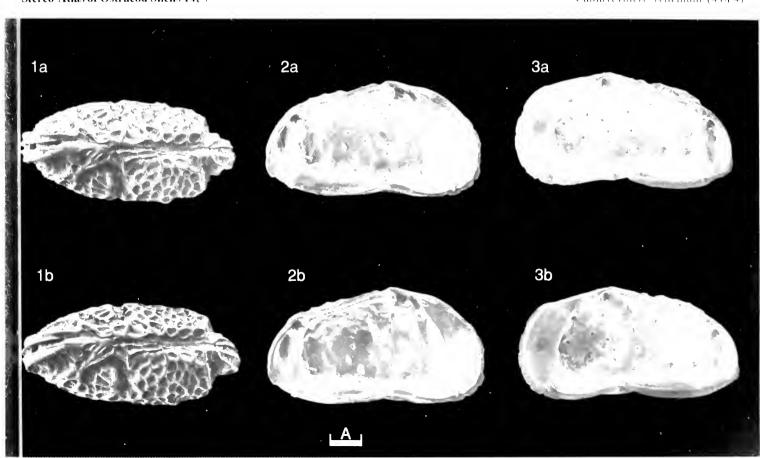
Explanation of Plate 14, 4

Fig. 1, car., ext. dors. (paratype, 1986.407, 635 μ m long); fig. 2, LV, int. lat. (paratype, 1986.408, 635 μ m long); fig. 3, RV, int. lat. (paratype, 1986.409, 645 μ m long). Scale A (100 μ m; ×90), figs. 1-3.



Stereo-Atlas of Ostracod Shells 14, 4

Cathayeythere reticulata (4 of 4)



ON SINOCYTHERE SINENSIS HOU

by Robin Whatley & Zhao Quanhong (University College of Wales, Aberystwyth, UK & Tongji University, Shanghai, China)

Genus SINOCYTHERE Hou, 1982

Type-species (by original designation): Sinocythere sinensis Hou, 1982

Diagnosis: Medium sized; subrectangular in lateral view with parallel dorsal and ventral margins, prominent posterior cardinal angle and obtusely rounded posterior margin. Eye-tubercle weakly developed but internal ocular sinus not developed. Surface reticulate with an anteromedian node surrounded

but internal ocular sinus not developed. Surface reticulate with an anteromedian node surrounded by a subcircular sulcus. Hinge hemiamphidont: anterior tooth in right valve conical, posterior tooth curved and distinctly dentate; in left valve, anterior socket enclosed ventrally by an anterior extension of the anteromedian conical tooth, posterior socket with an anti-slip toothlet ventromedianly, and median bar denticulate. Inner lamella relatively wide with shallow anterior vestibule; radial pore canals few, thin and simple. Adductor muscle scars small, consisting of a

vertical row of four scars all in contact; frontal scar single, oval.

Remarks: This genus is close to Spinoleberis Hanai, 1961 in many features except that the latter has a triangular outline and much narrower, more acute posterior margin in lateral view, and three longitudinal ribs. In external characters Sinocythere is somewhat similar to Palmenella Hirschman, 1916, but the latter bears a schizodont hinge. Cathaycythere Whatley & Zhao, 1987 (Stereo-Atlas October 1984) has circilar constants to the right of the right of the simple restants the

Ostracod Shells 14, 1-4) has similar ornament and also the circular sulcus surrounding the anteromedian node. The two genera differ in hingement and Sinocythere lacks the excavated posterior inner lamella so typical of Cathaycythere. Sinocythere and Cathaycythere are probably

worth including in a new family of Cytheracea.

Explanation of Plate 14, 6

Fig. 1, \circlearrowleft RV, ext. lat. (1986.410, 570 μ m long); fig. 2, \mathring{Q} car., rt. lat. (1986.411, 540 μ m long); fig. 3, \mathring{Q} car., lt. lat. (1986.411, 540 μ m long). Scale A (100 μ m; ×110), figs. 1-3.

Stereo-Atlas of Ostracod Shells 14, 7

Sinocythere sinensis (3 of 4)

Sinocythere sinensis Hou, 1982

1982 Sinocythere sinensis sp. nov. Hou in Hou et al., Cretaceous-Quaternary ostracode fauna from Jiangsu, 242, text-fig. 77; pl. 87, figs. 16-19.

1985 Sinocythere sinensis Hou; Zhao, Acta Oceanologica Sinica, pl. 2, fig. 10.

1985 Sinocythere sinensis Hou; Wang and Zhao, in Wang et al., Marine Micropaleontology of China, pl. 18, fig. 4.

Holotype: Nanjing Institute Geology & Paleontology, Academia Sinica; no. 4107, ♀ LV. Not figured herein.

Type locality: Jiangsu Province, Eastern China; Dongtai Formation, Quaternary.

Figured specimens: British Museum (Nat. Hist.) nos. 1986.410 (O'RV: Pl. 14, 6, fig. 1; Pl. 14, 8, fig. 3), 1986.411 (Q

car.: Pl. 14, 6, figs. 2, 3; Pl. 14, 8, fig. 1), 1986.412 (♀ LV: Pl. 14, 8, fig. 2). All Recent, collected from the littoral of Jiangsu Province, China; approx. lat. 34° 17′N, long. 120° 17′E.

Diagnosis: Irregularly polygonal surface reticulation with superimposed narrow, oblique posterodorsal-

anteroventral rib, and a bifid rib running from the anterior cardinal angle obliquely to mid-anterior. Female carapace strongly laterally compressed posterodorsally, posteriorly and ventrally; the male is inflated in these areas.

ventrally, the male is initiated in these aleas

Remarks: This species is very close to S. dongtaiensis Chen, 1982 in outline and overall ornamentation, but

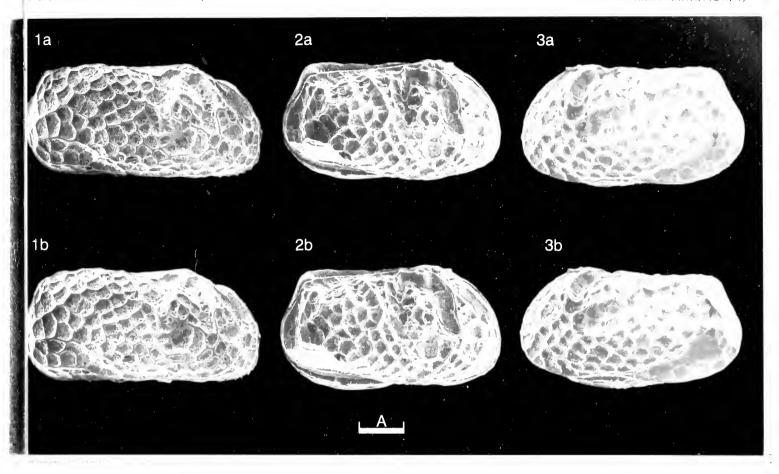
the latter is much more weakly reticulate.

Distribution: Pliocene to Recent, Eastern China. Recent specimens most abundant in littoral zone and inner

shelf shallower than 20m, rare in estuaries and water depths from 20 to 200m.

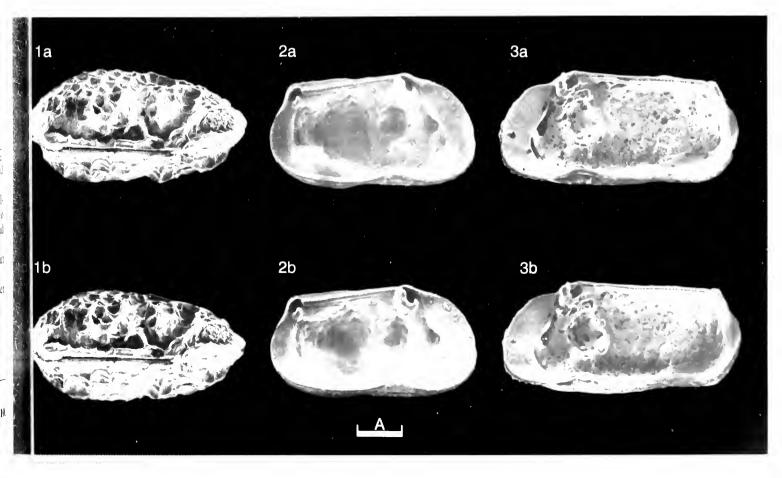
Explanation of Plate 14, 8

Scale A (100 μ m; ×110), figs. 1-3.



Stereo-Atlas of Ostracod Shells 14, 8

Sinocythere sinensis (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (3) 9–12 (1987)

595.337.14 (119) (510 : 161.120.34) : 551.313.1 + 551.35

ON ALBILEBERIS SINENSIS HOU

by Zhao Quanhong & Robin Whatley (Tongji University, Shanghai, China & University College of Wales, Aberystwyth, UK)

Genus ALBILEBERIS Hou, 1982

Type-species: Albileberis sinensis Hou, 1982

Diagnosis:

Small to medium, laterally compressed; subovate to rectangular in lateral view with greatest height anteromedianly. Bluntly truncate posterior and well rounded anterior margins. Surface smooth, pitted or reticulate. Hinge between paleomerodont and holomerodont; all positive elements in right valve, long ridge-like anterior tooth thickened anteriorly with a prominent square terminal toothlet, median bar smooth anteriorly and faintly crenulate posteriorly, posterior tooth curved and denticulate. Complementary negative elements in left valve with an anti-slip bar below median groove and anterior socket, and two small terminal anti-slip toothlets respectively at anterior and posterior ends. Inner lamella moderately wide with large vestibulae and narrow fused zone; radial pore canals moderate in number and simple. Adductor muscle scars a vertical row of four; frontal scar V-shaped; fulcral point crescentic and prominent.

Remarks:

Although the generic name Albileberis was first published in Guan et al. (Paleontological Atlas of Central & S China (4): Ostracoda, 1978), who attributed the genus to Hou from an earlier MS name, its type-species was not published until 1982 (Hou in Hou et al.). This genus is easily identified by its laterally compressed carapace, outline and peculiar hingement, and probably belongs to the Cytherideinae based on its internal characters.

Explanation of Plate 14, 10

Fig. 1, \bigcirc RV, ext. lat. (1986.413, 525 μ m long); fig. 2, \bigcirc LV, ext. lat. (1986.414, 525 μ m long); fig. 3, \bigcirc LV, ext. lat. (1986.415, 495 μ m long).

Scale A (100 μ m; ×120), figs. 1-3.

Stereo-Atlas of Ostracod Shells 14, 11

Albileberis sinensis (3 of 4)

Albileberis sinensis Hou, 1982

1982 Albileberis sinensis sp. nov. Hou, in Hou et al., Cretaceous-Quaternary ostracode fauna from Jiangsu., 240-241, text-fig. 75, pl. 88, figs. 1-7.

1985 Albileberis sinensis Hou; Zhao, Acta Oceanological Sinica, pl. 1, fig. 9.

1985 Albileberis sinensis Hou; Wang & Zhao, in Wang et al., Marine Micropaleontology of China, pl. 7, fig. 2, text-fig. 5.

Holotype: Nanjing Institute Geology & Paleontology, Academia Sinica; no. 41062; ♀ LV. Not figured herein.

Type locality: Jiangsu Province, E China; Dongtai Formation, Quaternary.

Figured specimens: British Museum (Nat. Hist.) nos. 1986.413 (♀ RV: Pl. 14, 10, fig. 1), 1986.414 (♀ LV: Pl. 14, 10,

fig. 2), 1986.415 (\circlearrowleft LV: Pl. 14, 10, fig. 3), 1986.416 (\updownarrow car.: Pl. 14, 12, fig. 1), 1986.417 (\updownarrow LV: Pl. 14, 12, fig. 2), 1986.418 (\updownarrow RV: Pl. 14, 12, fig. 3). All Recent, collected from the littoral of Jiangsu

Province, China; approx. lat. 34° 17'N, long. 120° 17'E.

Diagnosis: Subovate in lateral view with vertical truncated posterior margin, surface smooth with few very

weak reticulae around the margins.

Remarks: Markedly differs from the other two species in this genus. A. sheyangensis Chen, 1982 is much

more elongate and lower with obliquely truncated posterior margin, and A. asperata Guan, 1978

has an ornamentation of coarse reticulation.

Distribution: China, Quaternary to Recent. At the present day this species is abundant and widespread in

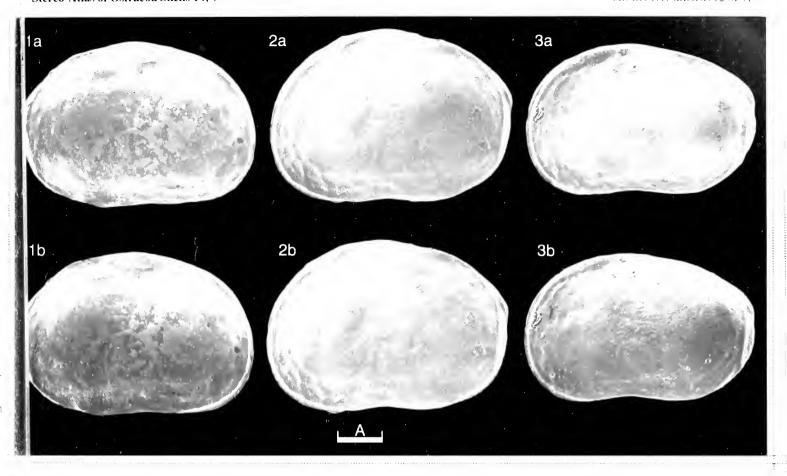
brackish and nearshore waters along the coast of the East China and Yellow Seas with a salinity range of 3‰ to normal sea water, including marshes, estuaries, littoral and the inner shelf

shallower than 50m.

Explanation of Plate 14, 12

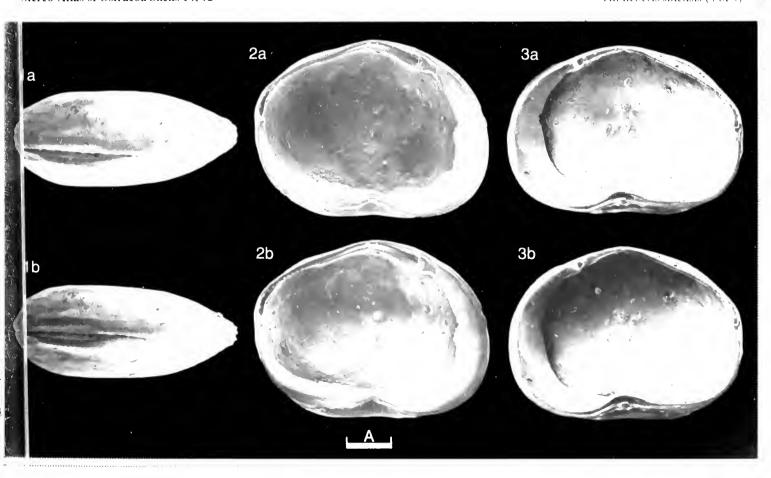
Fig. 1. \heartsuit car., ext. dors. (1986.416, 535 μ m long); fig. 2, \heartsuit LV, int. lat. (1986.417, 515 μ m long); fig. 3, \heartsuit RV, int. lat. (1986.418, 515 μ m long).

Scale A ($100\mu m$; ×120), figs. 1-3.



Stereo-Atlas of Ostracod Shells 14, 12

Albileberis sinensis (4 of 4)



ON SINOCYTHERIDEA IMPRESSA (BRADY)

by Zhao Quanhong & Robin Whatley (Tongji University, Shanghai, China & University College of Wales, Aberystwyth, UK)

Genus SINOCYTHERIDEA Hou, 1978

Type-species (by subsequent designation): S. latiovata Hou & Chen, 1982 (= Cytheridea impressa Brady, 1869; see below)

A genus of Cytherideidae characterized by its modified antimerodont hingement with a Diagnosis:

conspicuous anti-slip toothlet anteriorly in the left valve, by which it can be readily distinguished from such similar genera as Cyprideis Jones, 1857, Neocyprideis Apostolescu, 1965 and

Sarsicytheridea Athersuch, 1982.

Sinocytheridea was named by Hou in manuscript more than 20 years ago but remained Remarks:

unpublished until Guan et al. (1978) first applied this name for the genus in a published work, attributing the genus to Hou and using Hou's original description. S. latiovata Hou & Chen, 1982 was designated by Hou and Chen in Hou et al. (1982) as the type-species of Sinocytheridea. The present authors, however, have recently studied Brady's material from Hong Kong which is deposited in the Hancock Museum and consider S. latiovata and Cytheridea impressa Brady, 1869 to be conspecific. We therefore consider C. impressa to be the type-species of Sinocytheridea.

Explanation of Plate 14, 14

Figs. 1, \mathcal{Q} car., rt. ext. lat (paralectotype, 1.23.44, 740 μ m long); fig. 2, \mathcal{Q} LV, ext. lat. (1986.419, 720 μ m long); fig. 3, \mathcal{Q} car., ext. dors. (1986.421, 750 μm long).

Scale A (100 μ m; × 85), figs. 1-3.

Stereo-Atlas of Ostracod Shells 14, 15

Sinocytlieridea impressa (3 of 4)

Sinocytheridea impressa (Brady, 1869)

Cytheridea impressa sp. nov. G.S. Brady in L. De Folin and L. Perier (eds.), Les Fonds de la Mer., 158, pl. 16, figs. 13, 14.

Cyprideis velii Hu & Yeh, Geol. Soc., China (Taiwan), Proc., 21, 157-159, text-fig. 5, pl. 3, figs. 10-13.

1978 Sinocytheridea sinensis Hou; Hou in Guan et al., Paleontological Atlas Central & S China (4); Ostracoda, 240, pl. 65, figs. 1-5.

Sinocytheridea latiovata sp. nov. Hou & Chen in Hou et al., Cretaceous-Ouaternary ostracode fauna from Jiangsu, 164-165, text-figs. 26a-c, pl. 72, figs. 10-20.

1982 Sinocytheridea longa Hou & Chen, ibid. 165-166, text-figs. 27a-c, pl. 72, figs. 1-9.

Lectotype: Hancock Museum, Newcastle-upon-Tyne, England, no. 1.24.37, ♀ LV.

[Paralectotypes: Hancock Mus., nos. 1.24.38, Q RV; 1.23.44, Q car.]

Type locality: Hong Kong Harbour: Recent.

Figured specimens: Hancock Museum, Newcastle-upon-Tyne, England, nos. 1.24.37 (lectotype, ♀ LV: Pl. 14, 16, fig.

2), 1.24.38 (paralectotype, ♀ RV: Pl. 14, 16, fig. 3), 1.23.44 (paralectotype, ♀ car.: Pl. 14, 14, fig. 1), Brit. Mus. (Nat. Hist.) nos. 1986.419 (\$\Q\$ LV: Pl. 14, 14, fig. 2), 1986.421 (\$\Q\$ car.: Pl. 14, 14, fig.

3), 1986.420 (O LV: Pl. 14, 16, fig. 1). Nos. 1.23.44, 1.24.37 and 1.24.38 belong to the Brady Collection in the Hancock Museum and are from the type-locality; nos. 1986.419-431 are from the

Pohai Bya, China, lat. 39° 50'N, long. 118° 46'E, water depth: 15m.

Elongate-oval without obvious trace of angle in lateral view. RV larger than and slightly Diagnosis: overlapping LV along the periphery except anterior margin. Surface smooth with rounded shallow

pits (openings of sieve-type normal pore canals). Avestibulate, radial pore canals few, simple.

Adductor muscle scar a vertical row of four elongate scars; frontal scar V-shaped.

Remarks: Apart from the more elongate outline, S. longa Hou & Chen is identical in carapace features to S.

impressa. The authors believe that the former species is based on males of S. impressa.

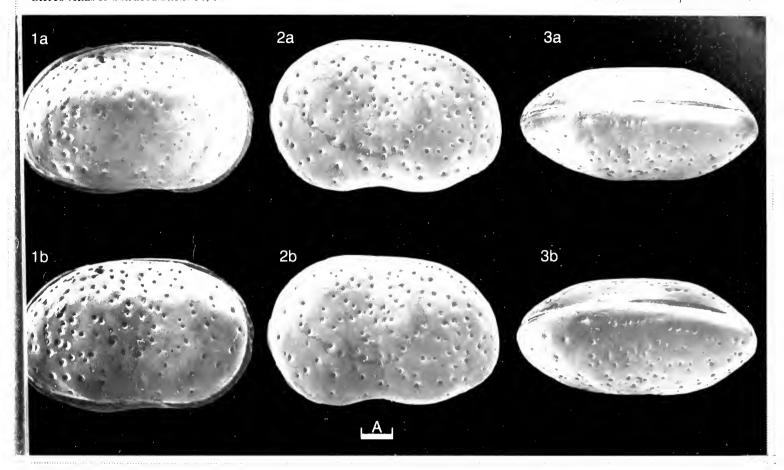
Distribution: Pliocene to Recent, China. The modern representatives occur widely in shelf, littoral, estuaries,

marshes, tidal pools and channels of the supralittoral zone along the entire coast of China with a salinity distribution ranging from about 2% to normal sea water and a water depth ranging from

middle shelf (50-100m) to supralittoral.

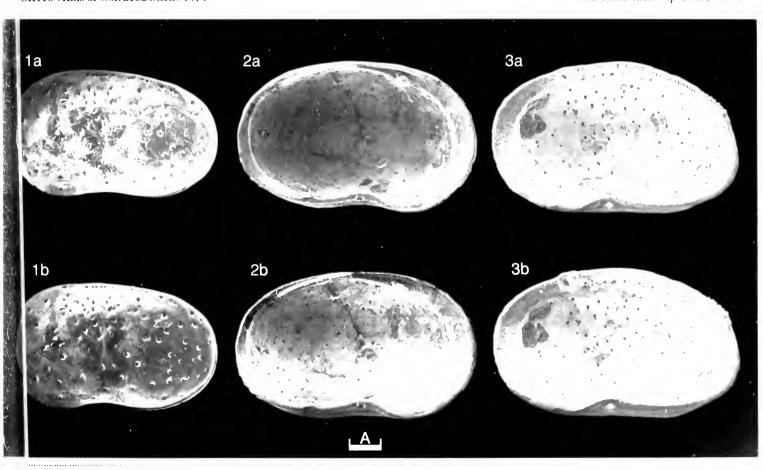
Explanation of Plate 14, 16

Fig. 1, \circlearrowleft LV, ext. lat. (1986.420, 660 μ m long); fig. 2, \circlearrowleft LV, int. lat. (lectotype, 1.24.37, 775 μ m long); fig. 3, \circlearrowleft RV, int. lat. (paralectotype, 1.24.38, 740 μ m long). Scale A (100 μ m; × 85), figs. 1-3.



Stereo-Atlas of Ostracod Shells 14, 16

Sinocytheridea impressa (4 ot 4)



ON PTERYGOCYTHEREIS VANNIEUWENHUISEI BROUWERS sp. nov.

by Elisabeth M. Brouwers (U.S. Geological Survey, Denver)

Ptervgocythereis vannieuwenhuisei sp. nov.

1986 Pterygocythereis sp. nov. C. A. Repenning, E. M. Brouwers, et al., Bull. U.S. Geol. Surv., 1687, pl. 1, fig. 1.

Holotype: U.S. National Museum no. 410130, of RV.

[Paratypes: U.S. National Museum nos. 410131-410134].

Type locality: Cutbank on a tributary of the Kalikpik River, Arctic coastal plain, North Slope, Alaska (lat. 70°

26.7'N, long. 152° 09.4'W); Pliocene. Outcrop consists of 1.8m of late Pliocene marine clay and sand overlain by 5.5m of Pleistocene fluvial and eolian sands. Deeper inner sublittoral to middle

sublittoral water depths; cold temperate to subfrigid marine climate.

In honour of Don Van Nieuwenhuise, research geologist at Amoco Production, Houston. Derivation of name: Figured specimens:

U.S. National Museum nos. 410130 (holotype, ♂ RV: Text-fig. 1), 410131 (paratype, ♀ LV: Pl. 14, 18, fig. 1), 410132 (paratype, ♂ RV: Pl. 14, 18, fig. 2), 410133 (paratype, ♂ LV: Pl. 14, 20, fig. 1), 410134 (paratype, LV: Pl. 14, 20, fig. 2). All from the type locality and horizon (locality

83-EB-187, 188, collected by E. Brouwers, 1983).

Short, high, rectangular lateral outline; large size; weak dimorphism. Three pairs of spines in Diagnosis:

median valve area; large, strong marginal spines. Wide marginal flange, continuous along anterior and venter. Numerous ventral marginal spines. Spinose anterodorsal margin with weak underlying flange. Strong posteroventral spinose prolongation. Left valve hinge has anterior socket with ventral rim; elongate, U-shaped posterior socket; cylindrical posteromedian tooth; weakly

crenulate median bar.

Explanation of Plate 14, 18

Fig. 1, \mathcal{Q} LV, ext. lat. (paratype, 410131, 1380 μ m long); fig. 2, \mathcal{O} RV, ext. lat. (paratype, 410132, 1460 μ m long). Scale A (100 μ m; ×125), figs. 1, 2.

Stereo-Atlas of Ostracod Shells 14, 19

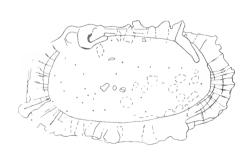
Pterygocythereis vannieuwenhuisei (3 of 4)

Remarks:

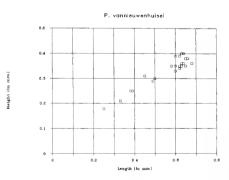
Pterygocythereis ranges from the Paleocene-Holocene, occurring commonly throughout the subtropical and temperate N Atlantic Ocean and rarely in the subfrigid Norwegian Sea. In the northwestern Atlantic Ocean, Pterygocythereis occurs in the southern cold temperate zone, but does not live in the northern cold temperate or subfrigid zones of the western N Atlantic. Pterygocythereis vannieuwenhuisei is related to the European P. mucronata-P. jonesii species complex and not to the more temperate NW Atlantic P. americana-P. inexpectata lineage.

Distribution:

?Late Miocene, early-late Pliocene (to 2.48 Ma): NE Alaska, three localities in Colvillian-aged sediments of the Gubik Fm. (Fish Creek, Kalikpik R., Miluveach Creek; 2.48 – 3.0 Ma, late Pliocene; Repenning et al., op. cit.), three localities in the upper Nuwok Member of the Sagavanirktok Fm. (Carter Creek, Barter Is., Manning Pt.; ?late Miocene, lower to middle Pliocene).



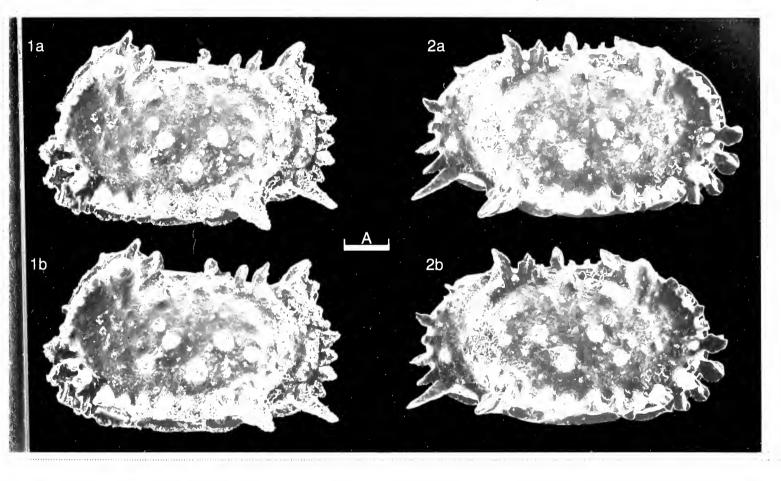
Text-fig. 1. Holotype (ORV, USNM no. 410130), camera lucida drawing, seen in transmitted light.



Text-fig. 2. Plot of length vs. height for 26 specimens from the type locality.

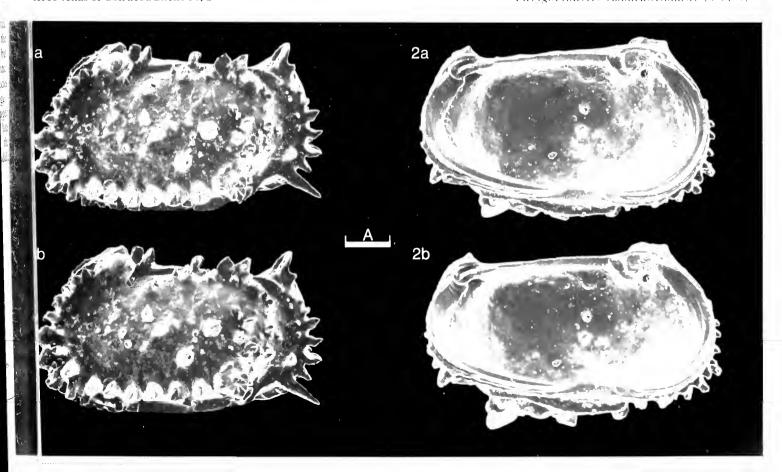
Explanation of Plate 14, 20

Fig. 1, \circlearrowleft LV, ext. lat. (paratype, 410133, 1280 μ m long); fig. 2, LV, int. lat. (paratype, 410134, 1300 μ m long). Scale A (100 μ m; ×125), figs. 1, 2.



tereo-Atlas of Ostracod Shells 14, 20

Pterygocythereis vannieuwenhuiset (4 of 4)



ON MUELLERINA HAZELI COLES & CRONIN sp. nov.

by Graham P. Coles & Thomas M. Cronin (University College of Wales, Aberystwyth & U.S. Geological Survey, Reston, Virginia)

Muellerina hazeli sp. nov.

Holotype: British Museum (Nat. Hist.) no. OS 12971, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12972-OS 12975. Four additional paratypes have been deposited in the U.S. Museum of Natural History: USNM nos.

409239-409242].

Type locality: Off the Florida Keys, United States continental slope; approx. lat. 24° 26'N, long. 81° 38'W;

Recent, water depth 107m.

Derivation of name: In honour of Joseph E. Hazel, in recognition of his studies on Muellerina from the Atlantic Coastal

Plain and shelf.

Figured specimens: British Museum (Nat. Hist.) nos. OS 12971 (holotype, Q LV: Pl. 14, 22, fig. 1; Pl. 14, 24, fig. 1),

OS 12972 (paratype, ♀ RV: Pl. 14, 22, fig. 2), OS 12973 (paratype, ♂ LV: Pl. 14, 22, fig. 3), OS 12974 (paratype, ♂ RV: Pl. 14, 24, fig. 2), OS 12975 (paratype, ♂ car.: Pl. 14, 24, fig. 3). All from

the type locality and horizon.

Explanation of Plate 14, 22

Fig. 1, \bigcirc LV, ext. lat. (holotype, OS 12971, 580 μ m long); fig. 2, \bigcirc RV, ext. lat. (paratype, OS 12972, 570 μ m long); fig. 3, \bigcirc LV, ext. lat. (paratype, OS 12973, 550 μ m long).

Scale A (100 μ m; ×130), figs. 1-3.

Stereo-Atlas of Ostracod Shells 14, 23

Muellerina hazeli (3 of 4)

Diagnosis: A species of Muellerina characterised by small size, relatively thick shell, simple subovate outline in dorsal view, and delicate ornament consisting of numerous discrete circular to ovate fossae with

several sharply defined narrow muri extending both above and below the muscle scar platform and

into the anterior field of the valve.

Remarks: M. hazeli most closely resembles M. ohmerti Hazel, 1983, but is distinctly smaller, being

equivalent in size to the A-1 instar of *M. ohmerti*, and has a distinctive, more delicately developed ornament. *M. hazeli* rarely occurs sympatrically with *M. ohmerti* north of Cape Hatteras (above 35°N), but is more abundant in deeper water on the upper continental slope, whereas *M. ohmerti* is a typical shelf species, most common at depths between 25 and 175m (Hazel 1970). *M. hazeli* is both the smallest and the most southerly distributed extant species of *Muellerina*. It is believed to have evolved from its parent species, *M. ohmerti*, during a high sea level stand in the late Pliocene

(Cronin & Coles, in prep.), and has since undergone little morphological change.

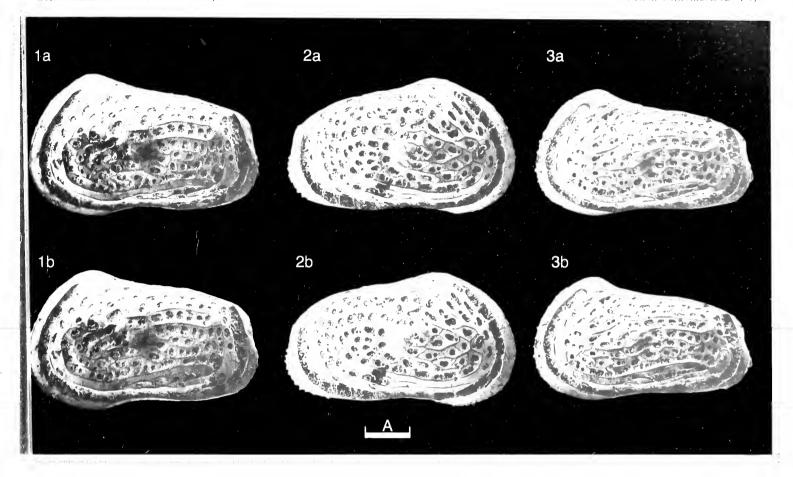
Distribution: Recent of the Atlantic continental shelf and slope from the Florida Keys (24° 25′N) to off New York at the head of Lydonia Canyon (40° 30′N). M. hazeli lives on the outer shelf and upper slope,

most commonly between 75 and 250m, having a maximum present-day depth range of 35 to 382m. It is also present in Pleistocene sediments in cores off the eastern United States from 32° 04′N to 38° 22′N, and in Pleistocene outcrops in the Norfolk and Wilmington submarine canyons.

Explanation of Plate 14, 24

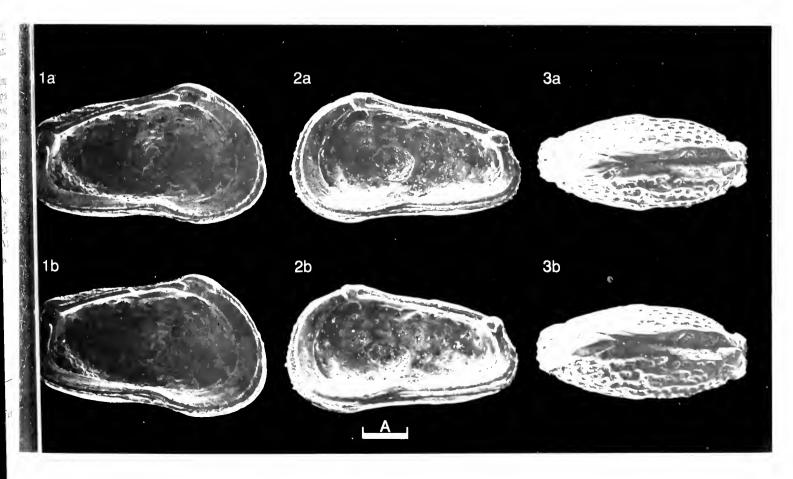
Fig. 1, \bigcirc LV, int. lat. (holotype, OS 12971, 580 μ m long); fig. 2, \bigcirc RV, int. lat. (paratype, OS 12974, 530 μ m long); fig. 3, \bigcirc car., dorsal (paratype, OS 12975, 570 μ m long).

Scale A (100 μ m; ×130), figs. 1-3.



Stereo-Atlas of Ostracod Shells 14, 24

Muellerma hazeli (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (7) 25-28 (1987) 595.337.21 (113.51) (44 : 161.002.47) : 551.351 + 552.54

ON HEALDIANELLA ? AREMORICA CRASQUIN sp. nov.

by Sylvie Crasquin (University of Lille, France)

Healdianella? aremorica sp. nov.

Holotype: University of Lille; France, ostracode Collection (COUL) no. 860, of carapace.

[Paratypes: COUL nos. 861, 862, 863, 865, 2155].

Type locality: Port Etroit Quarry (sample no. 85 MA 1), Laval syncline, Armorican Massif, France; lat. 47° 50'

54" N, long. 2° 39' 02"E. Sablé Limestone, uppermost Tournaisian, Carboniferous.

Derivation of name: From the latin aremoricus, Armorica, western province of Gaul.

Figured specimens: University of Lille, France, ostracode collection (COUL) nos. 860 (holotype, of car.: Pl. 14, 26,

fig. 1), 862 (♀ car.: Pl. 14, 26, fig. 2), 863 (♀ car.: Pl. 14, 26, fig. 3), 865 (♀ car.: Pl. 14, 28, fig. 1), 861 (juv. car.: Pl. 14, 28, fig. 2), 864 (juv. car.: Pl. 14, 28, fig. 3), 2155 (♂ car.: Pl. 14, 28, fig. 4). All from the Sable Limestone of type locality; uppermost Tournaisian, lower Carboniferous.

Explanation of Plate 14, 26

Fig. 1, \bigcirc car., rt. lat. (holotype, COUL 860, 0.58 mm long); fig. 2, \bigcirc car., rt. lat. (paratype, COUL 862, 0.58 mm long); fig. 3, \bigcirc car., rt. lat. (paratype, COUL 863, 0.60 mm long).

Scale A (200 μ m; × 140), fig. 1; scale B (200 μ m; × 85), figs. 2, 3.

Stereo-Atlas of Ostracod Shells 14, 27

Healdianella? aremorica (3 of 4)

Diagnosis:

Small, smooth species (adults 0.52-0.63 mm long) doubtfully assigned to *Healdianella*. Anterodorsal border straight; anterior border with a maximum of convexity located between 1/2 and lower 1/3 of valve height; ventral border is concave, with maximum concavity located in the anterior 1/3 of valve length; posterior border broadly rounded with maximum convexity located slightly below mid-height. In dorsal view, the carapace is laterally compressed in the medial region. Overlap is weak.

Sexual dimorphism: heteromorphs have a more obtuse dorsal angle in lateral view, and in dorsal view are wider behind a more pronounced median stricture. Tecnomorphs have a more acute dorsal angle in lateral view and in dorsal view are virtually of equal width throughout, the

median stricture being poorly developed.

Remarks:

This species looks like *Healdianella linevensis* Tschigova, 1958 from the upper Tournaisian of the Saratov-Leningrad area (*Trudy V.N.I.G.R.I.*, 14). *H. linevensis* differs in having a smaller length/height ratio, a more convex anterodorsal border and an anterior border which is not laterally compressed.

H. ? aremorica is assigned to Headianella with doubt because in dorsal view its carapace is laterally compressed in the medial region, a characteristic not observed in other species of that

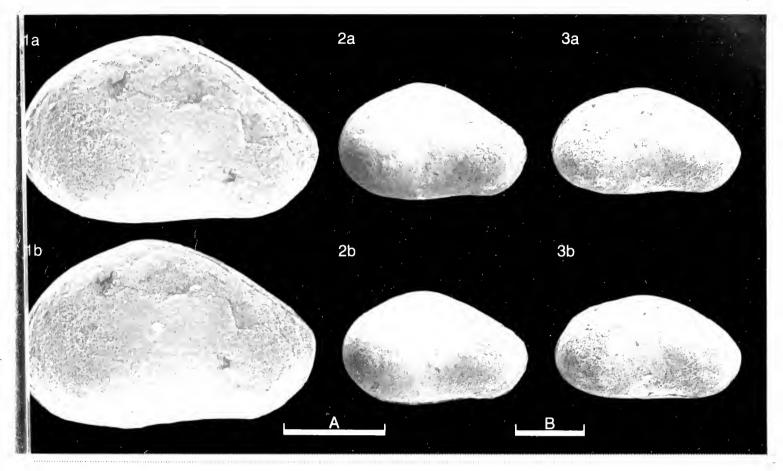
genus.

Distribution:

Laval syncline, Armorican Massif, France: uppermost Tournaisian-lower Visean, lower Carboniferous.

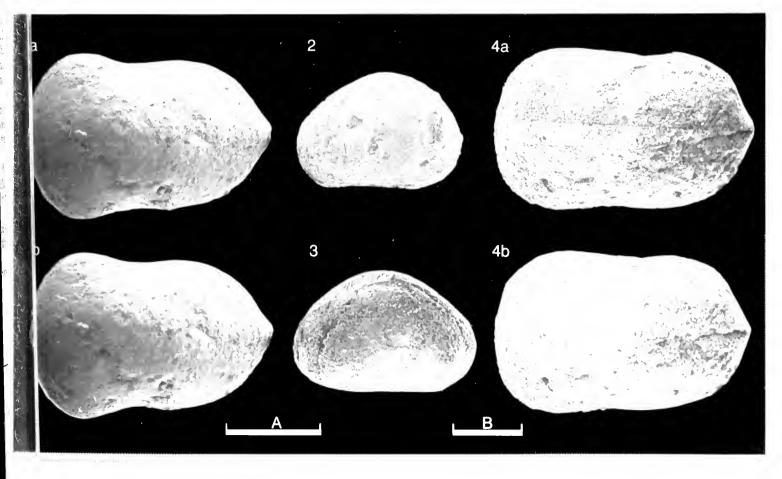
Explanation of Plage 14, 28

Fig. 1, \bigcirc car., dors. (paratype, COUL 865, 0.58 mm); fig. 2, juv. car., lt. lat. (paratype, COUL 861, 0.47 mm long); fig. 3, juv. car., rt. lat. (paratype, COUL 864, 0.40 mm long); fig. 4, \bigcirc car., dors. (paratype, COUL 2155, 0.55 mm long). Scale A (200 μ m; × 125), figs. 1, 3, 4; scale B (200 μ m; × 100), fig. 2.



tereo-Atlas of Ostracod Shells 14, 28

Healdianella? aremorica (4 of 4)



ON MAGHREBEIS TUBERCULATA MAJORAN gen. et sp. nov.

by S. Majoran

(Department of Historical Geology and Palaeontology, University of Uppsala, Sweden)

Genus MAGHREBEIS gen. nov.

Type-species: Maglirebeis tuberculata sp. nov.

Diagnosis:

Derivation of name: From the North African province of Maghreb (including Morocco, Algeria and Tunisia). Carapace small, subtriangular, inequivalved. Left valve larger, overhanging posterior, ventral and anterior margins of right valve. Ventral margin convex, converging posteriorly with straight dorsal margin. Thick, swollen ridge runs along evenly rounded anterior margin, denticulated prominently only on right valve, seldom and only feebly on left valve. Caudal process triangular, pointed at mid-height and armed with a swollen ridge. Ornament polymorphic. Ventromedian and dorsomedian areas bear pronounced lobate tubercles. Lateral surface variously pitted, ventral surface with 4-5 fine longitudinal ribs. Hinge ear of left valve is large, forms a thick, hook-like protuberance that overlaps right valve. Circular eye tubercle and ovate adductor muscle tubercle prominent. Hinge amphidont/heterodont; right valve has large posterior tooth and strong anterior

tooth with a large, spherical distal part fused to a smaller proximal part.

Remarks:

Similar outline to Veenia Butler & Jones, 1957 and Veeniacythereis Gründel, 1973, which differ by being larger, having 3 longitudinal ridges and lacking the curved, left hinge ear of Maghrebeis. The left hinges of all three are similar, but the right hinge of Maglirebeis differs in having a modified anterior tooth. Veenia also differs by its usually more pointed caudal process, and Veeniacythereis by its feeble or absent subcentral tubercle.

Cythereis? sp. of Rosenfeld & Raab (Bull. geol. Surv. Israel, 62, pl. 2, figs. 45-46, 1974; upper Cenomanian, Israel) probably belongs to Maghrebeis since it differs only by its smooth

Explanation of Plate 14, 30

Fig. 1, Q?car., lt. lat. (holotype, PMAL1, 500 μ m long); fig. 2, Q? car., dors., (PMAL2, 530 μ m long); fig. 3, Q? RV, int. lat. (PMAL3, 500μm long); fig. 4, Ω ? LV, int. lat., (PMAL4, 510μm long). Scale A (100μm; ×130), figs. 1-4.

Stereo-Atlas of Ostracod Shells 14, 31

Maghrebeis tuberculata (3 of 4)

Remarks: (cont.)

surface and possibly having one less dorsal tubercle. Also possibly congeneric is Cythereis lindiensis Bate, 1969 as reported by Grosdidier (Revue Inst. fr. Petrole., 28, pl. 13, fig. 104, 1973), which appears to be slightly larger and has more irregular dorsomedian and ventromedian tuberculation, a less pronounced subcentral tubercle and a smooth surface. Further differences are revealed by the original description and re-illustrated type material of C. lindiensis (Bate & Mellish, Stereo-Atlas Ostracod Shells, 13, 59-62, 1986). Another externally similar, considerably larger species is Grosdidier's Cythereis gr. malzi Bischoff, 1963 (Revue Inst. fr. Petrole, 28, pl. 14, fig. 105, 1973).

Maglirebeis tuberculata sp. nov.

Holotype: Type locality: Derivation of name:

Figured specimens:

Palaeontological Museum, University of Uppsala, Sweden, no. PMAL1, ♀? carapace.

Approx. 14km SW of Tocqueville, Algeria (approx. lat. 35° 52'N, long. 4° 55'E); Cenomanian. Latin, from the prominent dorsomedian and ventromedian tubercles.

Palaeontological Museum, University of Uppsala, Sweden, nos. PMAL1 (holotype, ♀? car.: Pl. 14, 30, fig. 1), PMAL2 (\bigcirc ? car.: Pl. 14, 30, fig. 2), PMAL3 (\bigcirc ? RV: Pl. 14, 30, fig. 3), PMAL4 (\bigcirc ? LV: Pl. 14, 30, fig. 4), PMAL5 (♀? car.: Pl. 14, 32, fig. 2), PMAL6 (♀? car.: Pl. 14, 32, fig. 1),

PMAL7 (6? car.: Pl. 14, 32, figs. 3, 4). All from the type locality and horizon.

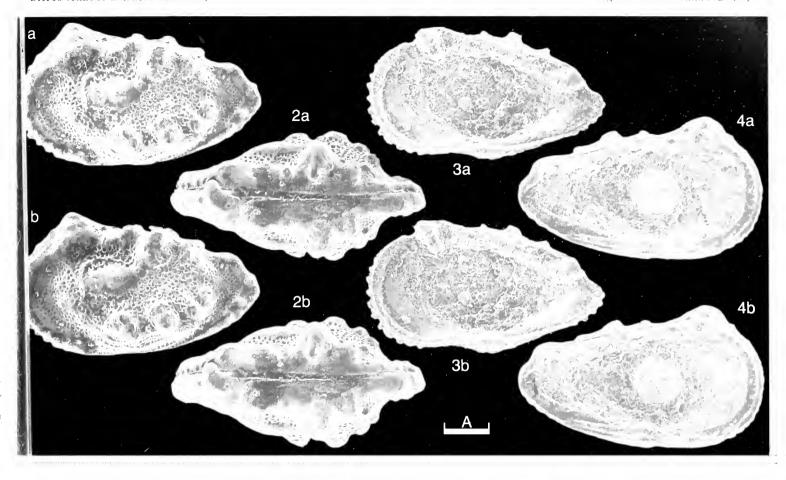
Diagnosis:

Maglirebeis with fine network of small pits, and 3 smooth, lobe-like tubercles respectively on dorsomedian and ventromedian regions. Two additional, smaller tubercles vertically arranged on posteromedian area. Swollen anterior and posterior ridges, pronounced eye tubercle, and ovate, adductor muscle tubercle are all smooth as are also some narrow, longitudinal fields on ventral surface. Ornament polymorphic with respect to size and configuration of dorsomedian and ventromedian tubercles, strength of anterior and posterior ridges, and presence of denticles along ventral section of caudal process. Shape differences might reflect sexual dimorphism: one type being dorsoventrally and laterally more compressed (= \bigcirc ?).

Uppermost Albian (or lower Cenomanian) to middle Cenomanian of N Africa. Distribution:

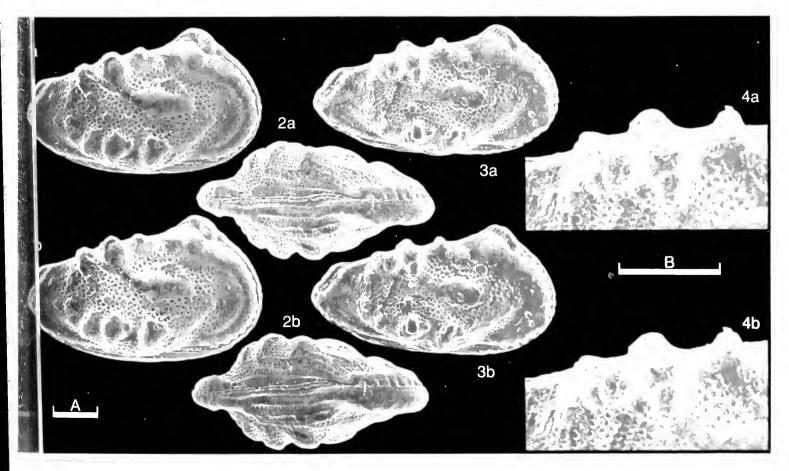
Explanation of Plate 14, 32

Fig. 1, ♀? car., rt. lat. (PMAL 6, 500μm long); fig. 2, ♀? car., vent., showing pitted surface with smooth, narrow, longitudinal fields (PMAL5, 500μm long); figs. 3-4, ♂? car. (compressed morph), (PMAL7, 500μm long): fig. 3, car., rt. lat.; fig. 4, detail of dorsomedian tuberculation. Scale A (100 μ m; ×130), figs. 1-3; scale B (100 μ m; ×280), fig. 4.



Stereo-Atlas of Ostracod Shells 14, 32

Maghrebeis tuberculata (4 of 4)



ON HOWEINA CAMPTOCYTHEROIDEA HANAI

by Noriyuki Ikeya & Ellen Compton-Gooding (Shizuoka University, Shizuoka, Japan & U.S. Geological Survey, Reston, VA)

Genus HOWEINA Hanai, 1957

Type-species (by original designation): Howeina camptocytheroidea Hanai, 1957

Diagnosis: Ovate Cytheruridae, right valve overlapping on dorsal margin, left valve overlapping on ventral margin. Greatest neight anterior, ventral margin nearly straight with a slight alate projection, eye

tubercle indistinct. Inner margin has modified S-shape along posterior margin.

Remarks: Howeing resembles Semicytherura Wagner, 1957 (see Whittaker, Stereo-Atlas Ostracod Shells, 2,

69-92, 1974); some might consider them synonymous since both have S-shaped posterior inner margins, but the validity of this criterion for recognising *Semicytherura* is questionable. In any case, right valves of *Howeina* have a large elongate anterior tooth, a knob-like posterior tooth and a smooth median element; in *Semicytherura* anterior and posterior teeth of the right valve are crenulate or have 2-3 knob-like projections, and the median element is smooth in the center with

sockets at its ends.

Howeina camptocytheroidea Hanai, 1957

1957 Howeina camptocytheroidea sp. nov. T. Hanai, J. Fac. Sci. Tokyo Univ., sec. 2, 11, 22-23, pl. 3, figs. 4a-c, text-figs. 5a, b.

1961 Howeina camptocytheroidea Hanai; T. Hanai, ibid., 13, 358, text-fig. 2, figs. 5a, b.

1971 Howeina camptocytheroidea Hanai; K. Ishizaki, Tohoku Univ. Sci. Rept., 2nd ser., (Geol.), 43, 79-80, pl. 2, fig. 21.

1977 Howeina camptocytheroidea Hanai; T. Hanai et al., Bull. Univ. Mus. Tokyo, 12, 56, pl. 3, figs. 1-7.

Explanation of Plate 14, 34

Fig. 1, \circlearrowleft LV, ext. lat. (IGSU-0-122, 618 μ m long); fig. 2, \circlearrowleft car., ext. dors. (IGSU-0-126, 653 μ m long); fig. 3, \circlearrowleft RV, ext. lat. (IGSU-0-123, 613 μ m long); fig. 4, \updownarrow car., ext. vent. (IGSU-0-121, 605 μ m long); fig. 5, \updownarrow RV, ext. lat. (IGSU-0-124, 625 μ m long). Scale A (100 μ m; × 100), figs. 1-5.

Stereo-Atlas of Ostracod Shells 14, 35

Howeina camptocytheroidea (3 of 4)

Holotype: University Museum, University of Tokyo, Tokyo, Japan, no. UMUT-CA-2612, Q right valve.

[Paratypes: nos. UMUT-CA-2613-2615].

Type locality: Upper Pliocene Setana Formation at Kaigarazawa, about 500m W of Nishinosawa, Kuromatsu-

nai, Suttsu-gun, Hokkaido (lat. 42° 39′ 37″N, long. 140° 17′ 37″E).

Figured specimens: Institute of Geosciences, Shizuoka University (IGSU) nos. 0-121 (\$\Q222\$ car.: Pl. 14, 34, fig. 4), 0-122

(\circlearrowleft LV: Pl. 14, 34, fig. 1), 0-123 (\circlearrowleft RV: Pl. 14, 34, fig. 3; Pl. 14, 36, fig. 7), 0-124 (\Lsh RV: Pl. 14, 34, fig. 5; Pl. 14, 36, figs. 4-6), 0-125 (\Lsh LV: Pl. 14, 36, figs. 1-3), 0-126 (\circlearrowleft car.: Pl. 14, 34, fig. 2). 0-121 is a Recent specimen from Mutsu Bay, northern Honshū (lat. 41° 20'N, long. 140° 55'E).

0-122-126 are from the type locality; 0-122,123 are disarticulated valves of the same individual.

Diagnosis: The valve surface has a pattern of pits that run parallel along the posterior and dorsal margins and somewhat longitudinally in the center of the valve. The anterior and posterior areas have a pattern

of irregular polygons delineated by fine ridges. The flat ventral margin has a series of ridges that run parallel to it. The posteroventral area is slightly depressed behind the alate projection.

Remarks: H. higashimeyaensis Ishizaki, 1971, H. leptocytheroidea (Hanai, 1957), and H. neoleptocytheroidea

(Ishizaki, 1966) each possess a distinctive pattern of prominent ridges and varying degrees of reticulation. They also have caudal processes that are more obvious than that of *H. camptocytheroidea*. Specimens illustrated by McDougall, Brouwers & Smith (*Bull, U.S. Geol. Surv.*, **1598**, 56, pl. 10, figs. 1,2, 1986) from Prudhoe Bay, Alaska, as *Cytherura* sp. B and

Cytherura sp. C., appear to be very similar to H. camptocytheroidea.

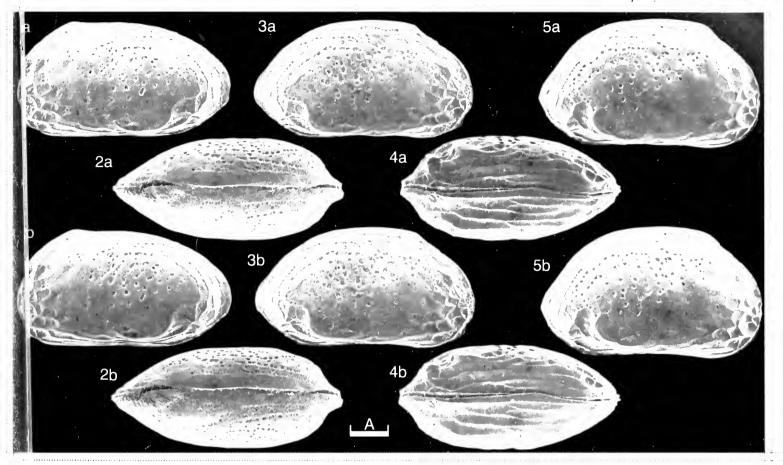
Distribution: A cold water species, H. camptocytheroidea is currently living in Suttsu and Uchiura Bays,

southern Hokkaido; Aomori and Mutsu Bays, Aomori Prefecture; and Otsuchi Bay, Iwate Prefecture. Late Pleistocene occurrences: the Nopporo Fm. Hokkaido; Shibikawa and Anden formations, Akita Prefecture; Hashidate Fm. Ishikawa Prefecture; and Jizodo, Yabu and Kiyokawa formations in Chiba Prefecture. In the late Pliocene, it occurs at the type locality; the Tomikawa Fm. Hokkaido; the Hamada Fm. Aomori Prefecture; and the Junicho Fm. Toyama

Prefecture.

Explanation of Plate 14, 36

Fig. 1-3, \bigcirc LV (IGSU-0-125, 650 μ m long); fig. 1, int. lat.; fig. 2, post. hinge; fig. 3, ant. hinge; figs. 4-6, \bigcirc RV (IGSU-0-124); fig. 4, int. lat., fig. 5, ant. hinge; fig. 6, post. hinge; fig. 7, \bigcirc RV, int. musc. sc., dorsal is to right (IGSU-0-123). Scale A (100 μ m; × 100), figs. 1, 4; scale B (100 μ m; × 160), figs. 2, 3, 5, 6; scale C (10 μ m; × 540), fig. 7.



ON SPINOLEBERIS EXIMIA (BOSQUET)

by J. F. Babinot & J. P. Colin

(Université de Provence, Marseille and Esso Production Research-European Lab., Bègles, France)

Genus SPINOLEBERIS Deroo, 1966

Type-species: Cythere eximia Bosquet, 1854 (by original designation).

Diagnosis:

Small-sized trachyleberidid (less than $650\,\mu\text{m}$) characterized by a well-marked hemispherical, sub-central tubercle; ventral ridge reduced to a strong posterior spinose tubercle and a median lamellar spine; strong spinose tubercle present at the postero-dorsal angle and a vertical spine on the middle part of the dorsal margin. A ridge connects the eye-tubercle and the sub-central tubercle; a weak longitudinal median ridge may be present; anterior margin bordered by two rows of strong spines. Surface of the valves smooth to very finely reticulate mostly on the anterior half. Sexual dimorphism distinct, males being longer than females. Amphidont hinge. Anterior marginal zone of medium width with about 20 straight pore canals. Muscle scars: three small scars disposed in a V-shape or a V-shaped scar with an additional round scar above the posterior branch; four adductor scars, the upper one being divided into two, the one below into three.

Remarks:

The genus *Spinoleberis* is relatively common in the late Cretaceous of Western and Central Europe. Typical species are restricted to the Campanian-Maastrichtian. Cenomanian and Turonian species such as *S. petrocorica* (Damotte, *Rev. Micropal*, **14**, 1, 1973), *S. krejcii* Pokornỳ (*Acta Univ. Carolinae Geol.*, 4, 1968) and *S. ectypus* Babinot (*Géobios*, **6**, 1, 1973) have a rather different morphology; they are deeply reticulate and do not display the characteristic spinose tubercles. Species attributed to the genus *Spinoleberis* by Donze (1970), have been recently placed in the newly errected genus *Navarracythere* Colin & Rodriguez-Lazaro (*Stereo-Atlas Ostracod Shells*, **13**, 63–66, 1986).

Explanation of Plate 14, 38

Fig. 1, $\$ car., ext. rt. lat. (20648–49, 560 μ m long); fig. 2, $\$ LV, ext. lat. (20646–47, 550 μ m long); fig. 3, $\$ RV, ext. lat. (20642–43, 560 μ m long). Scale A (250 μ m; \times 110), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 39

Spinoleberis eximia (3 of 4)

Spinoleberis eximia (Bosquet, 1854)

1854 Cythere eximia n. sp. J. Bosquet, Verhandel. geol. beschr. kaart Nederland, 2, 106, pl. 7. figs. 6a-d.

1936 Cythereis eximia (Bosquet); J. E. van Veen, Nat. hist. Maandbl., 25, 11-12, 26, pl. 7, figs. 1-6.

1958 Cythereis eximia (Bosquet); H. Howe & L. Laurencich, Introduction to the study of Cretaceous Ostracoda, 196-197.

1966 Spinoleberis eximia (Bosquet); G. Deroo, Meded. geol. Sticht., C, 2, 2, 165-166, pl. 6, figs. 72-74, pl. 26, figs. 822-824.

1966 Cythereis eximia (Bosquet); E. Herrig, Paläont. Abl., A, 2, 801-802, pl. 18, figs. 1-10, pl. 19, fig. 1.

1983 Spinoleberis eximia (Bosquet); B. Clarke, Mitt. Geol.-Paläont. Inst. Univ. Hamburg, 54, 110-111, pl. 7, figs. 11-12.

Holotype: Material deposited in the collections of the Institut Royal des Sciences Naturelles de Belgique,

Brussels under the reference "Crétacé Ostracodes 87 Arthr. Sec. I, Cret.", slide no. 44. e locality: Late Maastrichtian of St. Pietersburg, near Maastricht, southern Limburg, the Netherlands.

Type locality: Figured specimens:

These are deposited in the collections of Esso Production Research – European Laboratories at Bègles, France and the numbers all carry the prefix EPR-E. EPR-E 20648-49 (\bigcirc car.: Pl. 14, 38, fig. 1), 20646-47 (\bigcirc LV: Pl. 14, 38, fig. 2), 20642-43 (\bigcirc RV: Pl. 14, 38, fig. 3), 20650-51 (\bigcirc LV: Pl. 14, 40, fig. 1), 20654-55 (\bigcirc car.: Pl. 14, 40, fig. 2), 20816-17 (\bigcirc RV: Pl. 14, 40, fig. 3). All figured specimens are from Puits Maurits (250.5m), Maastricht, southern Limburg, The

Netherlands; late Maastrichtian.

Diagnosis: As for the genus. The surface of the muri of the reticulation is very finely pitted. A little knob

occurs in the middle of each mesh.

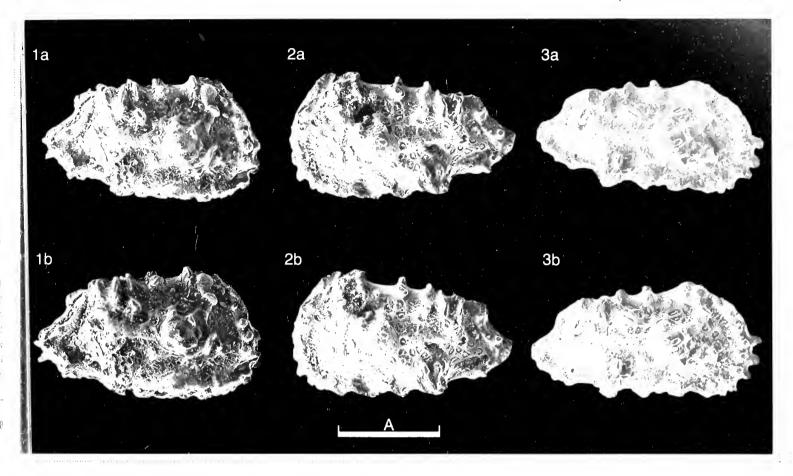
Remarks:

Deroo (1966), illustrated several species of the genus *Spinoleberis* in the type Maastrichtian. Most of the species are very similar to *S. eximia* and therefore extremely difficult to differentiate. Whether they are different species or merely ecotypes is highly questionable. These are *S. eximioides* (van Veen) and *S. pseudoeximia* Deroo. *Cythereis symmetrica* van Veen, 1936 (*Nat. Hist. Maandbl.*, 25, 11–12) is considered by Deroo (1966) and Clarke (1983) to be a juvenile of *S. eximia*, by Howe & Laurencich (1958), (1965) to belong in *S. tuberosa* (Jones & Hinde) and by Szczechura (1965) to belong in *S. spinifera* (van Veen).

Distribution: Late Maastrichtian of the Netherlands and Belgium. Early to late Maastrichtian of Germany.

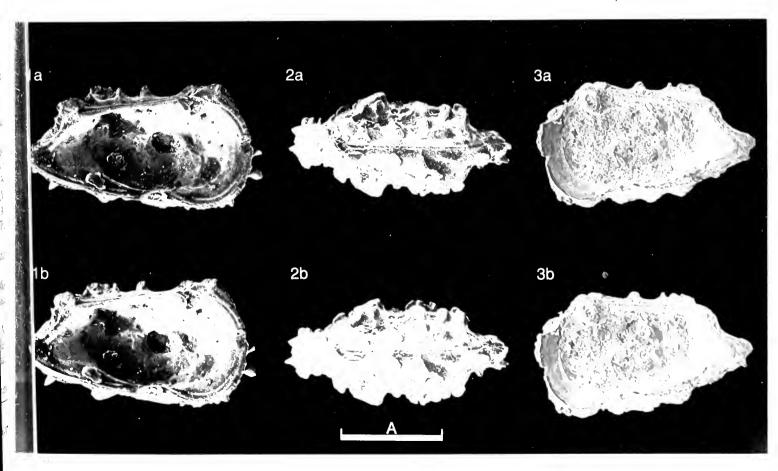
Explanation of Plate 14, 40

Fig. 1, Q LV, int. lat. (20650–51, 575 μ m long); fig. 2, O car., ext. dors. (20654–55, 555 μ m long); fig. 3, Q RV, int. lat. (20816–17, 545 μ m long). Scale A (250 μ m; ×110), figs. 1–3.



Stereo-Atlas of Ostracod Shells 14, 40

Spinoleberis eximia (4 of 4)



ON KOVALEVSKIELLA CAUDATA (LUTZ)

by P. Carbonel, J.-P. Colin & L. Londeix (University of Bordeaux, Talence & Esso Production Research, Bègles, France)

Kovalevskiella caudata (Lutz, 1965)

1965 Gouphocythere caudata sp. nov. A. K. Lutz, Geol. Jahrb., 82, 311, text-fig. 27, pl. 13, figs. 1, 2.

1969 Cordocythere caudata (Lutz); G. Carbonnel & S. Ritzkowski, Arch. Sci. (Genève), 22(1), 60.

1980 Kovalevskiella caudata (Lutz); J.-P. Colin & D. Danielopol, Paleobiol, continent., 11(1), 32, 37, fig. 17.

1985 Kovalevskiella caudata (Lutz); P. Carbonel, Bull. Centres, Recli. Explor,-Prod. Elf-Aquitaine, Mém. 9, pl. 90, figs. 7-10.

1986 Kovalevskiella caudata (Lutz); P. Carbonel, J.-P. Colin, D. L. Danielopol & L. Londeix, Géobios, 19(6), pl. 1, figs. 4-7.

Bundesanstalt für Bodenforschung, Hanover, no. 5421, LV. [Paratype: no. 5420, LV]

Type locality: Road cut between Undorf and Nittendorf near Regensburg. Bayaria, Federal Republic of

Germany; Tortonian, Late Miocene; freshwater molasse.

Figured specimens: Dept. Geol. & Oceanography, Univ. Bordeaux I, CO nos. 5103 (LV: P1. 14, 42, figs. 1-3; Pl. 14,

44, fig. 1), 5104 (RV: Pl. 14, 42, figs. 4-6), 5105 (car.: Pl. 14, 44, figs. 4, 5), 5106 (LV juv.: Pl. 14, 44, fig. 2), 5107 (RV juv.: Pl. 14, 44, fig. 3), 5108 (LV juv.: Pl. 14, 44, fig. 6). Aquitanian, Miocene, of Le Moras, near Labrède, Gironde, France; lat. 44° 41'N, long. 0° 34'W. Original,

German material could not be photographed.

Diagnosis: Carapace subrectangular, rounded anterior and posterior extremities; ornament typical of genus:

regularly disposed pustules. Well developed sulcus; large brood pouch, RV larger than LV; both cardinal hinge elements on LV trilobate. No sexual dimorphism. Two strong denticles on the

posteroventral part of LV.

Explanation of Plate 14, 42

Figs. 1-3, LV (CO 5103, 430μm long); fig. 1, ext. lat.; fig. 2, int. lat. hinge; fig. 3, int. lat. Figs. 4-6, RV (CO 5104, 433μm long); fig. 4, int. lat.; fig. 5, ext. lat.; fig. 6, int. lat. hinge. Scale A (200 μ m; ×135), figs. 1, 3-5; scale B (200 μ m; ×205), figs. 2, 6.

Stereo-Atlas of Ostracod Shells 14, 43

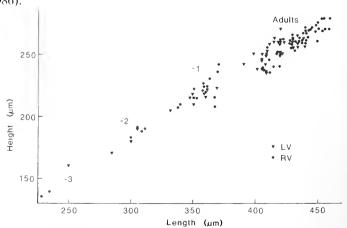
Kovalevskiella caudata (3 of 4)

Like other Kovalevskiella, K. caudata is parthenogenetic. It differs by the presence of 2 strong denticles on the posteroventral part of the left valve. Denticles are also present in larval stages, but only in the right valve. In the French locality studied, this species lived in a lagoonal to lacustrine environment (in oligo-to mesohaline waters) on very fine grained, marl bottom sediment. K. caudata is associated with Neocyprideis aguitanica Moyes or with Cundouopsis and Liuutocythere, and always with poorly diversified faunas. When the waters become fresh and more stable, Kovalevskiella disappears. Lutz (1965) described K. caudata from coaly marls in a freshwater molasse deposit. Its epibenthic life-style is very different to the hypogean or interstitial habitats of Recent Kovalevskiella (Colin & Danielopol 1980; Carbonel et al., 1986).

Distribution:

Miocene (Tortonian) near Regensburg, Germany (Lutz 1965); Miocene (Aquitanian) near Bordeaux, France (Carbonel 1985; Carbonel et al. 1986).

Text-fig. 1. Size dispersion of 116 left and right valves of K. caudata from Le Moras, near Labrède, Gironde, SW France.



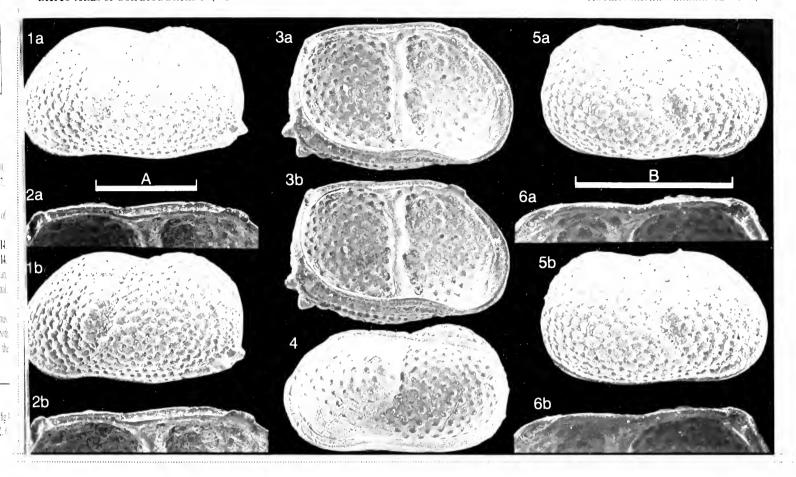
Explanation of Plate 14, 44

Fig. 1, LV ext. dors. (CO 5103, 430μm long); fig. 2, LV juv.-1, ext. lat. (CO 5106, 340μm long); fig. 3, RV juv.-2, ext. lat. (CO 5107) 285μm long). Figs. 4, 5, car. (CO 5105, 420μm long): fig. 4, ext. dors.; fig. 5, ext. vent. Fig. 6, LV juv.-3, ext. lat. (CO 5108, 215μm long). Scale A (200 μ m; ×135), figs. 1-6.

Stereo-Atlas of Ostracod Shells 14, 44

l.Frus.

Kovalevskiella candata (4 of 4)



4a 5a

a 3a

6a

4b

5b

6b

A

 $595.339.1 \ (113.333) \ (64 : 162.008.31) : 551.35 + 552.52$

ON CALOCARIA MAURAE VANNIER gen. et sp. nov.

by Jean Vannier (University of Rennes, France)

Genus CALOCARIA gen. nov.

Type-species: Calocaria maurae sp. nov.

Derivation of name: From the Greek karia, a walnut and kalos, beautiful; alluding to the shell shape and lateral

ornament. Gender feminine.

Diagnosis:

Myodocopid ostracode, oval in outline. Length 3.5mm; adults have a length:height ratio of approximately 1.1. Prominent anterior rostrum and rostral incisure. Composite external ornament: pattern of coarse ridges running obliquely to the ventral and dorsal margins, converging towards the middle of the valve and connected ventrally to a continuous marginal ridge; posteriorly to mid-length, ornament consists of a linear alignment of more and less coalescent

tubercles laterally merging with corrugations. Small arcuate muscle scar impression.

Remarks:

This new genus shares many similarities (rostrum, simple muscle scar impression, oval outline) with other Silurian 'cypridinids' and is tentatively included within this group. Nevertheless, the composite ornament of Calocaria is comparable to that of Silurian 'bolbozoids' (see Siveter, Vannier & Palmer, Palaeontology, text-fig. 4, in press, 1987) but neither the distinctive anterodorsal bulb nor sulcus typical of that group occur in Calocaria. By its well developed ornament Calocaria is distinguished from any other Silurian 'cypridinids', which are mainly smooth (see Siveter, Vannier & Palmer, Palaeontology, pls. 2, 3, 5, in press, 1987). As with numerous myodocopids from the Silurian of Europe, most specimens of Calocaria maurae show 'plastic' deformation of the valves, suggesting a rather thin, flimsy shell.

Explanation of Plate 14, 46

Figs. 1-3, RV (holotype, IGR 33100, 3055μm long): fig. 1, ext. lat.; fig. 2, ext. vent. obl.; fig. 3, ornament of lateral surface. Scale A (750 μ m; ×18), figs. 1, 2; scale B (100 μ m; ×75), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 47

Calocaria maurae (3 of 4)

Calocaria maurae sp. nov.

Institut de Géologie, University of Rennes (IGR), France, coll. no. 33100; RV. Holotype:

[Paratypes: IGR coll. nos. 33101, LV; 33103, RV; 33104, LV; 33106, LV].

Siltstones and mudstones in the Talmakent section near Talmakent (sample TA 84452 of J. J. Type-locality: Cornée collections, University of Aix-Marseille, France), Haut-Atlas, Morocco; lat. 31° 52′N,

long. 7° 45'W. Upper part of the Silurian; as determined by J. J. Cornée (work in progress).

From *maura*, Moorish, alluding to the region where this species occurs. Derivation of name:

Inst. de Géologie, Univ. Rennes (IGR), coll. nos. 33100 (holotype, RV: Pl. 14, 46, figs. 1-3; Pl. 14, Figured specimens:

48, figs. 1, 2) and 33101 (paratype, LV: Pl. 14, 48, figs. 3-6). From type locality; latex casts.

Diagnosis: As for the genus. Monotypic. Remarks:

C. maurae is the first Silurian myodocopid to be described from Africa. In the type locality, it is associated with numerous other myodocopid ostracodes, both 'bolbozoids' and 'cypridinids'. In this respect, this fauna is comparable to that recently documented from organic-rich, Silurian sediments of Britain and France (Siveter, et al., op. cit., in press). As a prelude to their further systematic studies of Silurian myodocopids the latter authors have noted the occurrence of similar myodocopid faunas in the same type of deposits (black to terrigenous mudstones) from northwestern Europe (e.g. Ludlow Series in Wales and the Armorican Massif, France), eastern Europe (Ludlow Series of Bohemia, Czechoslovakia) and now North Africa (herein). An outer shelf to shelf margin or even shelf slope environment is inferred, from faunal and sedimentological evidence, for the myodocopid occurrences in Europe (see Siveter, et al., op. cit., in press) and is

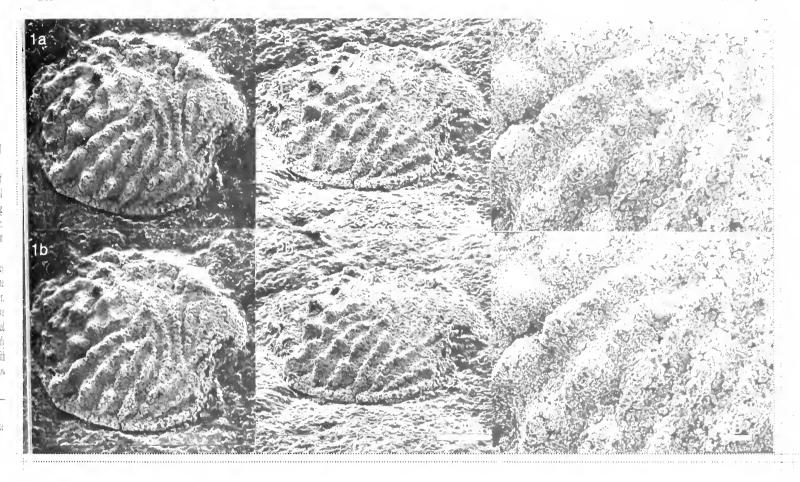
also likely in the case of the Moroccan material. At present, known only from the type locality.

Distribution: To J. J. Cornée (University of Aix-Marseille) for allowing me to study his material and to the Acknowledgements:

Humboldt Foundation (Bonn) for my Research Fellowship at Hamburg University.

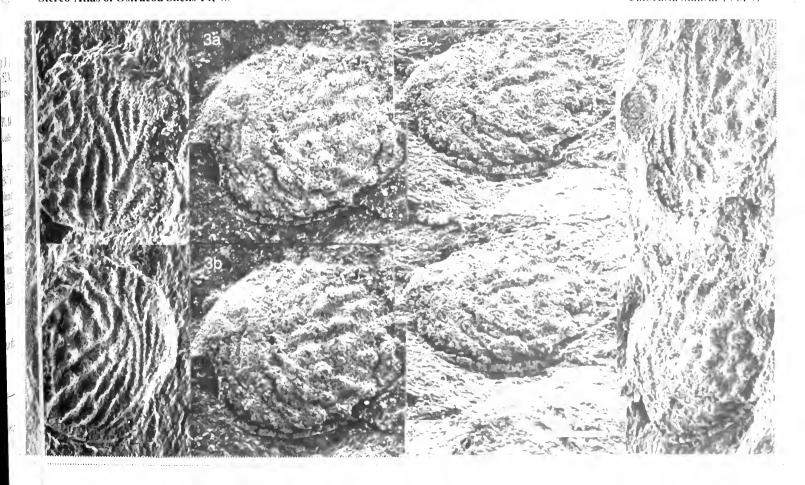
Explanation of Plate 14, 48

Figs. 1, 2, RV (holotype, IGR 33100, 3055μm long): fig. 1, ext. ant. obl.; fig. 2, ext. post. obl. Figs. 3-6, LV (IGR 33101, 3050μm long): fig. 3, ext. lat.; fig. 4, ext. vent. obl.; fig. 5, ext. ant. obl.; fig. 6, ext. post. obl. Scale A ($750\mu m$; ×18), figs. 1-6.



Stereo-Atlas of Ostracod Shells 14, 48

Calocaria maurae (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (13) 49–56 (1987) 595.336.13 (113.312) (437 : 161.013.49) : 551.35 + 552.52

ON SPINOHIPPULA ESURIALIS VANNIER, KRŮTA & MAREK gen. et sp.

by Jean Vannier, Miroslav Kruta & Ladislav Marek (University of Rennes, France; Academy of Sciences, Prague, Czechoslovakia)

Genus SPINOHIPPULA gen. nov.

Type-species: Spinohippula esurialis sp. nov.

Derivation of name:

Alluding to velar spines and ressemblance with species of the tribe Hippulini. Gender feminine.

Diagnosis:

Medium sized glossomorphitine (adults < 1.2mm long). Lateral surface virtually lacks lobation: sulci only expressed dorsally as two very poorly marked depressions, presumably representing S2 and S3. Strong velum extending from near anterior cardinal corner to posteromedium or posterodorsal part of valve and bearing a distinctive coarse denticulation along inner margin bordering a deep (maximum depth ventrally and anteriorly) fissum-like laterovelar furrow. Dimorphism mainly expressed in females by a broad crescent-like velar flange and a wide concave subvelar area (dolonal antrum) both ends of which connect with lateral surface of valve. Inner velar spines converging towards middle of domicilium tend to reach the lateral surface over the laterovelar furrow. Tecnomorphs have narrower velum reduced to a row of radiating velar spines,

with laterovelar furrow more open than in females; marginal sculpture unknown.

Remarks:

The inclusion of *Spinohippula* within the Glossomorphitinae (see R. Schallreuter, *Palaeontographica A*, **180**, 1983) is justified by the occurrence of a strong velar sculpture in both females and tecnomorphs, and well marked velar dimorphism. The shape of the velar flange in females is its most significant glossomorphitine feature, consisting of a massive adventral sculpture (Pl. **14**, 52, fig. 2) high the domicilium. Comparable features are in typical glossomorphitines such as *Collibolbina collis collis* (Schallreuter, 1964) (see R.

Explanation of Plate 14, 50

Figs. 1-4, Q LV (holotype, NM L26073, 1188 μ m long): fig. 1, ext. lat.; fig. 2, ext. vent. obl. (tilted 75°); fig. 3, ext. dors. obl. (tilted 45°), antero-vent. part of the valve. Scale A (300 μ m; ×69), figs. 1-3; scale B (200 μ m; ×115), fig. 4.

Stereo-Atlas of Ostracod Shells 14, 51

Spinohippula esurialis (3 of 8)

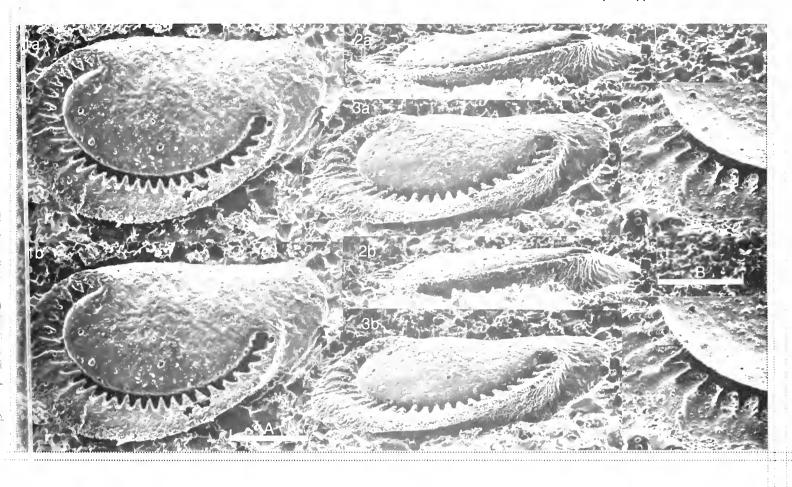
Remarks: (cont.)

Schallreuter, op. cit., 1983) from the middle Ordovician of Baltoscandia or *Gracquina hispanica* (Born, 1918) (see J. Vannier, *Palaeontographica* A, **193**, 1986) from the Llandeilo of France and Spain.

Schallreuter (op. cit., 1983) divided the subfamily Glossomorphitinae into three tribes on the basis of different types of velar dimorphism. Spinohippula shares strong similarities with representatives of the tribe Hippulini, especially Hippula and Parahippula. Although no typical torus is observed (in addition to the velar flange) in Spinohippula, the general morphology of its adventral region is strikingly comparable to that of Hippula (subgenera H. (Hippula) and H. (Cetona)) and Parahippula. Schematically, the shape (crescent-like sculpture), position and nature (main adventral sculpture with hollow spaces and/or radiating structures) of the velum (Text-fig. 2) are three important characteristics similar in Spinolippula from the middle Ordovician of Czechoslovakia, Parahippula from the middle Ordovician of United States, and numerous species of Hippula from the Ordovician of Europe and North America (see R. Schallreuter & M. Kruta, N, Jb. Paläont. (Mh), 8, 1980). In Spinohippula the row of velar denticulation (26 spines in holotype) (Pl. 14, 50, fig. 4; Pl. 14, 52, fig. 4) associated with a deep laterovelar furrow forms a semi-open peripheral groove (Text-fig. la) at the junction of the lateral, marginal and velar surfaces. By comparison, H. (Cetona) (Text-fig. 1b), an example of a typical unitoral hippuline, exhibits a row of short, flattened tubule-like spaces (= part of torus) connected to the velum, exactly at the same place as the semi-open groove in Spinohippula. Moreover, openings of these hollows (13 in females of H. (C.) cetona cetona = half the number of velar spines in the holotype of S. esurialis) are also impressed on the velar flange surface (cf. Text-figs. 1a & 1b). Similar comparisons could also be attempted with H. (Hippula) characterized by two tori (Text-fig. 2d). The velar flange of Parahippula (Text-fig. 1c) is considered by Kraft (Mem. geol. Soc. Am, 86, 1962) as a "hollow velate frill formed of two layers continuous with the outer layer of shell wall". The interpretation of this "hollow structure" is problematic, bearing in mind that two-layered structures frequently observed on silicified specimens (see D. J. Siveter, Stereo-Atlas Ostracod Shells, 12 (10), pl. 54, fig. 4) may be the result of diagenetic processes. Nevertheless, as stated by Kraft, "the logical structure of the carapace wall" of Parahippula is a convincing argument for true hollow spaces within the velum (as reconstructed in Text-fig.

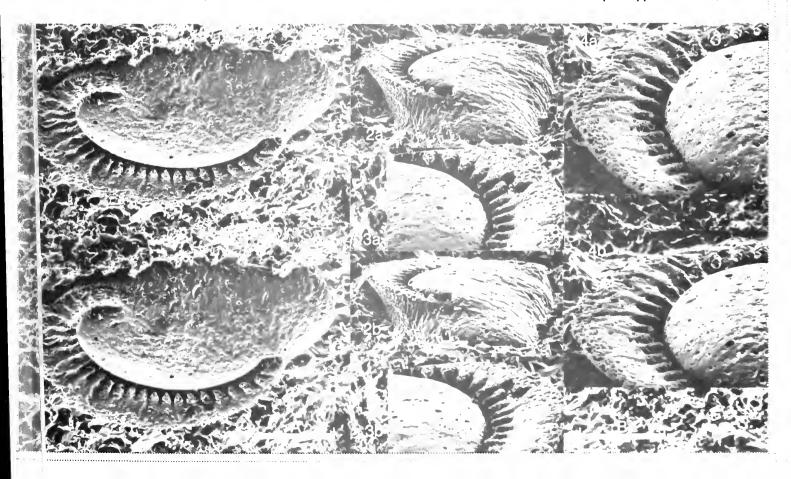
Explanation of Plate 14, 52

Figs. 1-4, \mathcal{Q} LV (holotype, NM L26073, 1188 μ m long): fig. 1, ext. dors. obl. (tilted 45°); fig. 2, ext. post. obl. (tilted 75°); fig. 3, ext. ant. obl. (tilted 75°), postero-vent. part of the valve; fig. 4, ext. post. obl. (tilted 55°). antero-vent. part of the valve. Scale A (300 μ m; ×69), figs. 1, 2; scale B (200 μ m; ×115), figs. 3, 4.

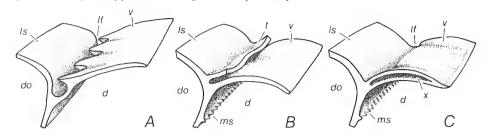


Stereo-Atlas of Ostracod Shells 14, 52

Spinohippula esurialis (4 of 8)



Remarks: (cont.) 1c). In this case: 1) the hollow spaces (13 in females; Text-fig. 2c) within the velar flange of Parahippula, and 2) its deep laterovelar furrow showing 12 secondary radiating tiny furrows on the velar flange, may represent homologous structures of the 13 toral hollows of H. (Cetona) (Text-fig. 2b) and the semi-open laterovelar groove of Spinohippula (Text-fig. 2a) respectively.



Text-fig. 1. Reconstructions of the adventral sculpture in three genera of the tribe Hippulini Schallreuter, 1983: A, *Spinolippula* gen. nov.; B, *Hippula*; C, *Parahippula*. All views represent medioventral cross-sections of valves (see Text-fig. 2). d = dolonal antrum; do = domicilium; $footnote{if} footnote{if} footnote$

Spinohippula esurialis sp. nov.

Holotype: National Museum, Prague (NM), Czechoslovakia, coll. no. L26073; ♀ LV.

[Paratypes: NM, Prague, coll. nos. L26074, tecnomorph RV; L26075, Q LV]. Casts of the holotype

and paratypes are in the Institute of Geology, University of Rennes, France.

Type locality: Ejpovice (borehole), 10km E of Plzen, WSW of Prague, Bohemia, Czechoslovakia; approx. lat. 49° 47'N,

long. 13° 38'E. Sandstones, Skalka quartzite Dobrotiva (Llandeilo ?) 'series', Ordovician.

Derivation of name: Latin, esurialis, hungry; referring to the teeth-like velar spines.

Explanation of Plate 14, 54

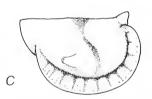
Figs. 1-3, tecnomorph RV (paratype, NM L26074, 838 μ m long): fig. 1, ext. lat.; fig. 2, ext. vent. obl. (tilted 45°); fig. 3, ext. post. obl., part of the valve. Scale A (300 μ m; ×93), figs. 1, 2; scale B (200 μ m; ×115), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 55



Remarks:







Text-fig. 2. Three genera of the tribe Hippulini Schallreuter, 1983. All lateral views of female right valves. A, *Spinohippula esurialis* gen. et sp. nov., from the Dobrotiva series (Llandeilo?) of Czechoslovakia, approximately ×95. B, *Hippula (Cetona) cetona cetona* (Schallreuter, 1964), from Backsteinkalk erratic boulders of northern Germany, middle Ordovician, approximately ×80 (after Schallreuter 1983, *op. cit.* pl. 3, fig. 1). C, *Parahippula ventrospina* (Kraft, 1962), from the middle Ordovician of Virginia, United States, approximately ×65 (after Kraft 1962, *op. cit.*, pl. 12, fig. 5). D, *Hippula (Hippula) latonoda* (Schallreuter, 1964), from the Upper Viru series (Caradoc) of Baltoscandia, approximately ×100 (after Schallreuter 1983, *op. cit.*, pl. 5, fig. 1).

Figured specimens: National Museum, Prague (NM), Czechoslovakia, coll. nos. L26073 (holotype, ♀ LV: Pl. 14, 50, figs. 1-4; Pl. 14, 52, figs. 1-4), L26074 (paratype, tecnomorph RV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♀ LV: Pl. 14, 54, figs. 1-3), L26075 (paratype, ♠ LV: Pl. 14, figs. 1-3), L26075 (parat

56, figs. 1-3). Silicone rubber casts of topotype specimens.

Diagnosis: As for the genus. Monotypic.

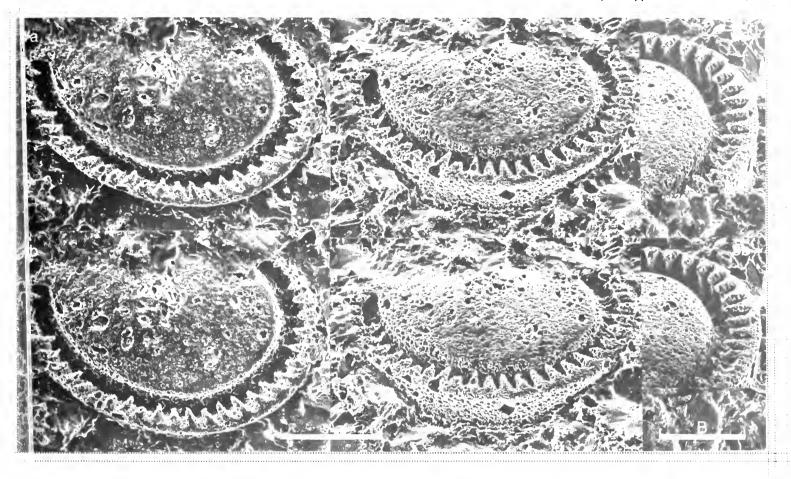
The laterovelar furrow represents a major concavity, widely extended ventrally, but not connected to the domiciliar cavity and protected from the outside by a row of spines; it could be interpreted as an external botulus-like brood concavity. More likely, this groove is homologous to a cavum (see R. Schallreuter in R. Maddocks (Ed.), *Proc. 8th Int. Symp. on Ostracoda, Houston, Texas*, 1983) or a fissum. Some tvaerenellids exhibit an arcuate cavum which tends to be closed by spines in the same way as the laterovelar furrow of *S. esurialis. Huckea luckea* (see R. Schallreuter, *Palaeontographica A*, 149, pl. 9, figs. 1-4, 1975) shows a ventral fissum positioned similarly to the furrow in *S. esurialis*. The exact function of the cavum (buoyancy control?) or fissum is still unknown but might represent an attempt to lighten the shell.

Distribution: At present known only from type locality.

Acknowledgements: To the Humboldt Foundation (Bonn) for my Research Fellowship at Hamburg University.

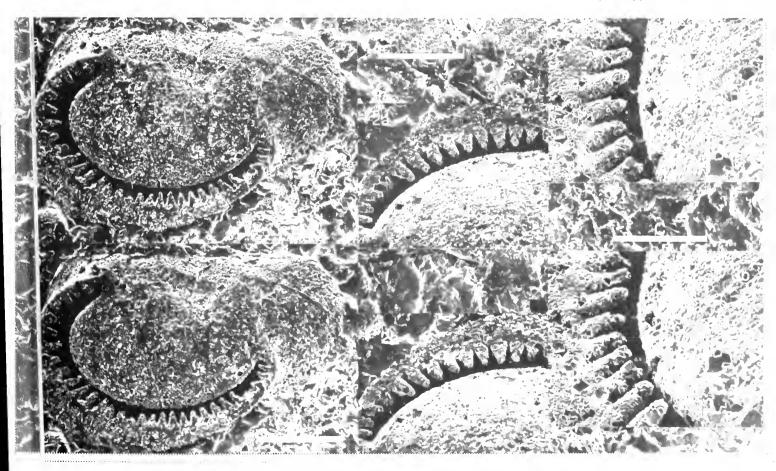
Explanation of Plate 14, 56

Figs. 1-3, Q LV (paratype, NM L26075, 1107 μ m long): fig. 1, ext. lat.; fig. 2, ext. dors. obl., postero-vent. part; fig. 3, ext. post. obl., medio-vent. part. Scale A (300 μ m; ×76), fig. 1; scale B (200 μ m; ×115), fig. 2; scale C (100 μ m; ×250), fig. 3.



tereo-Atlas of Ostracod Shells 14, 56

Spinohippula esurialis (8 of 8)



Stereo-Atlas of Ostracod Shells 14 (14) 57-64 (1987) 595.336.11 (113.331) (481 : 161.010.59) 551.35 + 552.54

ON BEYRICHIA (SAGENABEYRICHIA) SIVETERI POLLICOT subgen. et sp. nov.

by Paul D. Pollicott (University of Leicester, England)

Subgenus BEYRICHIA (SAGENABEYRICHIA) subgen. nov.

Type-species: Beyrichia (Sagenabeyrichia) siveteri sp. nov.

Derivation of name: Diagnosis:

Latin sagena, fish-net; alluding to the reticulate ornament of the lobes + the genus Beyrichia. Beyrichia with reticulo-tuberculate lobal ornament. Crumina elongate and relatively well assimilated with lobal area. Syllobium weakly cuspidate; anterior cusp slightly more prominant, posterior often lacking. Syllobial groove low and often above a well developed callus. Zygal arch

lacking.

Remarks:

The lobal reticulation of *B.* (Sagenabeyrichia) is unique within Beyrichia. Moreover, the occurrence of reticulation in an otherwise typical beyrichiine species has significance for beyrichiacean phyllogeny, particularly in the relationship between amphitoxotidines and beyrichiines. Henningsmoen (Geol. Fören. Stockh. Förli., 86, 387-9, 1965) thought that amphitoxotidines, with their typically tubulose velar frill, evolved from beyrichiines (Beyrichia subgenera). In contrast, Martinsson (Bull. geol. Instn. Univ. Uppsala, 42, 56, 1963) thought that a stabilized surface reticulation within the amphitoxotidines was entirely foreign to typical beyrichiines (although reticulation is known in atypical Beyrichiidae such as Bingeria: see A. Martinsson Bull. geol. Instn. Univ. Uppsala., 41, 1962), a subfamily which, furthermore, he

Explanation of Plate 14, 58

Fig. 1-3, ♂ RV (PMO 116.231, 2.00 mm long): fig. 1, ext. lat.; fig. 2, ext. vent.; fig. 3, ext. vent. obl. Figs. 4, 5, ♂ LV (PMO 116.232, 1.32 mm long): fig. 4, ext. dors. obl.; fig. 5, ext. lat. Scale A (370 μm; × 28), figs. 1-3; scale B (260 μm; × 40), figs. 4, 5.

Stereo-Atlas of Ostracod Shells 14, 59

Beyrichia siveteri (3 of 8)

Remarks: (cont.)

considered more 'advanced' by lacking a tubulose velum. Henningsmoen (*op. cit.*) thought that reticulation in the beyrichiines was an undeveloped possibility and, if found, its occurrence would indicate a possible beyrichiine derivation for the amphitoxotodines. The lobal reticulation of *Beyrichia* (*Sagenabeyrichia*) supports his idea.

B. (Sagenabeyrichia) further differs from many typical B. (Beyrichia) species by its better assimilated crumina, a feature which it has in common with species of B. (Simplicibeyrichia), especially B. (S.) callifera and B. (S.) duplicicalcarata (both Martinsson op. cit., 1962). B. (Sagenabeyrichia) differs markedly from B. (S.) globifera Martinsson, 1962 by its reticulation, lack of a calcarine spine, its often well developed syllobial groove/callus and in having a long, better defined preadductorial sulcus.

Beyrichia (Sagenabeyrichia) siveteri sp. nov.

1954 Beyrichia (Beyrichia) cf. kloedeni McCoy 1846; G. Henningsmoen, Norsk. geol. Tidsskr., 34, 40-43 (pars), pl. 2, fig. 7, 10-18, pl. 3, figs. 2-7; ? pl. 2, fig. 9.

Holotype: Paleontologisk Museum, Oslo, Norway, PMO 116.233; ♀ RV (broken posteriorly).

Type locality: Coastal section, southern tip of Kommersøya (east side), Holmestrand, Norway. Steinsfjorden Formation, '9cβ' of Kiaer (Skr. Vidensk. Selsk. Kristiania I Mat. – Naturv. Kl. 1906 II, 596 pp.);

Wenlock Series, Silurian. Approx. lat. 59° 32'N, long. 10° 18'E.

Derivation of name: Figured specimens:

After Dr. David J. Siveter, University of Leicester, England.

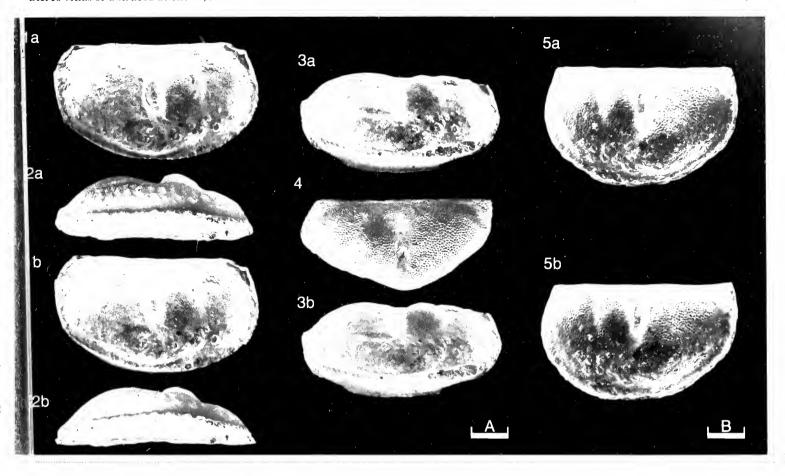
Paleontologisk Museum, Oslo, nos. **PMO 116.231** (\$\sigma\$ RV: Pl. 14, 58, figs. 1, 2, 3), **PMO 116.232** (\$\sigma\$ LV: Pl. 14, 58, figs. 4, 5, Pl. 14, 62, fig. 1), **PMO 116.233** (\$\sigma\$ RV: Pl. 14, 60, fig 1) **PMO 116.235** (\$\sigma\$ LV: Pl. 14, 60, figs. 2, 3), **PMO 116.235** (\$\sigma\$ LV: Pl. 14, 60, fig. 4, 5), **PMO 116.236** (\$\sigma\$ LV: Pl. 14, 62, fig. 4)

(\$\frac{14}{14}\$, 60, figs. 2, 3), PMO 116.235 (\$\frac{1}{2}\$ LV: Pl. 14, 60, fig. 4, 3), PMO 116.236 (\$\frac{1}{2}\$ LV: Pl. 14, 62, fig. 3), PMO 116.238 (\$\frac{1}{2}\$ LV: Pl. 14, 62, fig. 4), PMO 116.239 (\$\frac{1}{2}\$ LV: Pl. 14, 64, fig. 1), PMO 116.240 (\$\frac{1}{2}\$ LV: Pl. 14, 64, fig. 2), PMO 116.241 (\$\frac{1}{2}\$

RV: Pl. 14, 64, fig. 3).

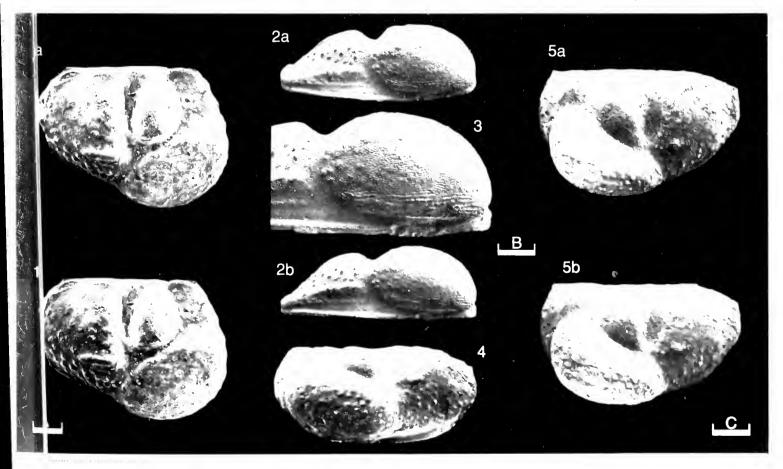
Explanation of Plate 14, 60

Fig. 1, \bigcirc RV, ext. lat. (holotype **PMO 116.233**, 2.00 mm long). Figs. 2, 3, \bigcirc RV (**PMO 116.234**, 2.50 mm long): fig. 2, ext. vent.; fig. 3, ext. vent. detail of crumina. Figs. 4, 5, \bigcirc LV (**PMO 116.235**, 2.48 mm long): fig. 4, ext. vent. obl.; fig. 5, ext. lat. Scale A (390 μ m; × 25), fig. 1; scale B (286 μ m; × 35), fig. 3; scale C (470 μ m; × 22), figs. 2, 4, 5.



tereo-Atlas of Ostracod Shells 14, 60

Beyrichia siveteri (4 ot 8)



Figured specimens: (cont.)

All figured specimens are from the Steinsfjorden Formation (9c), Sjorvoll, Ringerike, except for PMO 116.233 (holotype) and PMO 116.238, which are from the type horizon and locality. All specimens are prepared by mechanical preparation techniques from limestone slabs.

Diagnosis: Remarks:

As for the subgenus. B. (Sagenabeyrichia) is monotypic.

B. (S.) siveteri exhibits wide variation in both lobal reticulation and tuberculation. Most valves are reticulate over the entire lobal area, but in a few specimens, reticulation is lacking on the anterior lobe (possibly a feature of preservation?). Tuberculation varies from forms with extensive cover (mostly adults) to those in which it is lacking (small tecnomorphs). Reticulation is relatively smaller in larger forms, and tubercles are commonly restricted to a supra-velar field (Pl. 14, 58, figs. 1, 5).

Size variation of female adults is common within a single sample (see Text-fig. 1). This is thought to reflect mixed populations (chronodemes and/or ecodemes) rather than a possible case of precocious dimorphism (unknown in Beyrichiacea).

Distribution:

The Wenlock Series, Silurian of Norway. Collected from localities in the Steinsfjorden Formation (see Worsley, D. (ed.), *Nor. geol. unders. 384*, 1982) at Ringerike (9b-9e of Kiaer, *op. cit.*) and Holmestrand (9b-9c of Kiaer, *op. cit.*).

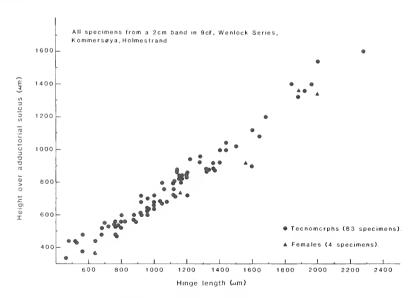
Explanation of Plate 14, 62

Fig. 1, ♂ LV, reticulation on syllobium (PMO 116.232, 1.32 mm long). Figs. 2, 5, ♂ RV (PMO 116.236, 2.32 mm long): fig. 2, ext. lat.; fig. 5, reticulation and tuberculation on syllobium. Fig. 3, ♂ LV, ext. lat. (PMO 116.237, 2.48 mm long). Fig. 4, ♂ LV, ext. lat. (PMO 116.238, 1.20 mm long).

Scale A (21 μ m; × 460), fig. 1; scale B (455 μ m; × 23), figs. 2, 3; scale C (230 μ m; × 40), fig. 4; scale D (62 μ m; × 150), fig. 5.

Stereo-Atlas of Ostracod Shells 14, 63

Beyrichia siveteri (7 of 8)

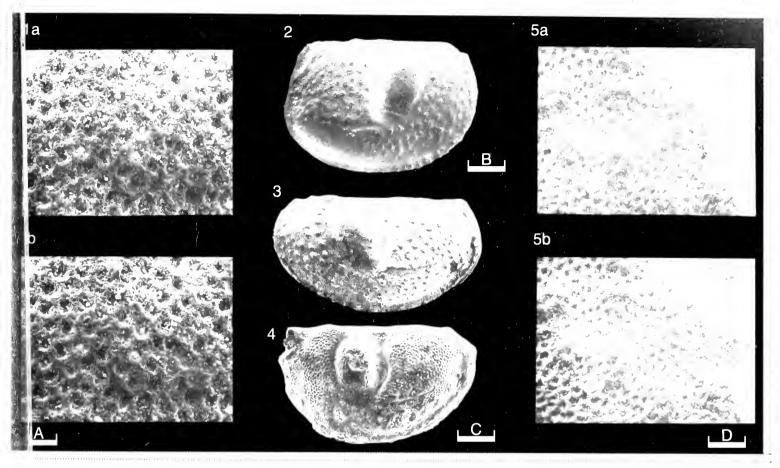


Text-fig. 1. Size variation within *B.* (*Sagenabeyrichia*) siveteri from the Steinsfjorden Formation (9c of Kiaer op. cit.), Wenlock Series at Kommersøya, Holmestrand, Norway.

Explanation of Plate 14, 64

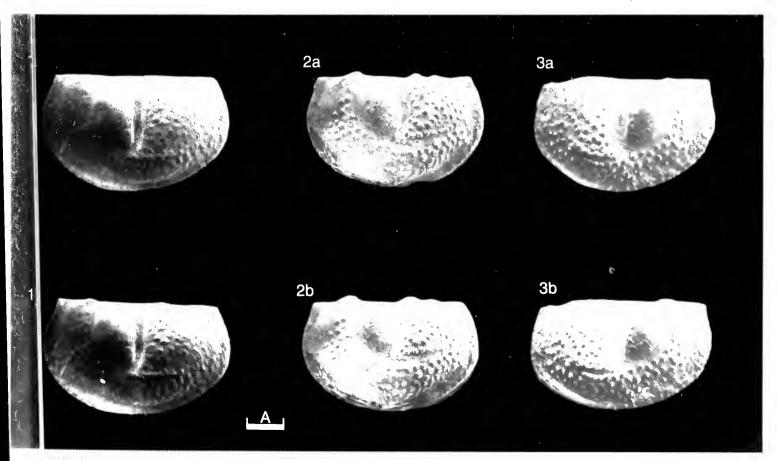
Fig. 1, O LV, ext. lat. (PMO 116.239, 2.72 mm. long). Fig. 2, O LV, ext. lat. (PMO 116.240, 2.68 mm. long). Fig. 3 O RV, ext. lat. (PMO 116.241, 2.68 mm. long).

Scale A (545 μ m; × 18), figs. 1-3.



ereo-Atlas of Ostracod Shells 14, 64

Beyrichia siveteri (8 of 8)



ON BYTHOCYTHERE INTERMEDIA ELOFSON

by David J. Horne (Geology Department, City of London Polytechnic)

Bythocythere intermedia Elofson, 1938

- 1868 Bythocythere constricta Sars; G. S. Brady, Trans. Linn. Soc. Lond., 26, (pars), 451–452, pl. 35, figs. 48–52 only (non pl. 35, fig. 47) (non Sars, 1866).
- 1938 Bythocythere intermedia sp. nov. O. Elofson, Ark. Zool., 30A, 10, text-figs. 14-21.
- Bythocythere intermedia Elofson; J. Athersuch, D. J. Horne & J. E. Whittaker, J. micropalaeontol., 2, 72–73, text-figs. 1, 2, 3a–g, 4r–t, 5b; pl. 2, figs. 1–4.
 - Type specimens: The whereabouts of Elofson's type material is not known.
 - Type locality: The Mittskaren, outside the mouth of Gullmar Fjord, W. Sweden, approx. lat. 58°15′ N, long.
 - 11°30′ E; Recent, marine, sublittoral.
- Figured specimens: British Museum (Nat. Hist.) nos. 1982.345 (O'LV: Pl. 14, 68, fig. 1; copulatory appendage:
 - Text-fig. 1), 1982.346 (♀ LV: Pl. 14, 66, fig. 2; RV: Pl. 14, 66, fig. 3), 1982.347 (♂ LV: Pl. 14, 68, figs. 2, 3), 1982.348 (♂ LV: Pl. 14, 66, fig. 1). All from Valentia, SW Ireland (approx. lat. 51°55′N, long. 10°20′W), taken from slides labelled "B. constricta" in the Norman Collection at the British Museum (Nat. Hist.); nos. 1982.345–347 are from slide 1900–3–6–379, no. 1982.348 is
 - from slide 1911.11.8 M3725.
 - Diagnosis: Moderately large (750-850 µm long) species of Bythocythere; carapace moderately inflated,
 - greatest width a little behind mid-length. Greatest height well behind mid-length. Dorsal margin convex in female, almost straight in male; ventral margin weakly sinuous in both sexes. Posterior margin denticulate. Dorsomedian sulcus weak. Male copulatory appendage with a relatively large,
 - subtriangular distal process.

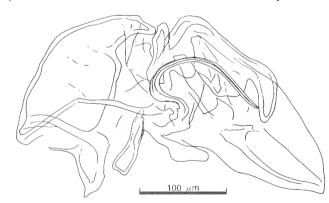
Explanation of Plate 14, 66

Fig. 1, \circlearrowleft LV, ext. lat. (1982.348, 840 μ m long); figs. 2, 3, \updownarrow (1982.346, 810 μ m long): fig. 2, LV, ext. lat.; fig. 3, RV, ext. lat. Scale A (100 μ m; \times 80), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 67

Bythocythere intermedia (3 of 4)

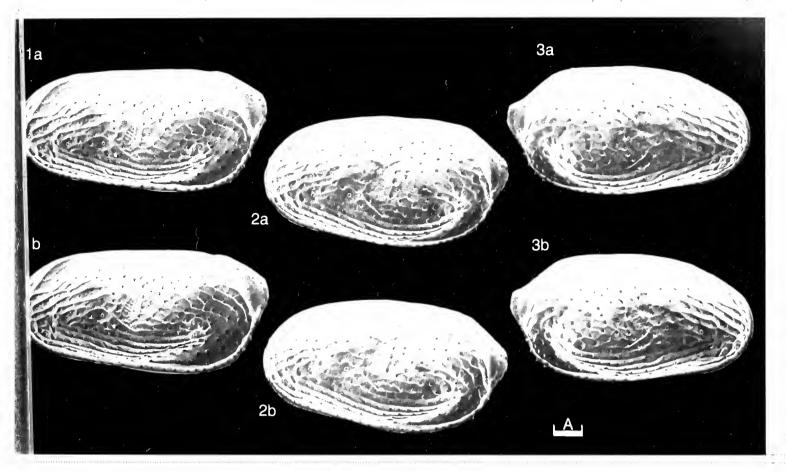
- Remarks: Early records of B. constricta Sars from British waters are now believed to be referable to either B. intermedia or B. zetlandica Athersuch, Horne & Whittaker, 1983 (see Horne Stereo-Atlas
 - intermedia or B. zetlandica Athersuch, Horne & Whittaker, 1983 (see Horne, Stereo-Atlas Ostracod Shells, 14, 69–72, 1987), neither of which possesses the deep median sulcus which is characteristic of Sars' species. B. zetlandica has a smooth posterior margin and is less elongate with a generally less rounded lateral outline than B. intermedia. A closely similar Miocene species, B. neerlandica Kuiper, 1918, is less elongate, less tapered anteriorly, and has a deeper dorsomedian
 - sulcus than B. intermedia.
- Distribution: Fairly common in sublittoral marine waters around British coasts, the southern North Sea, S
 - Norway and Sweden, and as far south as the Bay of Biscay.



Text-fig. 1 Bythocythere intermedia, male copulatory appendage (1982.345).

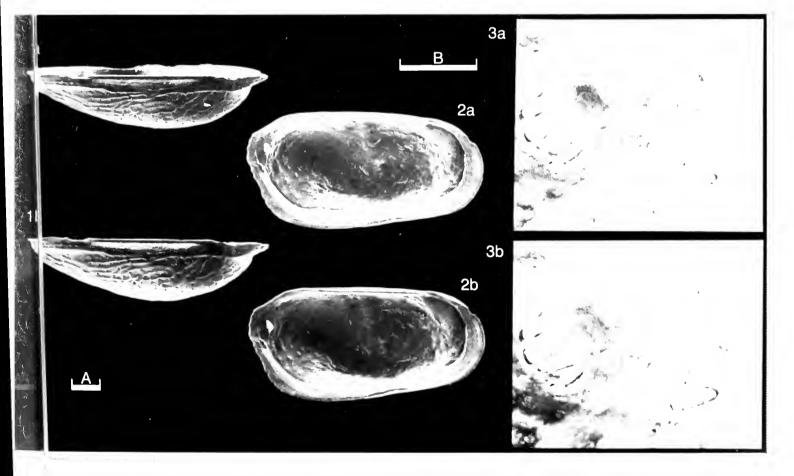
Explanation of Plate 14, 68

Fig. 1, \circlearrowleft LV, dors. (1982.345, 820 μ m long); figs. 2, 3, \circlearrowleft LV (1982.347, 790 μ m long); fig. 2, int. lat.; fig. 3, central muscle scar field. Scale A (100 μ m; ×80), figs. 1, 2; scale B (50 μ m; ×400), fig. 3.



tereo-Atlas of Ostracod Shells 14, 68

Bythocythere intermedia (4 of 4)



595.337.14 (119.9) (411 : 162.002.61 + 415 : 162.011.51) : 551.351

ON BYTHOCYTHERE ZETLANDICA ATHERSUCH, HORNE & WHITTAKER

by David J. Horne (Geology Dept, City of London Polytechnic)

Bythocythere zetlandica Athersuch, Horne & Whittaker, 1983

1868 Bythocythere constricta Sars; G. S. Brady, Trans. Linn. Soc. Land., 26, (pars), 451, pl. 35, fig. 47 only (non pl. 35, figs 48-52) (non Sars, 1866).

1983 Bythocythere zetlandica sp. nov. J. Athersuch, D. J. Horne & J. E. Whittaker, J. micropalaeontol., 2, 73, text-figs 41-n, 5c, pl. 2, figs 5-8.

Holotype: British Museum (Nat. Hist.) no. 1982.350, ♀ carapace and appendages.

[Paratype, no. 1982.351, of carapace and appendages.]

Type locality: Unst Haaf (fishing grounds off Unst), Shetland, approx. lat. 61° 00'N, long. 1° 30'W; Recent,

marine, sublittoral.

Figured specimens: British Museum (Nat. Hist.) nos. 1982.350 (holotype, ♀ LV: Pl. 14, 70, fig. 2; RV: Pl. 14, 70, fig.

3), 1982.351 (paratype, of LV: Pl. 14, 70, fig. 1), 1982.352 (of LV: Pl. 14, 72, fig. 1; copulatory appendage: Text-fig. 1), 1982.353 (of LV: Pl. 14, 72, figs 2-3). All taken from slides labelled "B. constricta" in the Norman Collection at the British Museum (Nat. Hist.): the holotype and paratype are from slide no. 1900-3-6-379; nos 1982.352 and 1982.353, both from Valentia, SW Ireland (approx. lat. 51° 55′N, long. 10° 20′W), are from slides 1900-3-6-379 and 1911.11.8. M3725

respectively.

Explanation of Plate 14, 70

Fig. 1, \circlearrowleft LV, ext. lat. (paratype, **1982.351**, 770 μ m long); figs. 2, 3, \circlearrowleft (holotype, **1982.350**, 790 μ m long): fig. 2, LV, ext. lat.; fig. 3, RV, ext. lat.

Scale A (100 μ m; × 80), figs 1-3.

Stereo-Atlas of Ostracod Shells 14, 71

Bythocythere zetlandica (3 of 4)

Diagnosis:

Moderately large (750-800 μ m long) species of *Bythocythere*; carapace strongly inflated, greatest width a little behind mid-length. Dorsal and ventral margins virtually straight, converging anteriorly, greatest height well behind mid-length. Posterior margin smooth. Dorsomedian sulcus weak. Distal process of male copulatory appendate relatively long, with a convex anterior margin and an almost straight posterior margin.

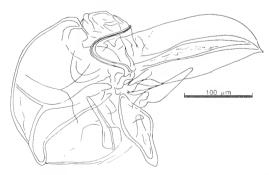
Remarks:

B. zetlandica was formerly confused with B. constricta Sars, which does not live in British waters; Sars' species has a characteristically deep dorsomedian sulcus, and the distal process of its male copulatory appendage is more symmetrical and slender than that of B. zetlandica. A similar NW European species, B. intermedia Elofson, 1938 (see Horne, Stereo-Atlas Ostracod Shells 14, 65-68, 1987), is more elongate than B. zetlandica and has a denticulate posterior margin and a more reputated a particular in lateral view.

rounded outline in lateral view.

Distribution:

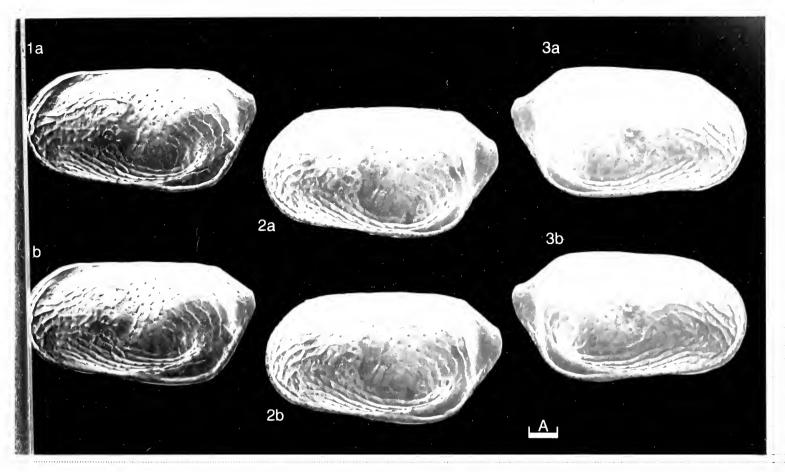
A marine species found in sublittoral waters around British coasts, particularly in the north.



Text-fig. 1 Bythocythere zetlandica, male copulatory appendage (1982.352).

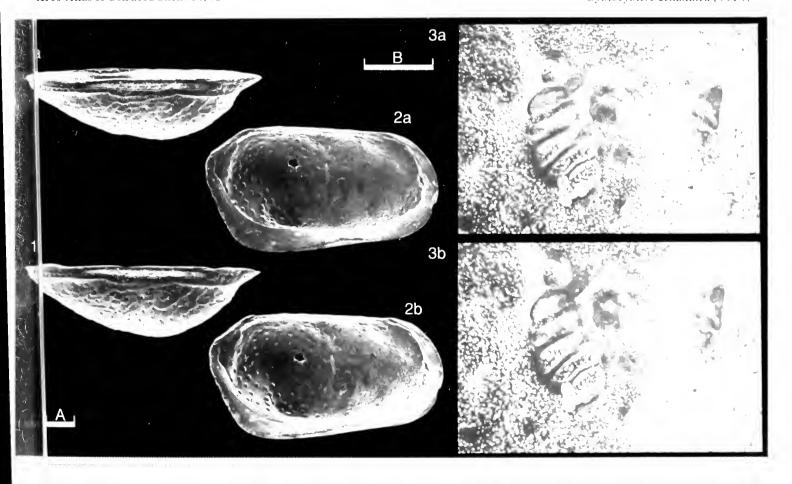
Explanation of Plate 14, 72

Fig. 1, \circlearrowleft LV, dors. (1982.352, 800 μ m long); figs 2, 3 \circlearrowleft LV (1982.353, 780 μ m long): fig. 2, int. lat.; fig. 3, central muscle scar field. Scale A (100 μ m; × 80), figs 1, 2; scale B (50 μ m; × 400), fig. 3.



tereo-Atlas of Ostracod Shells 14, 72

Bythocythere zetlandica (4 of 4)



Stereo-Atlas of Ostracod Shells: Vol. 14, Part 1

CONTENTS

- 14 (1) 1– 4 On Cathaycythere reticulata Whatley & Zhao gen. et sp. nov.; by R. C. Whatley & Zhao Quanhong
- 14 (2) 5–8 On Sinocythere sinensis Hou; by R. C. Whatley & Zhao Quanhong
- 14 (3) 9-12 On Albileberis sinensis Hou; by Zhao Quanhong & R. C. Whatley
- 14 (4) 13–16 On Sinocytheridea impressa (Brady); by Zhao Quanhong & R. C. Whatley
- 14 (5) 17–20 On *Pterygocythereis vannieuwenluisei* Brouwers sp. nov.; by E. M. Brouwers
- 14 (6) 21–24 On *Muellerina liazeli* Coles & Cronin sp. nov.; by G. P. Coles & T. M. Cronin
- 14 (7) 25-28 On Healdianella? aremorica Crasquin sp. nov.; by S. Crasquin
- 14 (8) 29–32 On Maghrebeis tuberculata Majoran gen. et sp. nov.; by S. Majoran
- 14 (9) 33–36 On *Howeina camptocytheroidea* Hanai; by N. Ikeya & E. Compton-Gooding
- 14 (10) 37-40 On Spinoleberis eximia (Bosquet); by J. F. Babinot & J. P. Colin
- 14 (11) 41–44 On Kovalevskiella caudata (Lutz); by P. Carbonel, J. P. Colin & L. Londeix
- 14 (12) 45-48 On Calocaria maurae Vannier gen. et sp. nov.; by J. Vannier
- 14 (13) 49–56 On *Spinohippula esurialis* Vannier, Krůta & Marek gen. et sp. nov.; by J. Vannier, M. Krůta & L. Marek
- 14 (14) 57-64 On Beyrichia (Sagenabeyrichia) siveteri Pollicott subgen. et sp. nov.; by P. D. Pollicott
- 14 (15) 65–68 On Bythocythere intermedia Elofson; by D. J. Horne
- 14 (16) 69-72 On Bythocythere zetlandica Athersuch, Horne & Whittaker; by D. J. Horne

Prepaid annual subscription (valid for Volume 14, 1987)
Individual subscription £22.00 or US \$50.00 for 2 parts (post free)
Price per Part: £22.00 or US \$50.00
Institutional subscription £45.00 or US \$80.00 for 2 parts (post free)

Price per Part: £45.00 or US \$80.00

Back volumes: Vol. 1 (4 Parts): £20.00; price per Part: £5.00

Vol. 2 (4 Parts): £28.00; price per Part: £7.00

Vol. 3 (2 Parts): £24.00; price per Part: £12.00

Vol. 4 (2 Parts): £30.00; price per Part: £15.00

Vol. 5 (2 Parts): £32.00; price per Part: £16.00

Vol. 6 (2 Parts): £40.00; price per Part: £20.00

Vol. 7 (2 Parts): £40.00; price per Part: £20.00

Vol. 8 (2 Parts): £60.00; price per Part: £30.00

Vol. 9 (2 Parts): £60.00; price per Part: £30.00

Vol. 10 (2 Parts): £60.00; price per Part: £30.00

Vol. 11 (2 Parts): £60.00; price per Part: £30.00

Vol. 12 (2 Parts): £60.00; price per Part: £30.00

Vol. 13 (2 Parts): £60.00; price per Part: £30.00

Postage extra in sales of all back Parts No trade discount is allowed on the subscription rate

Orders should be addressed to: Dr R. C. Whatley,
Department of Geology,
University College of Wales,
Aberystwyth, Dyfed.
Cheques should be made payable to B.M.S. (Stereo-Atlas Account)

SPECIAL OFFER

50% off all back part prices if

you become a subscriber to the Atlas

A Stereo-Atlas of Ostracod Shells

edited by R. H. Bate, D. J. Horne, J. W. Neale, and David J. Siveter

Volume 14, Part 2; 30th November, 1987

Published by the British Micropalaeontological Society, London-



Editors

Dr R.H. Bate, SSI (UK) Ltd., Tannery House, Tannery Lane, Send, Woking, Surrey GU23 7EF. Dr D.J. Horne, Department of Geology, City of London Polytechnic, Walburgh House, Bigland Street, London E1 2NG.

Prof. J.W. Neale, Department of Geology, The University, Hull HU6 7RH.

Dr David J. Siveter, Department of Geology, The University, Leicester LE1 7RH.

Editorial Board

Dr G. Bonaduce, Stazione Zoologica, 80121 Napoli, Italy.

Dr J.-P. Colin, Esso Production Research - European, 213 Cours Victor Hugo, 33321 Bègles, France.

Dr P. De Deckker, Research School of Pacific Studies, Australian National University, PO Box 4, Canberra ACT 2600, Australia.

Dr D. van Harten, Universiteit van Amsterdam, Geologisch Instituut, Nieuwe Prinsengracht 130, Amsterdam, The Netherlands.

Dr I. Purper, Departamento de Paleontologia e Estratigrafia, UFRGS, 90 000 Porto Alegre RS, Brazil. Dr R.E.L. Schallreuter, Universität Hamburg, Geologisch-Paläontologisches Institut, Bundesstrasse 55, D 2000 Hamburg 13, West Germany.

Dr Zhao Yuhong, Nanjing Institute of Geology & Palaeontology, Academia Sinica, Chi-Ming-Ssu, Nanjing, People's Republic of China.

Officers of the British Micropalaeontological Society

Chairman Dr A.C. Higgins, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN.

Secretary Dr P.P.E. Weaver, Institute of Oceanographic Sciences, Brook Road, Wormley, Godalming, Surrey GU8 5UB. Tel: 0428-79 4141.

Treasurer Dr J.E. Whittaker, Department of Palaeontology, British Museum (Natural History), Cromwell Road, London SW7 5BD. Tel: 01-589 6323.

Journal Editor Dr. L.M. Sheppard, SSI (U.K.) Limited, Chancellor Court, 20 Priestly Road, Guildford, Surrey GU2 5YL. Tel: (0483) 506605.

Newsletter Editor Dr R.L. Austin, Department of Geology, University of Southampton, Southampton SO9 5NH. Tel: (0703) 559122/557941

Conodont Group Chairman Dr R.J. Aldridge, Department of Geology, University of Nottingham, University Park, Nottingham NG7 2RD.

Secretary Dr P.M. Smith, Department of Earth Sciences, University of Cambridge, Downing Street, Cambridge CB2 3EQ. Tel: (0223) 355463 (or 276121).

Foraminifera Group Chairman Dr P. Copestake, Britoil, 150 St. Vincent Street, Glasgow G2 5LJ. Secretary Dr D.J. Shipp, Robertson Research Int. Limited, Ty'n-y-Coed, Llanrhos, Llandudno LL30 1SA. Tel: (0492) 81811.

Microplankton Group Chairman Dr G.L. Eaton, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN.

Secretary Dr A.J. Powell, BP Research Centre, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN. Tel: (09327) 62818.

Ostracod Group Chairman Dr D.J. Horne, Geology Department, City of London Polytechnic, Walburgh House, Bigland Street, London E1 2NG.

Secretary Dr C. Maybury, Department of Geology, University College of Wales, Aberystwyth, Dyfed SY23 3DB. Tel: (0970) 3111.

Palynology Group Chairman Dr M.C. Boulter, N.E. London Polytechnic, Romford Road, London E15 4LZ.

Secretary Dr J.E.A. Marshall, Department of Geology, The University, Southampton SO9 5NH. Tel: (0703) 559122.

Calcareous Nannofossil Group Chairman Mr M. Jakubowski, Robertson Research Int. Limited, Ty'n-y-Coed, Llanrhos, Llandudno, Gwynedd LL30 1SA.

Secretary Dr J. Crux, BP Research Centre, Chertsey Road, Sunbury on Thames, Middlesex TW16 7LN. Tel: (09327) 63062.

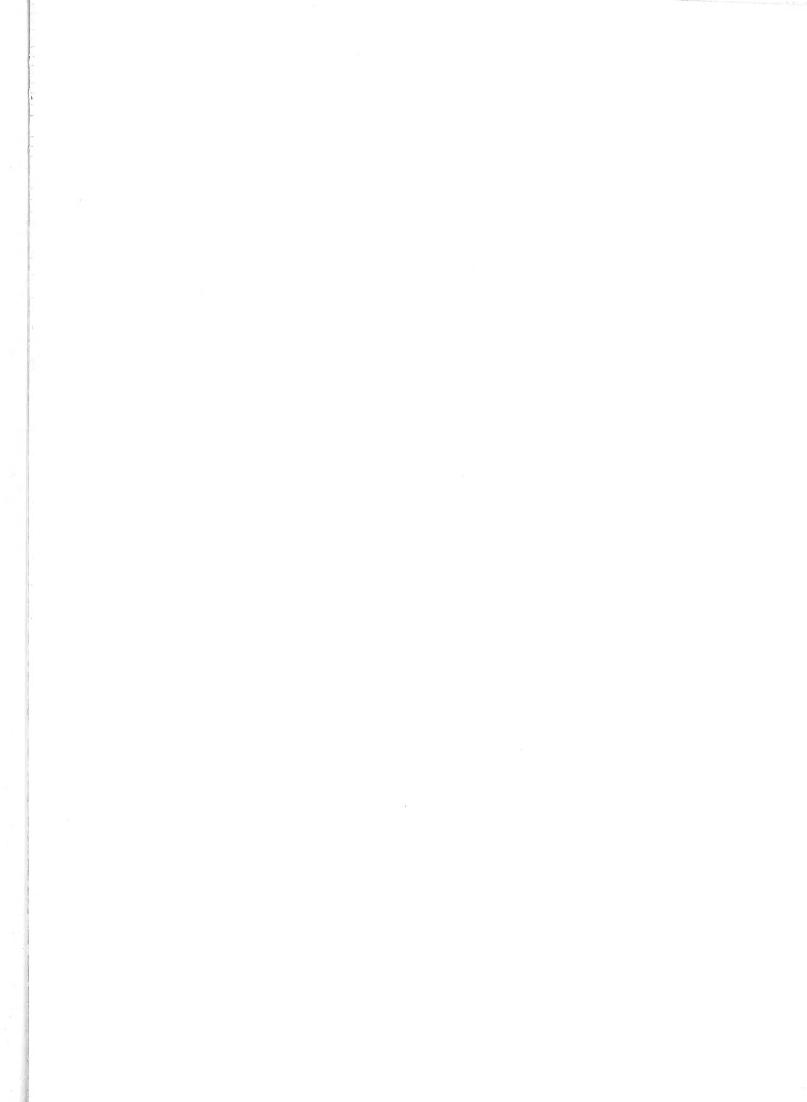
Instructions to Authors

Contributions illustrated by scanning electron micrographs of Ostracoda in stereo-pairs are invited. Format should follow the style set by the majority of papers in this issue. Descriptive matter apart from illustrations should be cut to a minimum; preferably each plate should be accompanied by one page of text only. Blanks to aid in mounting figures for plates may be obtained from any one of the Editors or Editorial Board. Completed papers should be sent to Dr David J. Siveter.



Financial support from The British Petroleum Company p.l.c. for the publication of this issue is gratefully acknowledged.

The front cover shows a left valve of *Neolimnocythere hexaceros* Delachaux, 1928, from Quaternary Deposits at Lago Junin, Peru. Photograph by Dr P. De Deckker, University of Monash, Victoria, Australia.



ON KUIPERIANA ROBUSTA WHATLEY & MAYBURY sp. nov.

by Robin Whatley & Caroline Maybury (University College of Wales, Aberystwyth)

Kuiperiana robusta sp. nov.

British Museum (Nat. Hist.) no. OS 12976, Q LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12977, OS 12978].

Blue Clay, sample no. 29, NW corner of Vicarage Pit, St. Erth, Cornwall, England (Nat. Grid *Type locality:*

Ref. SW 556352); Upper Pliocene.

Latin, from the robust nature of the valves. Derivation of name:

British Museum (Nat. Hist.) nos. **OS 12976** (holotype, ♀ LV: Pl. 14, 74, fig. 1; Pl. 14, 76, fig. 2), Figured specimens:

OS 12977 (paratype, ♀ RV: Pl 14, 74, fig. 2; Pl. 14, 76, figs. 1, 3, 4), OS 12978 (paratype, ♂ LV:

Pl. 14, 74, fig. 3). All from the type locality and horizon.

Explanation of Plate 14, 74

Fig. 1, ♀ LV, ext. lat. (holotype, OS 12976, 550 µm long); fig. 2, ♀ RV, ext. lat. (paratype, OS 12977, 560 µm long); fig. 3, ♂ LV, ext. lat. (paratype, OS 12978, 550 μm long).

Scale A (100 μ m; × 104), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 75

Kuiperiana robusta (3 of 4)

Diagnosis:

Medium-sized, strongly dimorphic with circular to subcircular, regularly disposed punctae medianly and reticulae peripherally. Dorsomedianly with 3 short, inclined sulcate depressions. Posterior marginal rim narrow and alar process bluntly rounded. Eye tubercle inconspicuous. Inner lamella moderately wide, undulose posteroventrally with a wide ventral flange and selvage and list developed. Hinge gongylodont with a long, thin, smooth groove/bar medianly; the posterior terminal element of the right valve is a curved tooth with a frill-like dorsal surface.

Remarks:

This species differs from the type-species, K. wanneri wanneri (Kuiper, 1918) (W. N. Kuiper, Oligocane und Miocane Ostracoden aus den Niederlanden, publ. PhD thesis, Groningen, 26–27, pl. 1, figs. 8a-c, 1918 and M.A.A. Bassiouni, *Roemeriana*, 3, 62-66, pl. 8, figs. 1-3, 1962) in that its reticulae are less regularly ordered ventrally, its eye tubercle is difficult to distinguish from ornament (whereas it is well defined in K. wanneri wanneri) and it possesses an alar process. (K. wanneri wanneri is inflated ventrally, but lacks a clearly defined alar protuberance). Both species possess punctate and reticulate ornament and have elongate, subrectangular lateral outlines.

The ratio of adult to juvenile specimens of K. robusta in the authors' material is low (1:43), with

only 5 adult specimens recovered.

Distribution:

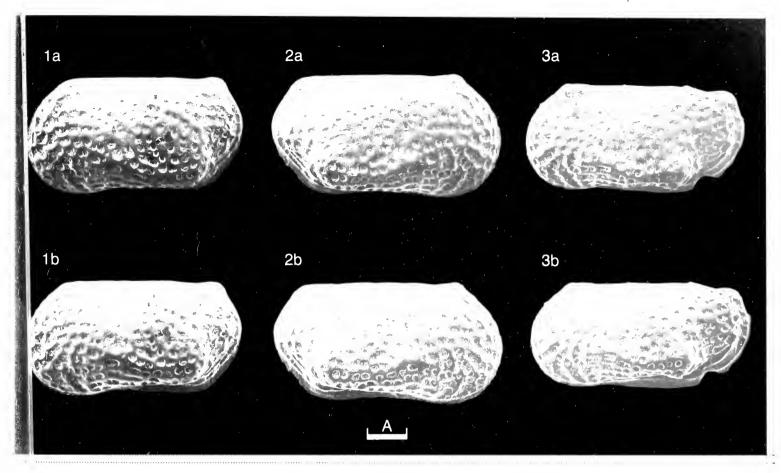
Upper Pliocene deposits of St. Erth, England (sample nos. 1-4, 7, 11, 16, 18, 22-23, 25-29) and Upper Pliocene (Redonian) deposits of Apigné (Borehole II, Le Temple du Cerisier), Le Bosq d'Aubigny and Saint-Jean-la-Poterie (sample no. 1549.15); NW France. See C. Maybury (Taxonomy, Palaeoecology and Biostratigraphy of Pliocene Benthonic Ostracoda from St. Erth and NW France, unpubl. PhD thesis, Univ. Wales, 1, 3-29, 1985) and J. -P. Margerel (Les Foraminifères du Redonien. Systématique, Répartition stratigraphique, Paléoécologie, Nantes, 1,

8–26, 1968) for geographical, stratigraphical and sample details.

Explanation of Plate 14, 76

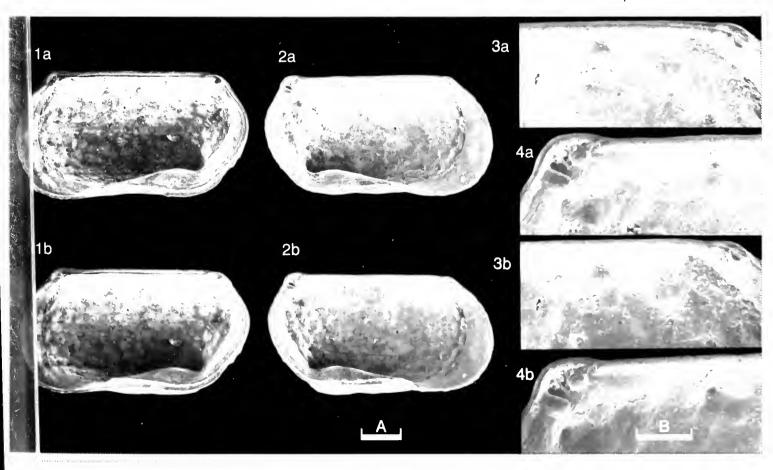
Figs. 1, 3, 4, \bigcirc RV (paratype, OS 12977, 560 μ m long): fig. 1, int. lat.; fig. 3, ant. hinge element; fig. 4, post. hinge element; fig. 2, \bigcirc LV, int. lat. (holotype, OS 12976, $550\mu m$ long).

Scale A (100 μ m; × 104), figs. 1, 2; scale B (40 μ m; × 330), figs. 3, 4.



tereo-Atlas of Ostracod Shells 14, 76

Kuiperiana robusta (4 of 4)



ON LOXOCAUDA SUBQUADRATA MAYBURY & WHATLEY sp. nov.

by Caroline Maybury & Robin Whatley (University College of Wales, Aberystwyth)

Loxocauda subquadrata sp. nov.

Holotype: British Museum (Nat. Hist.) no OS 12906, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12907-OS 12909].

Type locality: Blue Clay, sample no. 16, Vicarage Pit, St. Erth, Cornwall, England (Nat. Grid Ref. SW 556352);

Upper Pliocene.

Derivation of name: Latin,

Latin, from the outline of the valves in lateral view.

Figured specimens: British Museum (Nat. Hist.) nos. OS 12906 (holotype, Q LV: Pl. 14, 78, fig. 1), OS 12907

(paratype, ♀ RV: Pl. 14, 78, fig. 2). OS 12908 (paratype, ♂ LV: Pl. 14, 78, fig. 3; Pl. 14, 80, figs. 2–4), OS 12909 (paratype, ♂ RV: Pl. 14, 80, fig. 1). All from the type locality: specimen OS 12907 is from Mottled Clean Clay (sample no. 2); specimen OS 12908 is from a mixed sample (no. 7) and specimen OS 12909 is from the same sample as the holotype. See C. Maybury, *Taxonomy*, *Palaeoecology and Biostratigraphy of Pliocene Benthonic Ostracoda from St. Erth and NW France*,

unpub. PhD thesis, Univ. Wales, 1, 3-6, 1985 for sample details.

Explanation of Plate 14, 78

Fig. 1, Q LV, ext. lat. (holotype, **OS 12906**, 380 μ m long); fig. 2, Q RV, ext. lat. (paratype, **OS 12907**, 390 μ m long); fig. 3, O LV, ext. lat. (paratype, **OS 12908**, 430 μ m long). Scale A (100 μ m; ×160), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 79

Loxocauda subquadrata (3 of 4)

Diagnosis:

A very small to small, subquadrate species of *Loxocauda* characterised by a lateral surface with 4 obliquely disposed ridges posterodorsally and traces of a reticulum anteromedianly and anteroventrally; remainder smooth. Free marginal areas strongly compressed with a prominent, curved, sub-alar process posteriorly and posteroventrally. Hinge unusual: comprising in the left valve, a smooth bar with its anterior and posterior ends enclosed by narrow, horizontal, "u"-shaped sockets themselves bounded by "u"-shaped ridges. Muscle scars comprising four contiguous adductors, a "v"-shaped frontal and two subcircular mandibular scars.

Remarks:

The genus Loxocauda is known only from three previously described species: the type-species, L. muelleri Schornikov, 1969 (in: F. D. Mordukhai-Boltovskoi, (Ed.) Identification Key to the Fauna of the Black and Azov Seas, 2, 201, pl. 28, fig. 1, Kiev, 1969), L. fragilis (Sars, 1866) (G. O. Sars, Forh. Vidensk Selsk. Krist., 1865, 65–66, 1866 and An account of the Crustacea of Norway, 9, Ostracoda, pts. 13, 14, 222, pl. 102, fig. 3, 1926) and L. decipiens (G. W. Müller, 1894) (G. W. Müller, Fauna Flora Golf. Neapel, 21, 347–348, pl. 27, figs. 10–14, 24, pl. 29, figs. 2, 9, 1894). All these species differ from the new species in that they lack the traces of a reticulum and the sub-alar process which are characteristic of L. subquadrata. The present species (and all known Loxocauda species) resemble Pseudocythere Sars in shape and outline. The two genera differ, however, in their musculature, hingement and appendages.

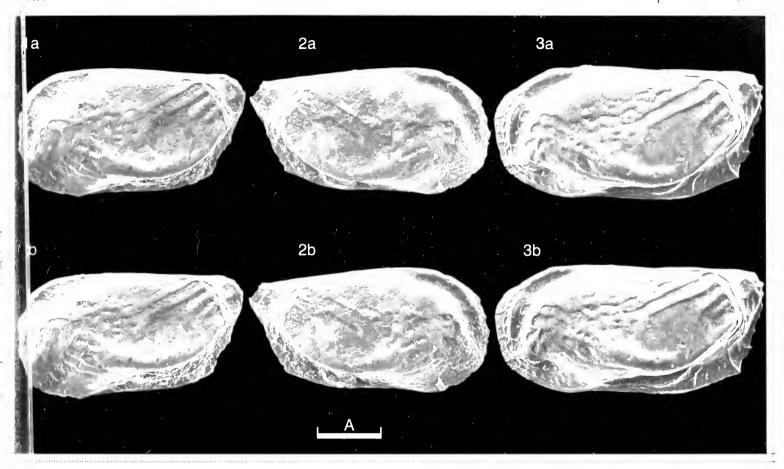
Distribution:

The species is known only from the Upper Pliocene deposits of St. Erth, Cornwall, England (samples nos. 1-4, 7, 16, 21, 23, 25-28, C. Maybury, op. cit.).

Explanation of Plate 14, 80

Fig. 1, \circlearrowleft RV, ext. lat. (paratype, OS 12909, 400 μ m long); figs. 2–4, \circlearrowleft LV, (paratype, OS 12908, 430 μ m long); fig. 2, int. lat.; fig. 3, ant. hinge element; fig. 4, post. hinge element.

Scale A (100 μ m; ×160), figs. 1, 2; scale B (40 μ m; ×400), figs. 3, 4.



ON SAGMATOCYTHERE MINUTA MAYBURY & WHATLEY sp. nov.

by Caroline Maybury & Robin Whatley (University, College of Wales, Aberystwyth)

Sagmatocythere minuta sp. nov.

Holotype: British Museum (Nat. Hist.) no. OS 12849, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12850 – OS 12853].

Type locality: Mixed sample, sample no. 7, Vicarage Pit. St. Erth, Cornwall, England (Nat. Grid Ref. SW

556352); Upper Pliocene.

Derivation of name: Latin, referring to the very small size of the species.

Figured specimens: British Museum (Nat. Hist.) nos. OS 12849 (holotype, Q LV: Pl. 14, 82, fig. 1), OS 12850

(paratype, ♀ RV: Pl. 14, 82, fig. 2), OS 12852 (paratype, ♂ RV: Pl. 14, 82, fig. 3), OS 12851 (paratype, ♂ LV: Pl. 14, 84, fig. 1), OS 12853 (paratype, ♀ RV: Pl. 14, 84, fig. 2), OS 12854 (paratype, ♂ LV: Pl. 14, 84, fig. 3). Specimens OS 12850 and OS 128854 from the same sample as the holotype; the remaining paratypes from Brown Clay (sample no. 28) at the type locality. See C. Maybury, *Taxonomy*, *Palaeoecology and Biostratigraphy of Pliocene Benthonic Ostracoda from St. Erth and NW France*, unpub. PhD thesis, Univ. Wales, 1, 3–6, 1985 for sample details.

Explanation of Plate 14, 82

Fig. 1, \bigcirc LV, ext. lat. (holotype, **OS 12849**, 370 μ m long); fig. 2, \bigcirc RV, ext. lat. (paratype, **OS 12850**, 370 μ m long); fig. 3, \bigcirc RV, ext. lat. (paratype, **OS 12852**, 380 μ m long).

Scale A (100 μ m; × 161), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 83

Sagmatocythere minuta (3 of 4)

Diagnosis:

A very small, strongly dimorphic, alate species of *Sagnatocythere* with a complex ornament of nodes and reticulae. In females there are two anterodorsal nodes, in males only one; their position is reflected internally by shallow circular to subcircular depressions. The anterior of the two anterodorsal nodes in the female and the anterodorsal node of the male is situated below the eye tubercle and is connected to it by a smooth rib. Posterodorsal node well defined and bearing a prominent, inverted u-shaped dorsal loop. Selvage well developed and blade-like mid-ventrally.

Remarks:

This species, Sagmatocythere alaefortis alaefortis Whatley & Maybury (Stereo-Atlas Ostracod Shells, 14, 85–88, 1987) and S. alaefortis gallica Whatley & Maybury (Stereo-Atlas Ostracod Shells, 14, 89–92, 1987) form a distinct group of Sagmatocythere whose noded and irregularly reticulate ornament distinguishes them from the "napoliana" and "multifora" groups. The "napoliana" group comprises S. napoliana (Puri, 1963) (see J. Athersuch, Stereo-Atlas Ostracod Shells, 3, 117–124, 1976), S. cristatissima (Ruggieri, 1967) (Riv. ital. Paleont. Stratigr., 73, 374–376, figs. 37–38) and S. wyatti Maybury & Whatley, 1987 (Stereo-Atlas Ostracod Shells, 14, 93–96). These strongly reticulate species all possess muri which are narrow and almost "blade-like". The "multifora" group comprises S. multifora (Norman, 1865) (In: G. S. Brady, Nat. Hist. Trans. Northumberland and Durham, 1, 18–19, pl. 6, figs. 13–16), S. littoralis (G. W. Müller, 1894) (Fauna Flora Golf. Neapel 21, 346, pl. 27, fig. 9, pl. 29, figs. 1, 7), S. paracercinata Whatley & Maybury, 1984 (Stereo-Atlas Ostracod Shells, 11, 21–24) and S. pseudomultifora Maybury & Whatley, 1984 (Stereo-Atlas Ostracod Shells, 11, 25–28). Species of this group are alate and possess regular reticulate ornament. The small size of the adults of S. minuta make it the smallest Sagmatocythere yet recorded.

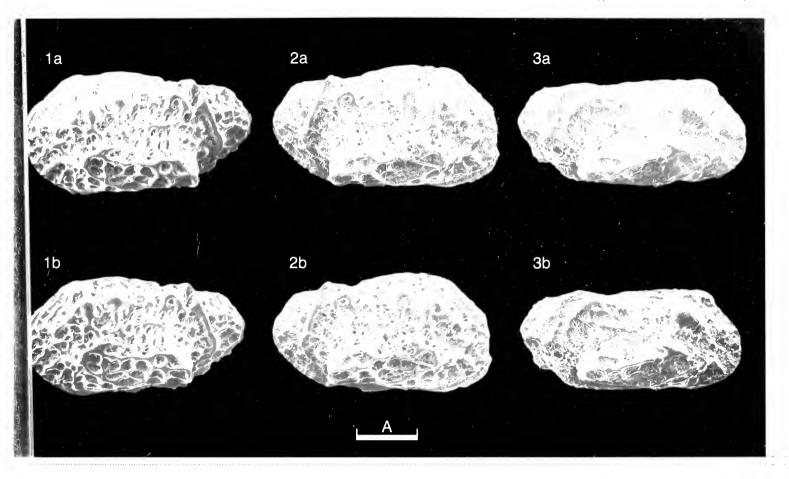
Distribution:

The species has been recovered from the Upper Pliocene deposits of St. Erth, Cornwall, England (sample nos 1–4, 7, 16, 18, 23, 25–28) and the Upper Pliocene (Redonian) deposits of Apigné (Le Temple du Cerisier), NW France. (See C. Maybury, *op. cit.*, for sample details).

Explanation of Plate 14, 84

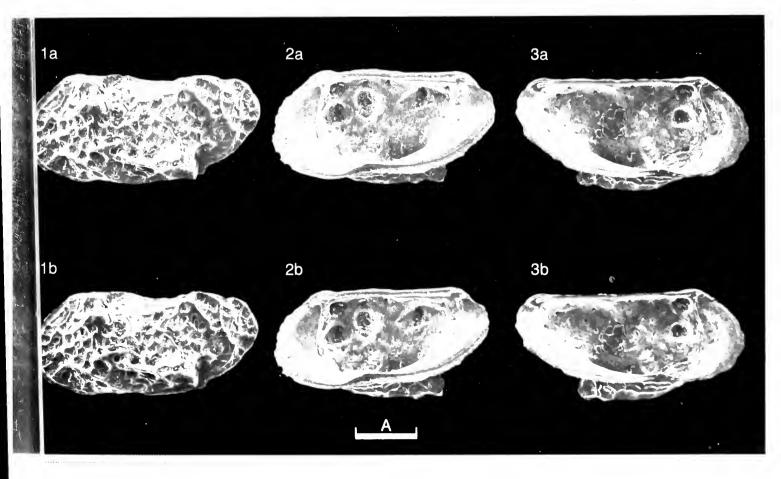
Fig. 1, \circlearrowleft LV, ext. lat. (paratype, OS 12851, 380 μ m long); fig. 2, \circlearrowleft RV, int. lat. (paratype, OS 12853, 370 μ m long); fig. 3, \circlearrowleft LV, int. lat. (paratype, OS 12854, 380 μ m long).

Scale A (100 μ m; × 161), figs. 1–3.



2reo-Atlas of Ostracod Shells 14, 84

Sagmatocythere minuta (4 of 4)



ON SAGMATOCYTHERE ALAEFORTIS ALAEFORTIS WHATLEY & MAYBURY sp. & subsp. nov.

by Robin Whatley & Caroline Maybury (University College of Wales, Aberystwyth)

Sagmatocythere alaefortis alaeforti sp. & subsp. nov.

Holotype: British Museum (Nat. Hist.) no. OS 12843, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12844-OS 12848].

Type locality: Brown Clay, sample no. 28, Vicarage Pit, St. Erth, Cornwall, England (Nat. Grid Ref. SW

556352); Upper Pliocene.

Derivation of name: Latin, from the strongly developed alae of this species.

Figured specimens: British Museum (Nat. Hist.) nos. OS 12843 (holotype, Q LV: Pl. 14, 86, fig. 1), OS 12844

(paratype, ♀ RV: Pl. 14, 86, fig. 2), OS 12845 (paratype, ♂ LV: Pl. 14, 86, fig. 3), OS 12846 (paratype, ♂ RV: Pl. 14, 88, fig. 1), OS 12847 (paratype, ♂ RV: Pl. 14, 88, fig. 2), OS 12848 (paratype, ♂ LV: Pl. 14, 88, fig. 3). Specimens OS 12845, OS 12846 and OS 12848 are from the same sample as the holotype. Specimen OS 12844 is from a bulk sample (sample no. 1) and specimen OS 12847 from a mixed sample; both are from the type locality and horizon. See C. Maybury, Taxonomy, Palaeoecology and Biostratigraphy of Pliocene Benthonic Ostracoda from St. Erth and NW France, unpub. PhD thesis, Univ. Wales, 1, 3-6, 1985 for sample details.

Explanation of Plate 14, 86

Fig. 1, Q LV, ext. lat. (holotype, **OS 12843**, 450 μ m long); fig. 2, Q RV, ext. lat. (paratype, **OS 12844**, 460 μ m long); fig. 3, Q LV, ext. lat. (paratype, **OS 12845**, 500 μ m long). Scale A (100 μ m; × 125), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 87

Sagmatocythere alaefortis alaefortis (3 of 4)

Diagnosis: A medium, subrectangular subspecies of Sagmatocythere with a straight dorsal margin and strongly developed alae. Anterior and posterior marginal areas flattened and with ornament less well

developed. Reticulate mid-dorsally and dorsomedianly. Ribs massive and angular in the alar region of the valve with two deep depressions midventrally. Eye tubercle irregular in outline and connected with a subrounded, anterodorsal node. Thickened ribs in the posterodorsal area of the

female and male left valve assume a more noded character in the male right valve.

Remarks: The posterodorsal protuberance/node and irregularly reticulate ornament of this species is similar

to that of certain species of Loxocorniculum Benson & Coleman, 1963 (Paleont. Contr. Univ. Kansas., no. 31, 38) such as the type-species, L. fischeri (Brady, 1869) (In: L. De Folin & L. Périer (eds.), Les Fonds de la Mer, 1(1), 154, pl. 18, figs. 15–16, 1869). The present authors, however, differentiate the two genera on the basis of their hinge structure: species of Sagmatocythere possessing a gongylodont hinge with a smooth median element and species of

Loxocorniculum a gongylodont hinge with a strongly denticulate median element.

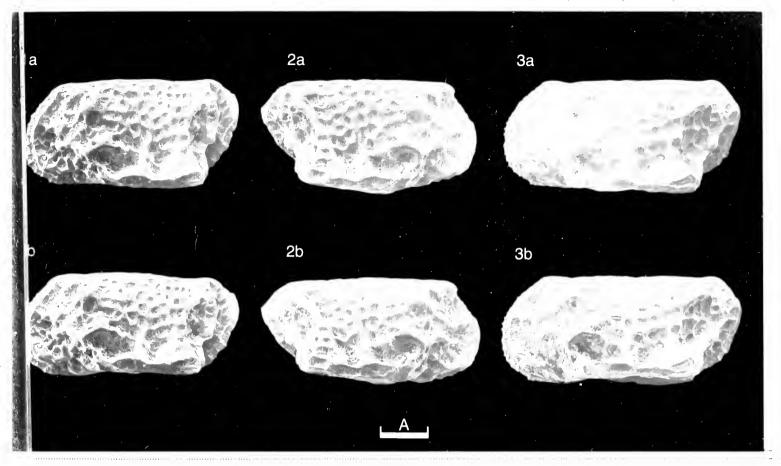
Distribution: Upper Pliocene deposits of St. Erth, Cornwall, England (sample nos. 1–4, 7, 11, 21, 23, 25–29; see

C. Maybury, op. cit., 1, 3–6 for sample details).

Explanation of Plate 14, 88

Fig. 1, O RV, ext. lat. (paratype, **OS 12846**, 500 μm long); fig. 2, Q RV, int. lat. (paratype, **OS 12847**, 450 μm long); fig. 3, O LV, musc. sc. (paratype, **OS 12848**, 500 μm long).

Scale A (100 μ m; ×125), figs. 1, 2; scale B (10 μ m; ×700), fig. 3.



reo-Atlas of Ostracod Shells 14, 88

Sugmatocythere alactorus (4 of 4)

2a

3b

ON SAGMATOCYTHERE ALAEFORTIS GALLICA WHATLEY & MAYBURY subsp. nov.

by Robin Whatley & Caroline Maybury (University College of Wales, Aberystwyth)

Sagmatocythere alaefortis gallica subsp. nov.

Holotype:

British Museum (Nat. Hist.) no. OS 12839, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12840 - OS 12842].

Type locality:

Shell-rich sand, Le Temple du Cerisier, SW of Rennes (approx. lat. 48° 07'N, long. 1° 41'W), NW

France; Upper Pliocene, Redonian.

Derivation of name:

Latin, referring to the fact that the subspecies has only been found in the Redonian deposits of

France.

Figured specimens:

British Museum (Nat. Hist.) nos. OS 12839 (holotype, ♀ LV: Pl. 14, 90, fig. 1), OS 12840 (paratype, ♀ RV: Pl. 14, 90, fig. 2), OS 12841 (paratype, ♂ LV: Pl. 14, 90, fig. 3), OS 12842

(paratype, O RV: Pl. 14, 92, figs. 1-4). All from the type locality and horizon.

Explanation of Plate 14, 90

Fig. 1, \mathcal{Q} LV, ext. lat. (holotype, OS 12839, 430 μ m long); fig. 2, \mathcal{Q} RV, ext. lat. (paratype, OS 12840, 440 μ m long); fig. 3, \mathcal{O} LV, ext. lat. (paratype, **OS 12841**, 480μm long). Scale A (100 μ m; × 135), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 91

Sagmatocythere alaefortis gallica (3 of 4)

Diagnosis:

A small subspecies of Sagmatocythere characterised by a straight dorsal margin in female specimens and a slightly concave dorsal margin in males. The ornament is regularly reticulate with the majority of fossae circular to suboval in outline. Fossae of the alar region of the valve are comparatively large and have irregular, angular outlines. In the left valve there is a prominent, postercdorsal loop; this is less prominent in the right valve. Inner lamella broad with a

conspicuous, blade-like selvage ventrally.

Remarks:

Sagmatocythere alaefortis gallica differs from S. alaefortis alaefortis Whatley & Maybury (Stereo-Atlas Ostracod Shells, 14, 85–88, 1987) in its smaller size and by having a greater portion of its lateral surface covered by a reticulum. The reticulum is also more regular in S. alaefortis gallica than in the nominate subspecies and the posterodorsal protuberance of S. alaefortis alaefortis is reduced to a posterodorsal loop in S. alaefortis gallica. The distinctive, blade-like selvage of S.

alaefortis gallica also serves to distinguish it from S. alaefortis alaefortis.

Distribution:

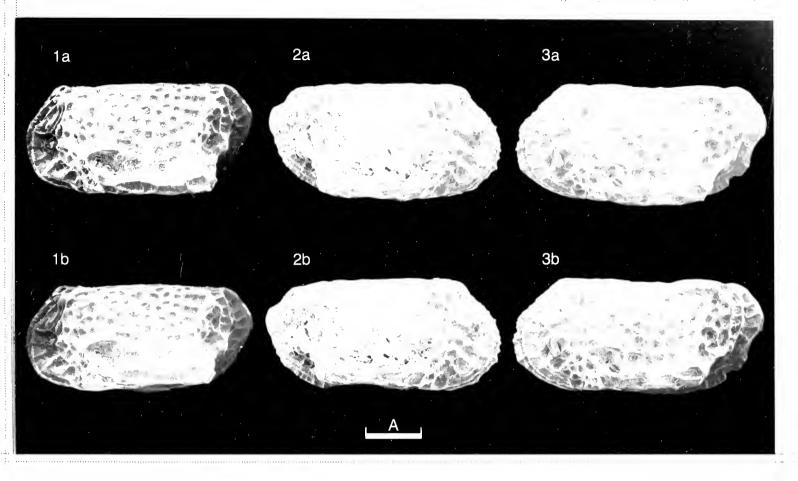
This subspecies occurs in the Redonian (Upper Pliocene) deposits of Apigné (Le Temple du Cerisier) and of Falleron (approx. lat. 46° 60'N; long. 1° 41'W). It has also been recovered in a mixed sample from NW France, also of Redonian age. See J. –P. Margerel, *Les Foraminifères du* Redonian. Systématique, Répartition stratigraphique, Paléoécologie, Nantes, 1, 8-26, 1968 for

geographical, stratigraphical and sample details.

Explanation of Plate 14, 92

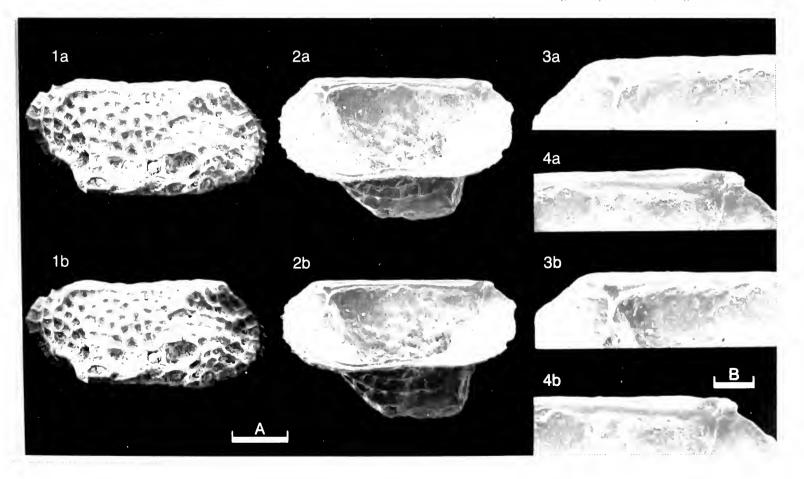
Fig. 1-4, O RV (paratype, OS 12842, 460μm long): fig. 1, ext. lat; fig. 2, int. lat.; fig. 3, ant. hinge element; fig. 4, post. hinge element.

Scale A $(100\mu m; \times 135)$, figs. 1, 2; scale B $(40\mu m; \times 265)$, figs. 3, 4.



Stereo-Atlas of Ostracod Shells 14, 92

Sagmatocythere alaefortis gallıca (4 ot 4)



ON SAGMATOCYTHERE WYATTI MAYBURY & WHATLEY sp. nov.

by Caroline Maybury & Robin Whatley (University College of Wales, Aberystwyth)

Sagmatocythere wyatti sp. nov.

Holotype:

British Museum (Nat. Hist.) no. OS 12861, ♀ LV.

[Paratypes: British Museum (Nat. Hist.) nos. OS 12862 - OS 12865].

Type locality:

Mixed sample, sample no. 7, Vicarage Pit, St. Erth, Cornwall, England (Nat. Grid Ref. SW

556352); Upper Pliocene.

Derivation of name: Figured specimens: Latin, in honour of Mr. Antony Wyatt in recognition of his work on 'wobbling continents'. British Museum (Nat. Hist.) nos. OS 12861 (holotype, ♀ LV: Pl. 14, 94, fig. 1), OS 12862 (paratype, of LV: Pl. 14, 94, fig. 2), OS 12863 (paratype, of RV: Pl. 14, 94, fig. 3), OS 12864 (paratype, O RV: Pl. 14, 96, figs. 1, 3, 4), OS 12865 (paratype, juv. LV: Pl. 14, 96, fig. 2). All specimens from the type locality; OS 12863 and OS 12865 are from the same sample as the holotype, but OS 12862 is from a sample of blue clay (no. 25) and OS 12864 from a mixed sample (no. 1). See C. Maybury, Taxonomy, Palaeoecology and Biostratigraphy of Pliocene Benthonic Ostracoda from St. Erth and NW France, unpub. PhD thesis, Univ. Wales, 1, 3-6, for sample details.

Explanation of Plate 14, 94 Fig. 1, \mathcal{P} LV, ext. lat. (holotype, OS 12861, 500 μ m long); fig. 2, \mathcal{O} LV, ext. lat. (paratype, OS 12862, 460 μ m long); fig. 3, \mathcal{O} RV, ext. lat. (paratype, OS 12863, $470\mu m$ long).

Scale A (100 μ m; × 127), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 95

Sagmatocythere wyatti (3 of 4)

Diagnosis:

A small to medium species of Sagmatocythere with an irregularly reticulate ornament of large fossae and narrow, blade-like muri. There are four obliquely disposed, subparallel muri posteriorly and a deeply excavated area posteroventrally. Eye tubercle small, smooth, tear-shaped and connecting with a narrow murus which extends anteroventrally, parallel to the anterior

Remarks:

This species and the type-species, Sagmatocythere napoliana (Puri, 1963) (see J. Athersuch, Stereo-Atlas Ostracod Shells 3, 117-124, 1976), a Miocene to Recent Mediterranean species, are similar in that certain units or 'cells' of the reticulum can be traced in both species. There is, for example, a prominent, polygonal posterior unit and a deeply excavated posteroventral area. In addition, there are 5-7 conjunctive pore conuli anteriorly, the muri of both species are narrow and blade-like and the fossae comparatively large. The two species differ in lateral outline; S. napoliana is much more elongate than S. wyatti and, whereas the former species has a dorsal margin with a concavity or 'saddle', the dorsal margin of S. wyatti is straight and obliquely sloped.

Distribution:

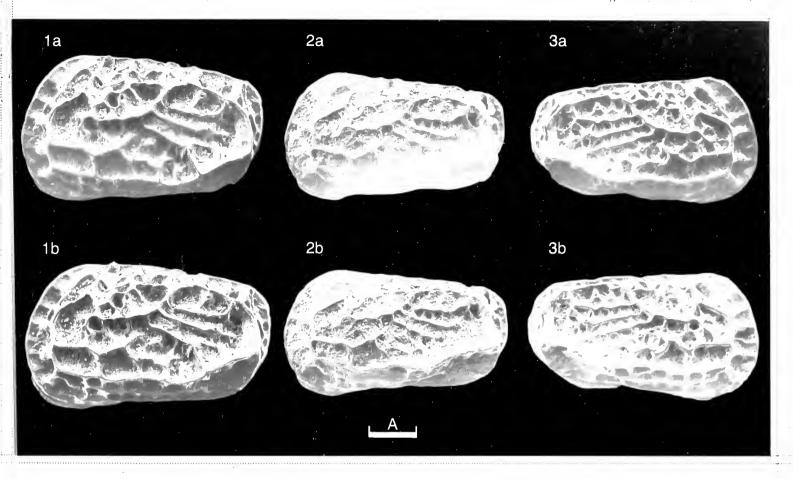
In addition to its occurrence in the Upper Pliocene deposits of St. Erth, Cornwall, England (sample nos. 1, 7, 10, 23, 25, 28–29; see C. Maybury, op. cit., for details), this species has been found in a Redonian (Upper Pliocene) sample of shell-rich sand from Le Temple du Cerisier, SW of Rennes (approx. lat. 48° 07'N, long. 1° 41'W), NW France (see, J. -P. Margerel, Les Foraminifères du Redonien. Systématique, Répartition stratigraphique, Paléoécologie, Nantes, 1,

7–13, 1968 for further sample details).

Explanation of Plate 14, 96

Figs. 1, 3, 4, ♂ RV (paratype, OS 12864, 460μm long): fig. 1, int. lat.; fig. 3, ant. hinge element; fig. 4, post. hinge element; fig. 2, juv. LV, musc. sc. (paratype, **OS 12865**, 410 µm long).

Scale A (100 μ m; × 127), fig. 1; scale B (10 μ m; × 740), fig. 2; scale C (40 μ m; × 330), figs. 3, 4.



 $595.337.14 \ (118.22.+119.9) \ (45:161.009.45+261.268:162.004.50+411:162.002.61+411:162.006.55+496.1:161.026.38):$ 551.351

ON CARINOCYTHEREIS CARINATA (ROEMER)

by John Athersuch & John E. Whittaker (B.P. Research Centre, Sunbury and British Museum (Natural History), London)

Genus CARINOCYTHEREIS Ruggieri, 1956

Type-species (by original designation): Cytherina carinata Roemer, 1838

Diagnosis:

Quadrate trachyleberid with three subparallel ponticulate, sometimes discontinuous carinae; ventral carina strongest. Ventral margin carinate; anterior margin carinate and/or denticulate; posterior margin spinose or denticulate; area between carinae tuberculate or mammilate. Male RV dimorphic; posteroventral region of valve devoid of ventral carina and compressed. Eye tubercle prominent. Hinge amphidont or heterodont; anterior tooth of RV stepped; posterior tooth crenulate.

Seta of second podomere of antennula long in both sexes; endopodite of antenna with three long setae; exopodite dimorphic, long and three-jointed in male, short and possibly only two-jointed in female.

Remarks:

Carinocythereis differs from Occlusacythere Ruggieri & Russo, 1980, in possessing ponticulate carinae.

Expanation of Plate 14, 98

 $1000 \,\mu\text{m}$ long).

Scale A (250 μ m; ×60), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 99

Carinocythereis carinata (3 of 6)

Carinocythereis carinata (Roemer, 1838)

Cytherina carinata sp. nov. F.A. Roemer, Neues Jb Miner. Geogn. Geol. Petrefakt., 1838, 518, pl. 6, fig. 28. 1838

Cythereis antiquata sp. nov. W. Baird, Natural History of British Entomostraca, Ray Soc., London, 176, pl. 20, fig. 2. 1850

Cythere antiquata (Baird); G. S. Brady, Trans. Linn. Soc. Lond., 26, 417, pl. 30, figs. 17-20. 1868

Carinocythereis antiquata (Baird); F. E. Caraion, Revue Biol. Buc., 5, 123, figs. 4a, b. 1960

Carinocythereis carinata Roemer; G. Ruggieri & A. Russo, Boll. Soc. paleont. ital., 19, 30, pl. 2, fig. 8; text-fig. 2 (neotype). 1980

Carinocythereis antiquata (Baird); N. Doruk, Stereo-Atlas Ostracod Shells, 8, 63-70. 1981

Neotype: Designated by Ruggieri & Russo, op. cit., a female RV; housed in the Institute of Palaeontology, University of Modena, Italy, cat. no. 19252. (Refigured herein, Pl. 14, 100, fig. 1). The original type material of C. carinata is missing (only label exists) from the Roemer Collection, Roemer Museum, Hildesheim, West Germany (Athersuch & Whittaker, 1986, Br. Micropalaeontologist,

Type locality: Figured specimens: Castellarquarto, Piacenza, N Italy (approx. lat 45°00'N, long. 9°40'E); Late Pliocene. British Museum (Nat. Hist.) nos. 1984.180 (♀ car.: Pl. 14, 98, fig. 1; Pl. 14, 100, fig. 3), 1984.181 (O'RV: Pl. 14, 98, fig. 2), 1984.182 (O'RV: Pl. 14, 98, fig. 3). Io 5884 (O'LV: Pl. 14, 100, fig. 2), 1984.212 (o' copulatory appendage: Text-fig. 1).

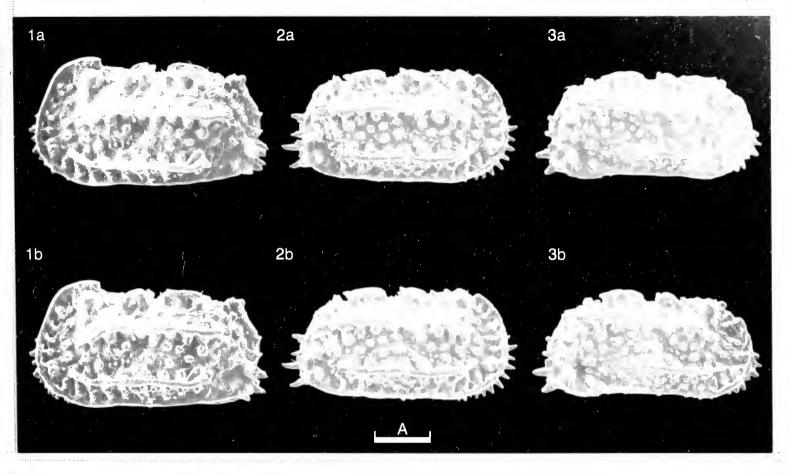
Institute of Palaeontology, University of Modena, no. 19252 (neotype, ♀ RV: Pl. 14,100,

1984.180 is from between Plymouth Sound and Start Point, Devon, SW England (lat. 50° 10′ N, long. 4° 00′ W), collected at depth of 38 m by S. J. Sturrock. 1984.181, 182 are from Unst Haaf (fishing grounds), Shetland (approx. lat. 61°00′N, long. 1°30′W), ex. Norman Collection slide no. 1900.3.6.268, collected 1867. Io 5884 is from Urla Bay, W Turkey (approx. lat. 38°19'N, long. 26°47′E), collected by N. Doruk (and figured by her (1981, op. cit.) as "C. antiquata (Baird)"). 1984.212 is from Rothesay Bay, Isle of Bute (approx. lat. 55°50'N, long. 5°05'W), SW Scotland, ex Brady Collection. All Recent. University of Modena no. 19252, from type locality; Late Pliocene.

Explanation of Plate 14, 100

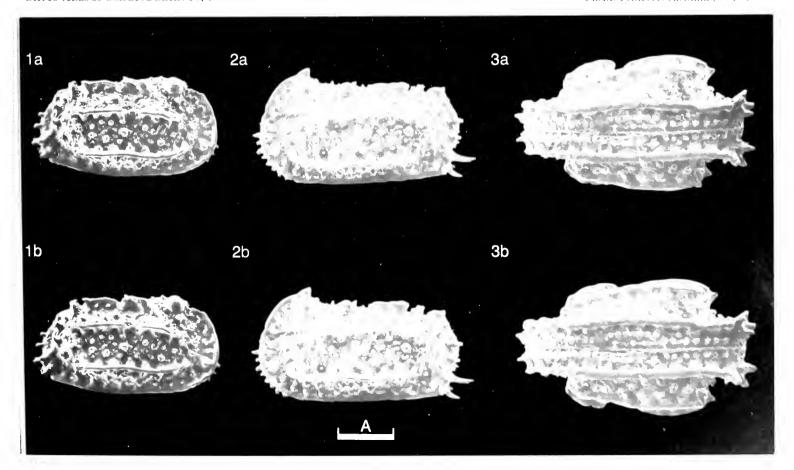
Fig. 1, Ω RV, ext. lat. (Neotype, Univ. of Modena no. 19252, 820 μm long); fig. 2, ♂ LV, ext. lat. (Io 5884, 960 μm long); fig. 3 Ω car., ext. vent. (1984.180).

Scale A (250 μ m; ×60), figs. 1–3.



Stereo-Atlas of Ostracod Shells 14, 100

Carinocythereis carmata (4 of 6)



Disgnosis:

Anterior margin with marginal carina which is entire and ponticulate throughout. Ventrolateral carina not produced anteriorly. Male copulatory appendages distinctive.

Remarks:

In designating a neotype for *Carinocythereis carinata*, Ruggieri & Russo (1980, *op. cit.*) chose a specimen identical to *C. antiquata* (Baird) in all aspects except for size. This was particularly unfortunate since Roemer's original illustration is so unclear as to make its interpretation entirely subjective, whereas *C. antiquata*, although lacking a type specimen, is readily determinable from Baird's original drawing. Nevertheless, *C. antiquata* and the neotype of *C. carinata* are, in our opinion, quite clearly conspecific (compare Pl. 14, 98, fig. 2 and Pl. 14, 100, fig. 1) and as a result the latter name takes priority.

C. whitei (Baird) (see J. Athersuch & J. E. Whittaker, Stereo-Atlas Ostracod Shells, 14, 103–110, 1987) differs from C. carinata principally in the disposition of the carinae. C. carinata possesses a marginal carina ventrally which extends without a break from the posteroventral angle around the anterior margin to the eye tubercle. C. whitei has a similar marginal carina which occupies the same position, but in contrast to C. carinata, it is replaced anteroventrally by a row of short, stout marginal spines. In addition, the carina does not form such a prominent crest above the eye tubercle. Both species possess ponticulate ventrolateral carinae. In C. whitei this carina extends anteriorly to run parallel to the anterior margin. A similar carina is found in C. carinata but it is restricted to the ventrolateral region, the anterior part being replaced by a row of four to five small tubercles. C. whitei is consistently smaller than C. carinata amongst living populations. Fossil specimens of C. carinata are also significantly smaller than their Recent counterparts. Minor differences in the male appendages are also useful in distinguishing between these two species.

In the juveniles (A-1, A-2) of both species, the marginal carinae are entire. However, differences seen in the ventrolateral carinae of the adults are also apparent in the juveniles. In addition, the juveniles of C. white i are proportionately less high and more tapered posteriorly then those of C. carinata.

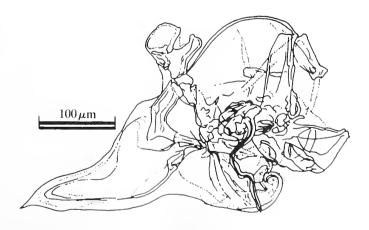
Stereo-Atlas of Ostracod Shells 14, 102

Carinocythereis carinata (6 of 6)

Distribution:

Recent: British coasts, most frequently in the north (BMNH and Brady Collection, Hancock Museum); French Atlantic Coast (Yassini, 1969, Bull. Inst. Géol. Bassin Aquitaine, 7); Mediterranean (Pugliese et al., 1978, Pubbl. Staz. zool. Napoli, 40); Black Sea (Caraion, op. cit.). Pliocene and Pleistocene: Mediterranean (Doruk, op. cit. and herein).

There is some evidence (e.g. Pugliese *et al.*, 1978), at least in the Mediterranean, that *C. carinata* (= *C. antiquata*) tends to occur more frequently in deeper water $(40-130 \,\mathrm{m})$ than *C. whitei* (= *C. bairdii*) $(20-90 \,\mathrm{m})$.



Text-fig. 1. O' copulatory appendage (1984.212). Drawing by D. J. Horne.

Stereo-Atlas of Ostracod Shells 14 (24) 103-110 (1987)

Carinocythereis whitei (1 of 8)

595.337.14 (119.1 + 119.9) (429:162.005.51 + 420:162.004.50 + 420:162.005.50 + 420:161.000.52 + 564.3:161.033.35 + 45:161.014.40):551.351

ON CARINOCYTHEREIS WHITEI (BAIRD)

by John Athersuch & John E. Whittaker (B.P. Research Centre, Sunbury and British Museum (Natural History), London)

Carinocythereis whitei (Baird, 1850)

- 1850 Cythereis Whitei sp. nov. W. Baird, The Natural History of British Entomostraca, Ray Soc., London, 175, pl. 20, figs. 3, 3a.
- 1865 Cythereis aspera sp. nov. G. S. Brady, Ann. Mag. nat. Hist., ser. 3, 16, 190, pl. 9, figs. 16-19.
- 1868 Cythere Whiteii (Baird); G. S. Brady, Trans. Linn. Soc. Lon., 26, 416, pl. 30, figs. 21-24.
- 1969 Carinocythereis bairdii sp. nov. F. Uliczny, Hemicytheridae und Trachyleberididae (Ostracoda) aus dem Pliozän der Insel Kephallinia (Westgrienchenland), Univ. of Munich, 79, pl. 5, fig. 7; pl. 16, fig. 7.
- 1971 Carinocythereis carinata (Roemer); P. Carbonel & J. Moyes, Revta esp. Micropaleont., 13, 147–154, pl. 1, figs. 1, 4; pl. 2, figs. 1–9 (non Cytherina carinata Roemer, 1838).
- 1976 Carinocythereis antiquata (Baird); G. Bonaduce, G. Ciampo & M. Masoli, Pubbl. Staz.zool.Napoli, 40, 49, pl. 25, figs. 8-10.
- 1985 Carinocythereis whitei (Baird); J. Athersuch, D. J. Horne & J. E. Whittaker, J. micropalaeontol., 4, 153–158, pl. 1, figs. 12–15; pl. 2, figs. 7, 8.
 - Lectotype: Designated herein, a female carapace from the Baird Collection, ex. slide no. 50.42; housed in the

Brit.Mus. (Nat.Hist.), London, cat. no. 1984.174 (now split into two valves).

Type locality: Tenby, Dyfed, SW Wales (lat. 51° 41'N, long. 4° 43'W); Recent.

Explanation of Plate 14, 104

Fig. 1, Q LV, ext. lat. (Lectotype, 1984.174, 860 μ m long); fig. 2, Q RV, ext. lat. (Lectotype, 1984.174, 840 μ m long); fig. 3, O RV, ext. lat. (1984.173, 890 μ m long). Scale A (250 μ m; \times 75), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 105

Carinocythereis whitei (3 of 8)

Figured specimens:

British Museum (Nat. Hist.) nos. **1984.174** (Lectotype, $\Q LV + RV$: Pl. **14**, 104, figs. 1, 2); **1984.173** ($\Q^2 RV$: Pl. **14**, 104, fig. 3); **OS 12312** ($\Q LV$: Pl. **14**, 106, fig. 1); **OS 12313** ($\Q RV$: Pl. **14**, 106, fig. 3); **1984.175** (juv. A-1 car.: Pl. **14**, 108, fig. 1); **1984.176** ($\Q RV$: Pl. **14**, 108, fig. 2); **1984.177** ($\Q RV$: Pl. **14**, 108, fig. 3); **1984.178** ($\Q^2 C$ car.: Pl. **14**, 110, figs. 1; copulatory appendage: Text-fig. 1a); **1984.179** ($\Q LV$: Pl. **14**, 110, figs. 2, 3); **1984.213** ($\Q^2 C$ copulatory appendages: Text-fig. 1b).

The lectotype (1984.174) is from the sole remaining syntypic slide in the Baird Collection (ex. 50.42) at the Brit. Mus. (Brit.Hist.); collected by T. R. Jones. 1984.173, 175 and 178 are from the Norman Collection (Brit.Mus. (Nat.Hist.)): 1984.173 and 175 from Dartmouth, Devon, SW England (lat. 50° 21′N, 3° 37′W) (ex slide no. 1911.11.8.M 3372); 1984.178 from Plymouth, Devon (approx. lat. 50° 22′N, 4° 08′W). 1984.176 and 1984.213 were collected alive by J. Athersuch from coarse sand in Famagusta Bay. Cyprus (approx. lat. 35° 10′N, long. 33° 58′E), water depth 30m, salinity 39.4‰, during November 1973. 1983.177 and 179, from the Bay of Naples (approx. lat. 40° 50′N, long. 14° 17′E), were kindly provided by Dr. G. Bonaduce. OS 12312–12314 are from the Nar Valley Clay, East Winch, Norfolk (lat. 00° 32′E, long. 52° 44′N), collected by P. G. Cambridge and B. M. Funnell; Pleistocene (Hoxnian?).

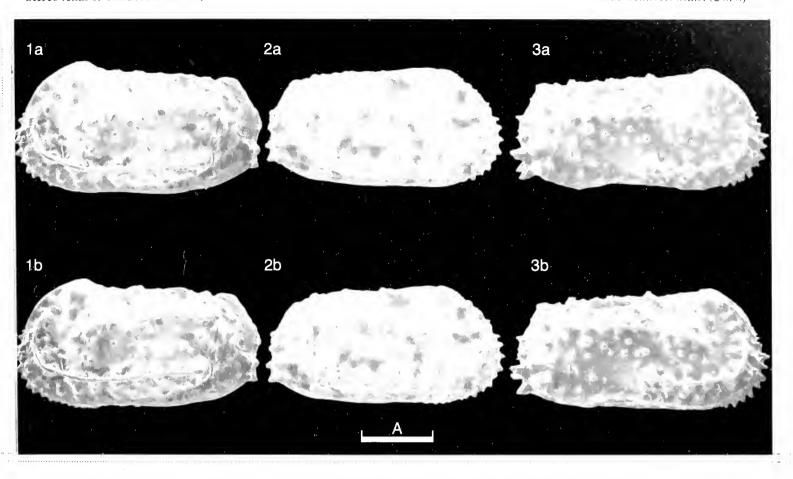
Diagnosis:

Anterior margin with carina which is entire and ponticulate dorsally, and disconnected ventrally to form a row of short spines. Ventrolateral carina extends to run parallel to anterior margin. Male copulatory appendage distinctive.

Explanation of Plate 14, 106

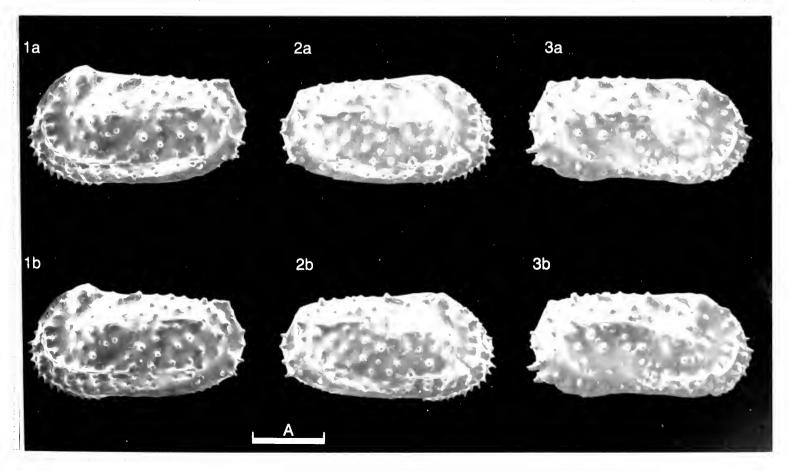
Fig. 1, Q LV, ext. lat. (OS 12312, 745 μ m long); fig. 2, Q RV, ext. lat. (OS 12313, 740 μ m long); fig. 3, Q RV, ext. lat. (OS 12314, 780 μ m long).

Scale A (250 μ m; × 75), figs. 1–3.



Stereo-Atlas of Ostracod Shells 14, 106

Carinocythereis whitei (4 of 8)



Remarks:

This species was recognised by Baird (1850, op. cit.), Brady (1868, op. cit.) and Brady, Crosskey & Robertson (1874, Palaentogr. Soc. Monogr.) as being distinct from C. carinata (Roemer, 1838) = C. antiquata (Baird, 1850). All of these authors, however, illustrated and described poorly preserved specimens of C. whitei, a fact that has tended to mask the true differences between these two species. (Worn specimens appear more nodose when the carinae are abraded). However, an examination of Baird's syntypes, one of which is illustrated herein (Pl. 14, 104, figs 1, 2) leaves us in no doubt as to the true identity of C. whitei. The main difference between C. whitei and C. carinata (Roemer) is in the length and disposition of the ventrolateral and anterolateral carinae (see also Remarks on C. carinata (Roemer) in J. Athersuch & J. E. Whittaker, Stereo-Atlas Ostracod Shells, 14, 97–102, 1987). There is some variation in the development of the carinae in both Recent and fossil forms (cf. Pl. 14, 106, figs. 2, 3; Pl. 14, 108, figs 2, 3), a factor which seems to be related to calcification of the carapace as a whole.

Until Athersuch, Horne & Whittaker (1985, op. cit.) reinstated the name C. whitei, G. S. Brady & A. M. Norman (Scient. Trans. R. Dubl. Soc., ser. 2, 4, 1889) were apparently the last authors to regard it as a distinct species in Britain and the only records under this name in the Mediterranean appear to be those of Ruggieri, 1956 (Att Soc.ital. Sci.nat., 95) and Uliczny, 1969 (op. cit.). Otherwise, the name whitei seems to have fallen into disuse and specimens referable to this species have usually been described as either C. antiquata (Baird) or C. bairdii Uliczny. Recent: British coasts (most frequently in the south), French Atlantic coast and widespread

Distribution:

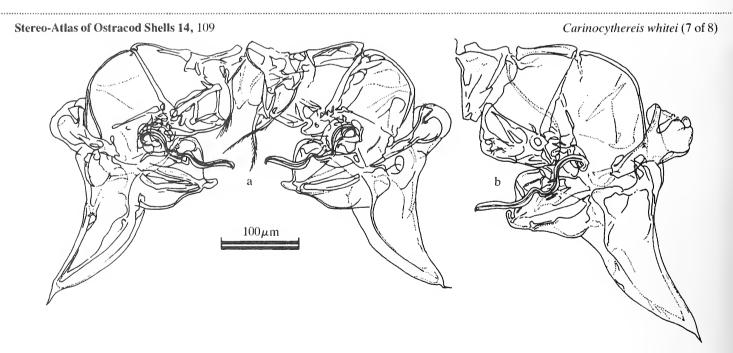
Recent: British coasts (most frequently in the south), French Atlantic coast and widespread throughout the Mediterranean (recorded as *C. antiquata* or *C. bairdii*). A sublittoral species found at depths of 20–60m or more.

Fossil: Pleistocene and Pliocene of the Mediterranean (under a variety of names) (Uliczny, op. cit.; Ruggieri, op.cit.); Pleistocene of England (as C. aspera).

Explanation of Plate 14, 108

Fig. 1, juv. A–1 car., ext. lt. lat. (1984.175, $700\mu m$ long); fig. 2, Q RV, ext. lat. (1984.176, $890\mu m$ long); fig. 3, Q RV, ext. lat. (1984.177, $780\mu m$ long).

Scale A (250 μ m; × 75), figs 1–3.



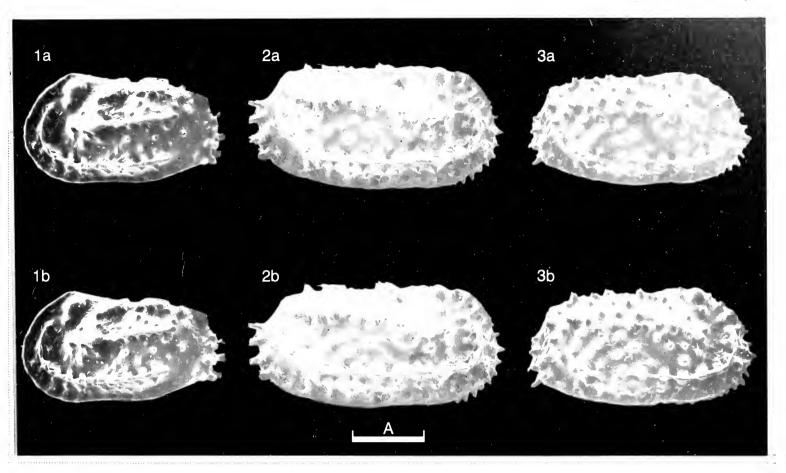
Text-fig. 1. of copulatory appendages: a, (1984.178), Recent of Britain; b, (1984.213), Recent of the Mediterranean. Drawings by D. J. Horne.

Explanation of Plate 14, 110

Fig. 1, \bigcirc car., ext. vent. (1984.178, 890 μ m long); fig. 2, \bigcirc LV, int. lat. (1984.179, 800 μ m long); fig. 3, \bigcirc LV, int. musc. sc. (1984.179).

Scale A (250 μ m; × 75), figs. 1,2; scale B (50 μ m; × 310), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 110



Carinocythereis whitei (8 of 8) 1a 2a 2b 1b

Stereo-Atlas of Ostracod Shells 14 (25) 111-114 (1987)

595.337.14 (119.15-118.21) (510 : 161.103.27) : 551.313.1 + 552.52

ON ABROTOCYTHERE QUINQUICORNIS ZHAO gen. et sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Genus ABROTOCYTHERE gen. nov.

Type-species: Abrotocythere quinquicornis sp. nov.

Derivation of name: Greek, meaning beautiful; in reference to its surface ornamentation + cythere.

Diagnosis: Genus small (adults $390-460\mu m$), subrectangular to truncated oval in side view, with well developed posteroventral cyathus in the right valve. Right valve hinge with narrow elongate tooth

anteriorly and large rounded tooth posteriorly separated by a groove which is very wide in its anterior half and disappears under the dorsal margin posteriorly. Muscle scar pattern a vertical

row of four adductor scars with rounded mandibular and frontal scars.

Remarks: Appendages and soft parts are unknown but the nature of the adductor muscle scar pattern places

the new genus in the Cytheracea. The general carapace features place it in the Limnocytheridae and it can be assigned to the Subfamily Timiriaseviinae. Here it has affinities with the Kovalovskiella (Rosacythere) and Theriosynoecum groups of Colin & Danielopol (Palaeobiologie Continentale XI, 1, 13-17, 1980). Although the hinge structure is very similar, it differs from Theriosynoecum in the general nature of the ornamentation as well as in shape, which in Abrotocythere is much more rectangular and less rounded, particularly posteriorly. It is closest to the Cretaceous genus Rosacythere Colin, 1980 from which it differs most markedly in having the positive elements of the hinge structure in the right valve. In size $(390-460 \mu m)$ it is also much smaller than both Rosacythere (560–600 μ m) and Theriosynoecum (620–1420 μ m). Abrotocythere may be regarded as a Tertiary derivative of Rosacythere and may thus belong in the Kovalovskiella

group.

Explanation of Plate 14, 112

Fig. 1, 2, RV, (holotype, 103070, 390 μ m long): fig. 1, ext. lat., fig. 2, ext. dors. Scale A (100 μ m; × 245), figs. 1, 2.

Stereo-Atlas of Ostracod Shells 14, 113

Abrotocythere quinquicornis sp.nov.

Abrotocythere quinquicornis (3 of 4)

Holotype:

Academia Sinica Nanjing Institute of Geology and Palaeontology, China; coll. no. 103070, RV. [Paratypes: eight valves, Academia Sinica Nanjing Institute of Geology and Palaeontology

nos. 103071, 90870-90876].

Type locality:

Section at Gaocanzi, Zhongshui town, Weining County, Guizhou province, SW China; lat. 27°

Derivation of name: Figured specimens:

20'N, long. 103° 39'E. From a marl lens in mudstones of Miocene (or possibly Oligocene) age. Latin; reference to the five horn-like spines or pore conuli in the posterior half of the shell. Academia Sinica Nanjing Institute of Geology and Palaeontology, nos. 103070 (holotype, RV: Pl. 14, 112, figs. 1, 2), 103071 (paratype, RV: Pl. 14, 114, figs. 1, 2). Both figured specimens are from

the type locality and horizon.

Diagnosis:

A small $(390\mu m)$ species of *Abrotocythere*, subrectanglar in side view with strong infracurvature, the anterodorsal margin sloping gently at about 45° to the vertical. Straight dorsal and ventral margins are parallel, slightly concave in their median part and truncated posteriorly by the vertical posterior margin. Left valve slightly larger than right valve with ventral overlap medianly and a well developed posteroventral cyathus in the right valve. Wide, shallow "V" shaped sulcus anterodorsally giving a dorsal view reminiscent of a calabash. Primary puncta pentgonal or subrounded with secondary pitting. Pore canal openings are clearly visible at the top of the pore conuli and on the ridges of the reticulation anteriorly. Inner lamella narrow with very small vestibules at each end, selvage strong. Hinge typical of genus. A vertical row of four adductor scars lies on a platform in front of the strongly vaulted posterior part of the shell and these scars are also seen on the external surface. There are two oval mandibular scars anteroventrally and a round frontal scar level with the topmost adductor scar.

Remarks:

A. quinquicornis differs from A. ovata Zhao (Stereo-Atlas Ostracod Shells, 14, (26) 115-118, 1987) in its smaller size and in the development of five prominent tubercles/spines posteriorly.

Distribution:

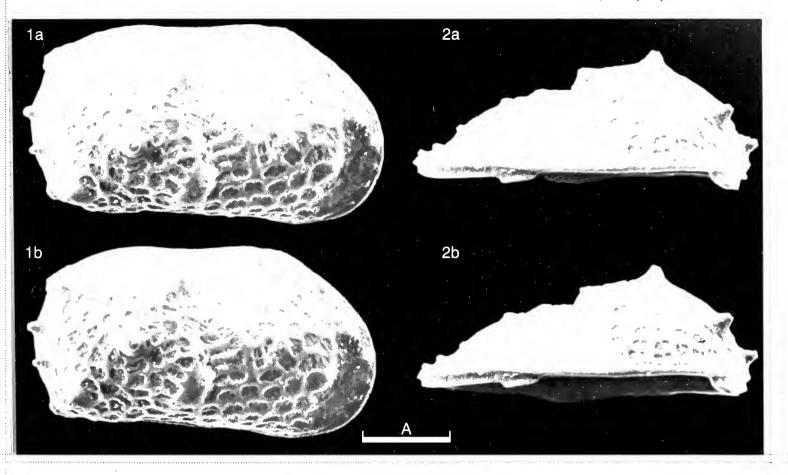
Abrotocythere quinquicornis and Abrotocythere ovata Zhao have been found in the Guizhou Province, SW China in beds of (?) Oligocene-Miocene age; they are associated with gastropods

which are thought to occupy an oligohaline niche.

Acknowledgment:

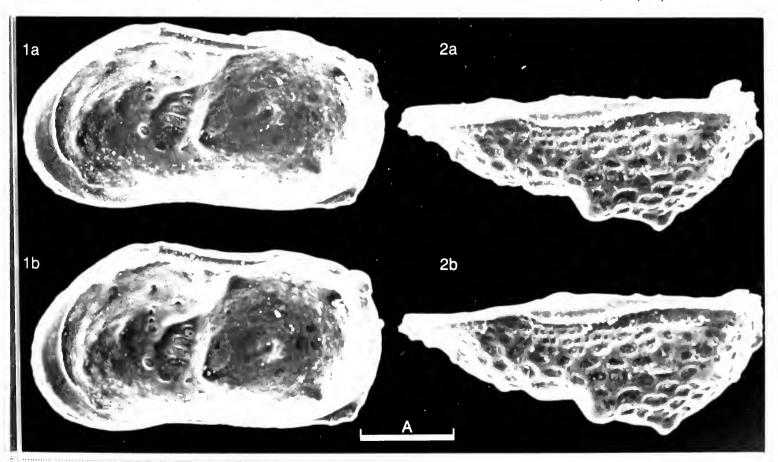
This study was undertaken while a visiting Research Scholar in the Department of Geology, University of Hull, England.

Explanation of Plate 14, 114



Stereo-Atlas of Ostracod Shells 14, 114

Abrotocythere quinquicornis (4 of 4)



ON ABROTOCYTHERE OVATA ZHAO sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Abrotocythere ovata sp. nov.

Holotype: Academia Sinica Nanjing Institute of Geology and Palaeontology, China, coll. no. 103072; RV.

[Paratypes: two valves, Academia Sinica Nanjing Institute of Geology and Palaeontology,

nos. 103073(1)-103074(2)].

Type locality: Section at Gaokanzi, Zhongshui town, Weining county, Guizhou province, SW China, lat. 27°

20'N, 103° 39'E. From a marl lens in mudstones of Miocene (or possibly Oligocene) age.

Derivation of name: Referring to the oval outline of the shell.

Figured specimens: Academia Sinica Nanjing Institute of Geology and Palaeontology nos. 103072 (holotype, RV: Pl.

14, 116, fig. 1; Pl. 14, 118, fig. 2), 103073(1) (paratype, RV: Pl. 14, 118, fig. 1), 103073(2) (paratype, LV: Pl. 14, 116, fig. 2). All of the figured specimens are from the type locality and

horizon.

Diagnosis: Small, truncated oval in lateral view, cordate in dorsal view. Dorsal and ventral margins

subparallel. Right valve with strong posteroventral cyathus. Wide, shallow V-shaped sulcus anterodorsally. Surface with subrounded fossae with secondary pitting, in some areas arranged in the manner of fish scales. Size of fossae variable becoming smaller anterodorsally near the shallow V-shaped sulcus. The elongate, round, posteroventral tubercle lies slightly behind mid-length and is inclined downwards anteriorly at an angle of about 30° to the horizontal. Inner lamella narrow

Explanation of Plate 14, 116

Fig. 1, RV, ext. lat. (holotype, 103072, 460 μ m long); fig. 2, LV, ext. lat. (paratype, 103073(2), 445 μ m long). Scale A (100 μ m; × 205), figs. 1; scale B (100 μ m; × 177), fig. 2.

Stereo-Atlas of Ostracod Shells 14, 117

Abrotocythere ovata (3 of 4)

Diagnosis: (cont.)

with very small vestibule anteriorly. Hinge characteristic of the genus with elongate anterior tooth plate, large rounded posterior tooth and connecting groove in the right valve. A vertical row of four adductor scars lies on a platform in front of the swollen posterior part of the shell. There are two small rounded and closed mandibular scars anteroventrally.

Remarks:

This species occurs with Abrotocythere quinquicornis Zhao (Stereo-Atlas Ostracod Shells, 14, 111, 1987) to which it is obviously closely related. It differs in a number of important respects. With a length generally $450-460\mu m$, A.ovata is consistently larger than A.quinquicornis (390 μm). Ornamentation also differs consistently. The present species, whilst showing the basic elements of sulcus, reticulation and tuberculation differs in a number of important respects. A.ovata lacks the five prominent tubercles/spines of A.quinquicornis. In this it might simply be considered a morph of the latter species but for the fact that the tubercle that is developed is elongated in a direction virtually at right angles to the direction the one which occurs in roughly the same position in A.quinquicornis. They can not be regarded as homologous and the pattern of fossae round these respective tubercles is also quite different. Similarly these same differences suggest that this is not a case of sexual dimorphism and the current taxon is regarded as a species different from, but co-eval with, A.quinquicornis.

A.ovata is somewhat similar in outline to Kovalevskiella phreaticola (Danielopol) (Colin and Danielopol, Paleobiologie Continentale, XI, 1, pl. 2, fig. 1, 2, 1980) and other Kovalevskiella species (Colin and Danielopol, Paleobio, Contin., XI, 1, 13–14, pl. 1–5, 1980), but there are clear

differences in ornamentation, hinge structure and muscle scar pattern.

Distribution:

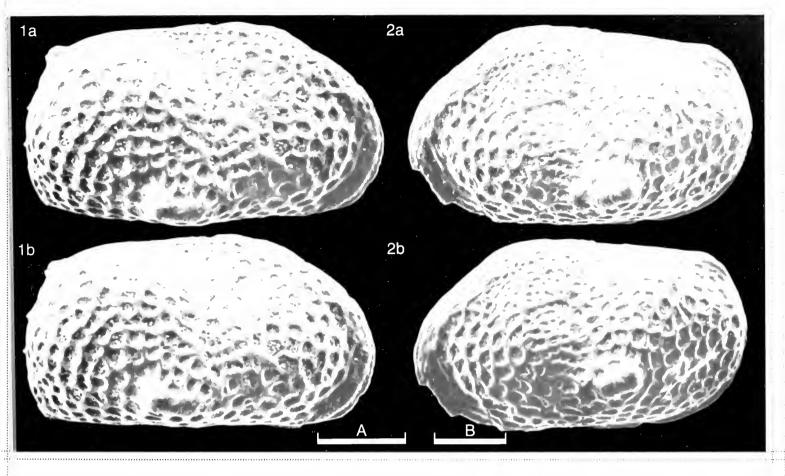
Abrotocythere ovata and Abrotocythere quinquicornis (Stereo-Atlas Ostracod Shells, 14, 111, 1987) have been found in Guizhou Province, SW China, in beds of Miocene (or possibly Oligocene) age. They are associated with gastropods which are thought to have lived in an oligohaline habitat.

Acknowledgment:

This study was undertaken while a visiting Research scholar in the Department of Geology, University of Hull, England.

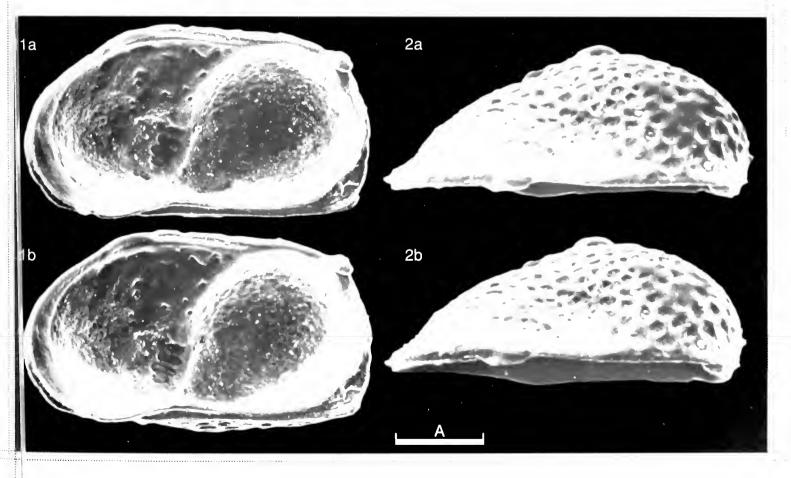
Explanation of Plate 14, 118

Fig. 1, RV int. lat. (paratype, 103073(1), 450 μ m long); fig. 2, RV ext. dors. (holotype, 103072, 460 μ m long). Scale A (100 μ m; × 212), figs. 1, 2.



Stereo-Atlas of Ostracod Shells 14, 118

Abrotocythere ovata (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (27) 119-122 (1987) 595.337.14 (119.1) (510 : 161.104.26) : 551.312.4 + 552.52

ON LEUCOCYTHERE WEININGENSIS ZHAO sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Leucocythere weiningensis sp. nov.

Holotype: Academia Sinica Nanjing Institute of Geology and Palaeontology, China, coll. no. 103064;

carapace.

[Paratypes: valve and carapace, Academia Sinica Nanjing Institute of Geology and

Palaeontology, nos. 103065-103066].

Type locality: Borehole CK-17 at Caohai Lake, Weining County, Guizhou Province, SW China; lat. 26° 51'N,

104° 12'E. At a depth of 21m from the surface; black mudstones of Pleistocene age.

Derivation of name: From its occurrence in the Weining County, Guizhou Province, SW China.

Figured specimens: Academia Sinica Nanjing Institute of Geology and Palaeontology nos. 103064 (holotype, car.: Pl.

14, 120, figs. 1, 2), 103065 (paratype, RV: Pl. 14, 122, fig. 1.), 103066 (paratype, car.: Pl. 14, 122,

fig. 2). All of the figured specimens are from the type locality and horizon.

Explanation of Plate 14, 120

Figs. 1, 2, car. (holotype, 103064, 470 μ m long): fig. 1, ext. lt. lat.; fig. 2, ext. rt. lat. Scale A (100 μ m; × 201), figs. 1, 2.

Stereo-Atlas of Ostracod Shells 14, 121

Leucocythere weiningensis (3 of 4)

Diagnosis:

Carapace small; dorsal margin straight, inclined towards posterior; ventral slightly concave in the median part. Highest and widest about one-third length from the anterior end. Surface reticulate with secondary pitting in the fossae. Two vertical dorsal sulci in the anterior half of the shell, the most prominent lying just in front of mid-length. Pores occur at the intersections of some muri of which two or three anteriorly, and about half a dozen posteriorly form fairly prominent pore conuli. There is a small backward projecting spine at about three-quarters length and at about one-fifth the height above the ventral margin. Some specimens show a sulcus immediately behind this spine (Pl. 14, 120, fig. 2). Vertical row of four adductor scars placed low on the shell on the anterior side of the internal ridge, with two rounded mandibular scars more ventrally. Hinge merodont with straight toothplate subdivided into three toothlets anteriorly, a locellate groove and a prominent elliptical tooth posteriorly in the right valve.

Remarks:

This species is related to Leucocythere plena Zhao (see Stereo-Atlas Ostracod Shells, 14, 123, 1987), but the latter is more swollen posteriorly, with a concave posterior outline in dorsal view,

and the hinge structure is less well developed.

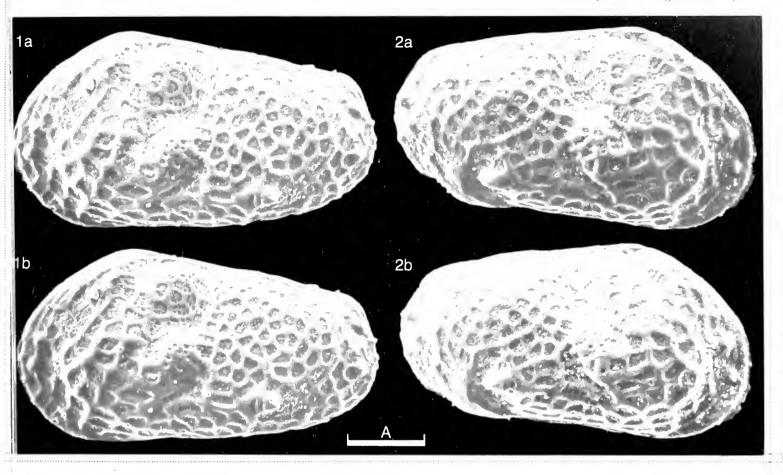
Distribution: Acknowledgement:

This species has so far only been found in Pleistocene deposits in Guizhou Province, SW China. This study was undertaken as a visiting research scholar at the Department of Geology, University

of Hull, England.

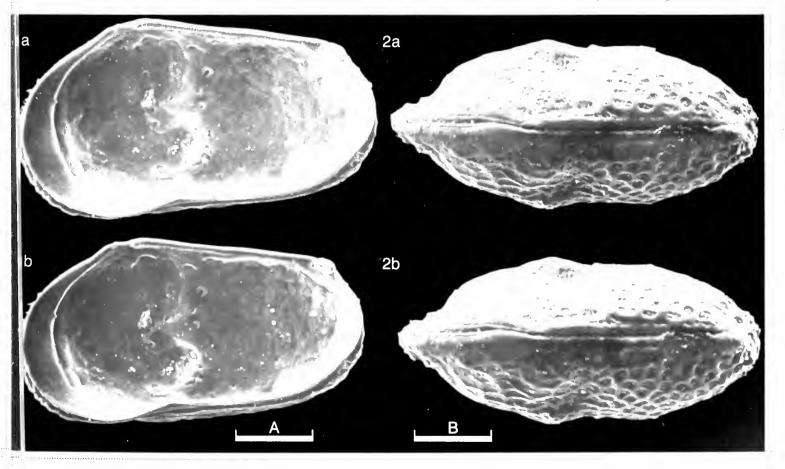
Explanation of Plate 14, 122

Fig. 1, RV int. lat. (paratype, 103065, 450 μ m long); fig. 2, car., ext. dors. (paratype, 103066, 470 μ m long). Scale A (100 μ m; × 210), fig. 1; scale B (100 μ m; × 205), fig. 2.



itereo-Atlas of Ostracod Shells 14, 122

Leucocythere weiningensis (4 of 4)



595.337.14 (119.1) (510 : 161.104.26) : 551.312.4 + 552.52

ON LEUCOCYTHERE PLENA ZHAO sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Leucocythere plena sp. nov.

Holotype: Academia Sinica Nanjing Institute of Geology and Palaeontology, China, coll. no. 103067;

carapace.

[Paratypes: valve and carapace, Academia Sinica Nanjing Institute of Geology and

Palaeontology, nos. 103068-103069].

Type locality: Borehole CK-17 at Caohai Lake, Weining County, Guizhou Province, SW China; lat. 26° 51'N,

104° 12′E. At a depth of 21m from the surface; black mudstones of Pleistocene age.

Derivation of name: From the latin plenus, plump, stout; in reference to the swollen posterior half of the shell. Figured specimens: Academia Sinica Nanjing Institute of Geology and Palaeontology nos. 103067 (holotype, car.: Pl.

14, 124, figs. 1, 2), 103068 (paratype, RV: Pl. 14, 126, fig. 1.), 103069 (paratype, car.: Pl. 14, 126,

fig. 2). All of the figured specimens are from the type locality and horizon.

Explanation of Plate 14, 124

Figs. 1, 2, car. (holotype, **103067**, 520 μ m long): fig. 1, ext. lt. lat.; fig. 2, ext. rt. lat. Scale A (100 μ m; × 178), figs. 1, 2.

Stereo-Atlas of Ostracod Shells 14, 125

Leucocythere plena (3 of 4)

Diagnosis: Small to medium sized carapace with gently concave dorsum inclined posteriorly. Highest anteriorly at about one-third the length. Two dorsal vertical sulci in the anterior half of the shell;

sub-central tubercle; posterior half of shell swollen. Sparse pore conuli developed over the surface of the shell and developed posteriorly where they form distinct tubercles. Ornamentation of subdued reticulation with round secondary pits occupying the fossae. Ten marginal pore canals anteriorly. Vertical row of four adductor scars in the ventral part of the shell and lying on the anterior flank of the median internal ridge which defines the posterior limit of the sub-central tubercle. Hinge merodont with narrow, well-defined anterior and posterior toothplates with thin,

sinuous groove in between.

Remarks: This species is closely related to L. weiningensis Zhao (see Stereo-Atlas Ostracod Shells, 14, 119,

1987) but differs in its concave dorsum, less differentiated hinge, subdued ornamentation and marked posterior swelling. The present species is also related to *L. subquadrata* Huang & You, 1982 (Huang, Yang & You, *Palaeontology of Xizang*, Book IV, 377, fig. 6, pl. 14, fig. 3, 1982, Beijing), but that species lacks the carapace sulci and does not show the posterior inflation of

L.plena.

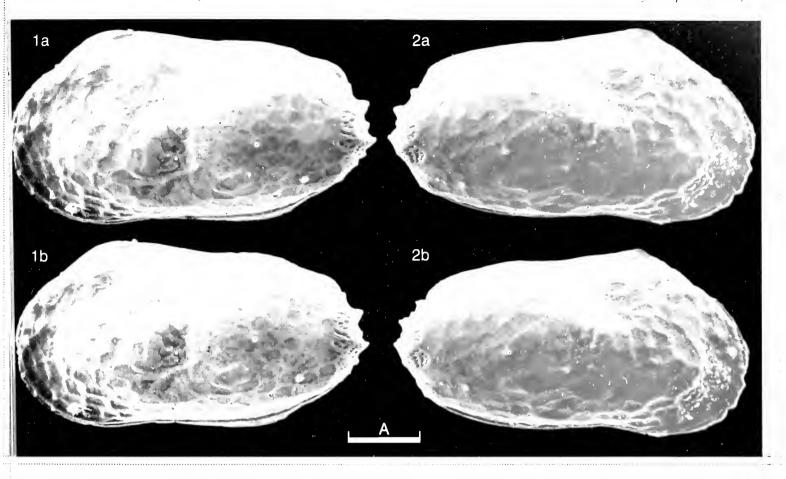
Distribution: L.plena has so far only been found in Pleistocene deposits in Guizhou Province, SW China.

Acknowledgement: This study was undertaken as a visiting Research Scholar at the Department of Geology,

University of Hull, England.

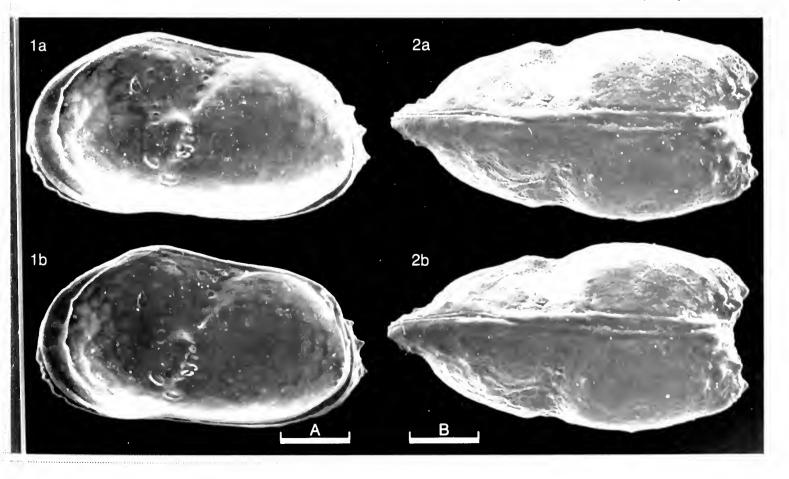
Explanation of Plate 14, 126

Fig. 1, RV int. lat. (paratype, 103068, 490 μ m long); fig. 2, car., ext. dors. (paratype, 103069, 520 μ m long). Scale A (100 μ m; × 188), fig. 1; scale B (100 μ m; × 186), fig. 2.



Stereo-Atlas of Ostracod Shells 14, 126

Leucocythere plena (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (29) 127-130 (1987)

595.337.14 (119.1) (510 : 161.104.26) : 551.312.4 + 552.52

ON LIMNOCYTHERE XINANENSIS ZHAO sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Limnocythere xinanensis sp. nov.

Holotype: Academia Sinica Nanjing Institute of Geology and Palaeontology, China, coll. no. 103060; ♀ LV.

[Paratypes: three female valves and carapaces, Academia Sinica Nanjing Institute of

Geology and Palaeontology, nos. 103061-103063].

Type locality: Borehole CK-17 at Caohai Lake, Weining County, Guizhou Province, SW China; lat. 26° 51'N,

104° 12'E. At a depth of 17m from the surface; black shale of Pleistocene age.

Derivation of name: From its occurrence in the Xinan region of China.

Figured specimens: Nanjing Institute of Geology and Palaeontology nos. 103060 (holotype, Q LV: Pl. 14, 128, fig. 1),

10361 (paratype, \bigcirc RV: Pl. 14, 128, fig. 2), 10362 (paratype, \bigcirc RV: Pl. 14, 130, fig. 1), 103063 (paratype, \bigcirc car.: Pl. 14, 130, fig. 2). All of the figured specimens are from the type locality and

horizon.

Diagnosis: Shell reniform but more broadly rounded in front, dorsal side straight, ventral side curved. Median

and anterior dorsal, vertical sulci occur, of which the median is the stronger. Surface ornamentation of five nodes and primary and secondary reticulation. Two nodes lie in the dorsal half of the shell on either side of the median sulcus. The other nodes lie in the ventral half of the shell, one behind the median sulcus, the other two smaller nodes lie one above the other in front of the median sulcus. Hinge merodont with terminal undivided toothplates linked by a groove in the

Explanation of Plate 14, 128

Fig. 1, \bigcirc LV, ext. lat. (holotype, **103060**, 480 μ m long); fig. 2, \bigcirc RV, ext. lat. (paratype, **103061**, 490 μ m long). Scale A (100 μ m; \times 187), figs. 1, 2.

Stereo-Atlas of Ostracod Shells 14, 129

Limnocythere xinanensis (3 of 4)

Diagnosis: (cont.)

right valve. Row of four closely pressed adductor muscle scars centrally placed on the ridge which represents the expression of the median sulcus internally, frontal scar oval on the same level as the two uppermost adductors and two rounded mandibular scars more ventrally placed. In dorsal view pointed anteriorly and somewhat arrow-shaped. Three small dorsal spines in the posterior half of

the right valve.

Remarks:

L.xinanensis is closely related to L.stationis Vávra, 1891 but differs clearly from the latter species as originally figured (Archiv Naturw. Landesd. Böhmens, 8, 109, fig. 38, 1891) in being much more slender and more pointed posteriorly in dorsal view and in tapering more posteriorly and not being so evenly rounded anteriorly in lateral view. L.stationis was thought to be confined to Europe until Martens (Hydrobiologia, 110, 138–141, figs. 9–16, 1984) recorded it from the Sudan and gave good illustrations, Martens' material is much nearer to the Chinese material in dorsal view but in lateral aspect does not taper so much posteriorly and the dorsal margin shows a more pronounced break in slope than does L.xinanensis where the dorsal margin is long and straight. Martens notes the variability of dorsal spines in L.stationis where up to three may be found although they were completely absent from his African specimens. L.xinanensis shows a similar variability in the development of these spines. L.xinanensis from the lower part of the present section consisted of many females and rare males neither of which carried dorsal spines. In the middle of the section specimens with one dorsal spine were found and both males and females were present, whilst in the upper part of the section many males and females occurred which had three dorsal spines.

De Deckker's Australian species *L.dorsosicula* (*Proc. R. Soc. Vict.*, **93**, 43–45, figs. 1, 2a–i, 1981) has between three and six spines dorsally and also differs in its much reduced turberculation.

Comparisons with other species are not close.

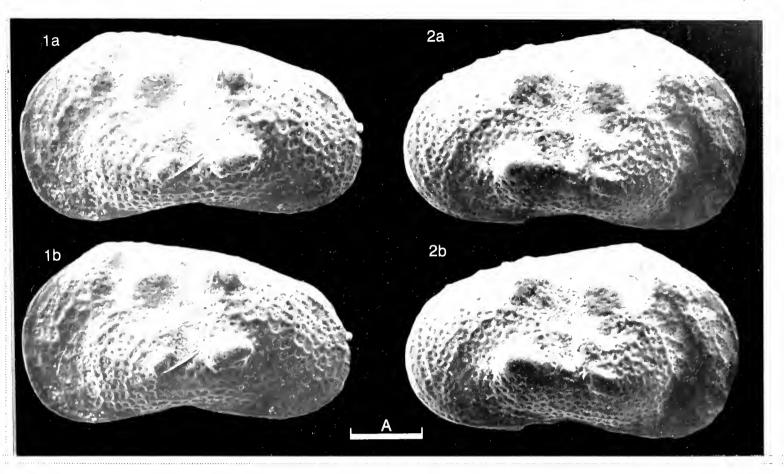
Distribution: Limnocythere xinanensis has been found in the Guizhou Yunnan Province in China in deposits

ranging from Pleistocene to Recent in age.

Acknowledgement: This study was undertaken as a visiting Research Scholar at the Department of Geology, University of Hull, England.

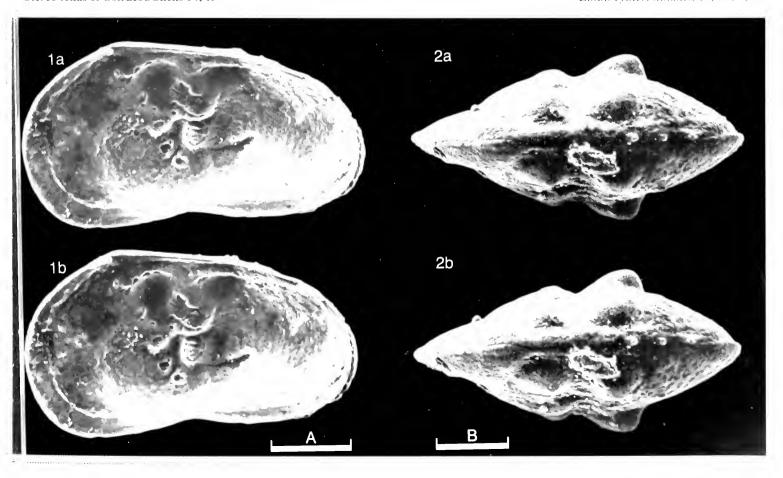
Explanation of Plate 14, 130

Fig. 1, Q RV, int. lat. (paratype, **103062**, 440 μ m long); fig. 2, Q car. ext. dors. (paratype, **103063**, 480 μ m long). Scale A (100 μ m; \times 207), fig. 1; scale B (100 μ m; \times 183), fig. 2.



Stereo-Atlas of Ostracod Shells 14, 130

Limnocythere xinanensis (4 of 4)



Stereo-Atlas of Ostracod Shells 14 (30) 131-134 (1987) 595.337.14 (119.1) (510 : 161.104.26) : 551.312.4 + 552.52

ON METACYPRIS APHTHOSA ZHAO sp. nov.

by Zhao Yuhong

(Academia Sinica Nanjing Institute of Geology and Palaeontology, China & University of Hull, England)

Metacypris aphthosa sp. nov.

Holotype: Academia Sinica Nanjing Institute of Geology and Palaeontology, China, coll. no. 103074(a); Q

carapace.

[Paratypes: two male valves, one female carapace and one male carapace, Academia Sinica

Nanjing Institute of Geology and Palaeontology, nos. 103074(b)-103074(e)].

Type locality: Borehole at Caohai Lake, Weining County, Guizhou Province, SW China; lat. 26° 51'N, 104°

12'E. At a depth of 27m from the surface; black mudstones of Pleistocene age (Q2-3).

Derivation of name: Greek aphthosus, measles; in reference to the surface ornamentation.

Figured specimens: Academia Sinica Nanjing Institute of Geology and Palaeontology nos. 103074(a) (holotype, Q

car.; RV: Pl. 14, 132, figs. 1, 3), 10374(b) (paratype, ♀ car: Pl. 14, 132, fig. 2), 103074(c) (paratype, ♂ RV: Pl. 14, 134, fig. 1), 103074(e) (paratype, ♂ car: Pl. 14, 134, fig. 2), 103074(d) (paratype, ♂ RV: Pl. 14, 134, fig. 1). All of the figured specimens are from the type locality and

horizon.

Diagnosis: Distinct sexual dimorphism. Females medium-sized, rounded-rectangular in lateral view and cordate in dorsal view with the greatest width posteriorly. Males small, elongate in lateral view and

oval in dorsal view with the greatest width posteriorly. Males small, elongate in lateral view and oval in dorsal view with the greatest width at mid-length. Larger left valve overlaps right valve. Surface reticulate. Females have a very faint trace of a dorsal sulcus which is not seen in any of the males. Two to four rows of very fine pits occur marginally and are well seen along the dorsal

Explanation of Plate 14, 132

Figs. 1, 3, \bigcirc car., RV (holotype, 103074(a), 480 μ m long): fig. 1, ext. lat., fig. 3, int. lat. Fig. 2, \bigcirc car. (paratype, 103074(b), 480 μ m long).

Scale A (200 μ m; × 134), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 133

Metacypris aphthosa (3 of 4)

margins of the valves in dorsal view (Pl. 14, 132, fig. 2: Pl. 14, 134, fig. 2). Each valve develops five tubercles anteriorly and four or five posteriorly. These are constant in position and there is no difference between the sexes. Hinge merodont, right valve with a long, smooth anterior toothplate, a shorter, thicker, smooth posterior toothplate and slightly sinuous interconnecting groove. Right valve free margin with strong selvage and posteroventrally the valve bulges down well below the valve margin. Typical cytheracean muscle scar pattern with a row of four adductor scars, the outer two oval, the central two very elongated. Two small, rounded mandibular scars occur anteroventrally.

Remarks:

Distribution:

This species is very similar to *Metacypris changzhouensis* Chen (*Acta Palaeon. Sinica*, 13(1), 7, pl. 2, figs. 9, 13, 1965) but in the latter the tubercles lack the constancy and regular distribution seen in *M.aphthosa. M.changzhouensis* differs further in that the right valve is larger than the left valve, and in addition it is also a bigger species (length 680 µm). Differences are also apparent in dorsal view when the posterior part of the carapace is compared. It also differs from *Metacypris unibulla* Hou & Chen (*Acta Palaeon. Sinica*, 13(1), p. 9, pl. 1, figs. 5, 9 1965) because the latter only has one posterior tubercle, is thinner in dorsal view and differs in size amongst other things. The present species differs from *Metacypris cordata* Brady & Roberston (Brady and Roberston, *Ann. Mag. nat. Hist.*, Ser. 4. 6, 19–20, pl. VI, figs. 1, 9, 1870; Pinto & Sanguinetti, *Esc. Geol. P. Alegre*, 4, pl. II, figs. 1 a–e, 1962; Colin & Danielopol, *Palaebiologie Continetale*, XI, 1, 29–30, pl. 14, figs. 5–9, 1980), in that in the latter the right valve is the larger, there is no surface tuberculation and it is longer and narrower than *Metacypris aphthosa* which is a short and very inflated species.

Males, females and younger instars are all found together in the deposits examined although the females are more than twice as abundant as the males and instars together.

M. aphthosa has so far only been found in Pleistocene deposits in Guizhou Province, SW China.

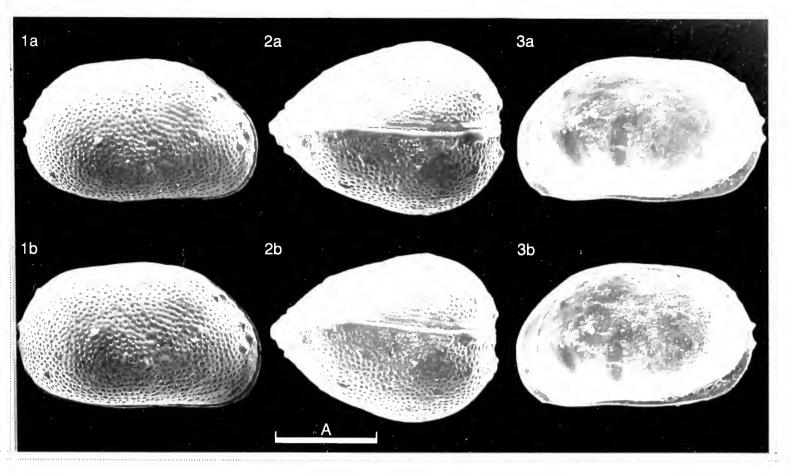
Acknowledgement: This study was undertaken as a visiting Research Scholar at the Department of Geology, University of Hull, England.

Explanation of Plate 14, 134

Fig. 1, O RV, ext. lat. (paratype, 103074(e), 440 μ long); fig. 2, O car., dors (paratype, 103074(e), 400 μm long); fig. 3, O RV, int. lat. (paratype, 103074(d), 420 μm long). Scale A (200 μm; × 146), figs. 1–3.

Stereo-Atlas of Ostracod Shells 14, 134

Metacypris aphthosa (4 of 4)



1a 2a 3a

1b 2b 3b

595.337.14 (116.333) (669 : 161.001.06) : 551.35

ON BENINEA IBECETENENSIS APOSTOLESCU gen. et sp. nov.

by Vespasian Apostolescu (5, rue J. -C. Bézanier, 78360-Montesson, France)

Genus BENINEA gen. nov.

Type-species: Beninea ibecetenensis sp. nov.

Derivation of name:

from Benin, W Africa.

Diagnosis:

Cytheridae essentially characterized by its hinge. Right valve: anterior element consisting of a strong, rounded tooth, a long crenulate groove and a posterior plate-like cardinal element bearing five strong crenulations; left valve: large anterior socket, long crenulate ridge slightly arched and a posterior strongly crenulate socket. No accommodation groove.

Carapace subovoid in side view, elongate ovate dorsally. Anterior margin broadly rounded, posterior margin obliquely rounded. Left valve larger than right; dorsal margin regularly arched, with greatest height in middle part. Anterior margin of right valve more angular. Surface smooth with well developed normal sieve-type pore-canals. Eye tubercle absent. Sexual dimorphism pronounced; males more elongate than females.

Central muscle scars: vertical row of four coalescent rounded scars and two equally rounded scars in front (Text-fig. 1).

Narrow marginal zone; line of concrescence coincides with the inner margin. Radial pores

straight, simple and up to eight on anterior margin (Text-fig. 1). Remarks:

Externally, Beninea is comparable to Bopaina Apostolescu, 1961 and "Clithrocytheridea" senegali Apostolescu, 1961 from the Senonian of Senegal (Rev. Inst. franç Pétrole, 16, (7–8), 779–867). Except for the absence of an accommodation groove and the median ridge on the left valve, the hinge of Beninea is close to Apatocythere Triebel, 1940 (Senckenbergiana, 22, (3/4), 160–227),

Explanation of Plate 14, 136

Fig. 1, \circlearrowleft car., rt. lat. (paratype, P-351, 560 μ m long); fig. 2, \circlearrowleft car., rt. lat. (holotype, H-350, 510 μ m long); fig. 3, \circlearrowleft car., lt. lat. (holotype, H-350, $510\mu m$ long).

Scale A (200 μ m; × 110), fig. 1; scale B (200 μ m; × 120), fig. 2; scale C (200 μ m; × 130), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 137

Remarks: (cont.)

Dordoniella Apostolescu, 1955 (Cah. géol., 33, 329-330), and Schulapacythere Malz, 1970 (Senckenbergiana, 51, (5/6), 401–409). In external view, Beninea differs from these genera by the shape of the carapace and the absence of an eye tubercle.

Beninea ibecetenensis sp. nov.

Holotype:

V. Apostolescu Collection, Lab. Micropaleontol., Mus. natl. Hist. nat., Paris, France, no. H-350; Q carapace.

[Paratypes: 12 carapaces and valves; same repository as holotype].

Type locality:

Ibeceten borehole (at 181-182m), near the town of Anthieme, Nigeria, Benin, W Africa (see Apostlescu, 1961, Rev. Inst. franç Pétrole, 16 (7-8), tab. 3, 786); early Senonian, Cretaceous.

Derivation of name:

Figured specimens:

From the bore-hole Ibeceten, the type locality. Mus. natl. Hist. nat. Paris, France, V. Apostolescu Collection, no. H−350 (holotype, ♀ car.: Pl. 14, 136, figs. 2, 3), P-351 (paratype, o' car.: Pl. 14, 136, fig. 1; Pl. 14, 138, fig. 1), P-352 (paratype, ♀ LV: Pl. 14, 138, fig. 2), P-353 (paratype, ♀ RV: Pl. 14, 138, fig. 3). All from the type-locality, Ibeceten borehole (at 181–182m), Benin, W Africa. Early Senonian, occurring

together with other ostracodes such as Cophinia apiformis (Reyment, 1960).

Diagnosis: Distribution: As for the genus.

Early Senonian, Cretaceous, of the Benin-Togo basin, W Africa.

Acknowledgment:

Dr. J. P. Colin, Esso Production Research-European Lab. (Bégles) is thanked for providing the S.E.M. micrographs (taken by C. Lété) and for reading the text.





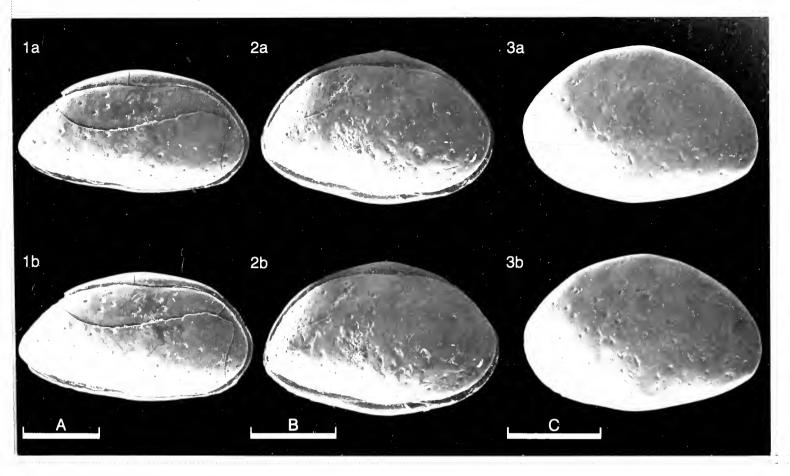


Text-fig. 1. B. ibecetenensis: a, internal view, left valve; b, internal view, right valve; c, dorsal view, right valve.

Explanation of Plate 14, 138

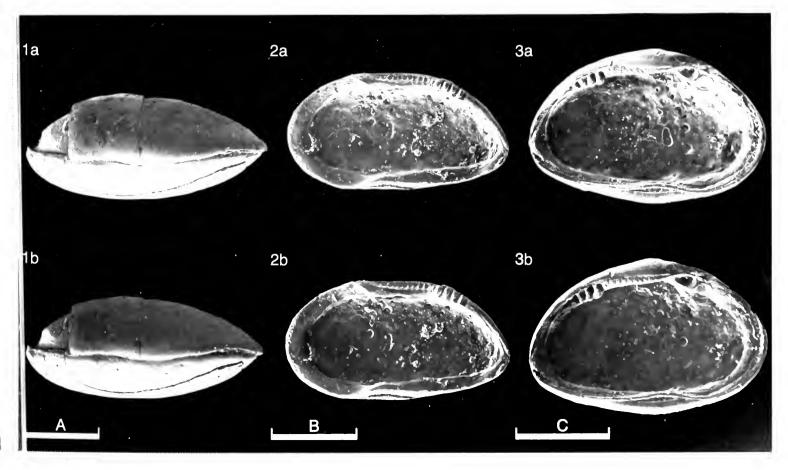
Fig. 1, \circlearrowleft car., ext. dors. (paratype, P-351, 560 μ m long); fig. 2, \circlearrowleft RV, int. lat. (paratype, P-352, 480 μ m long); fig. 3, \circlearrowleft LV, int. lat. (paratype, P-353, $490\mu m \log$).

Scale A (200 μ m; × 110), fig. 1; scale B (200 μ m; × 120), fig. 2; scale C (200 μ m; × 130), fig. 3.



Stereo-Atlas of Ostracod Shells 14, 138

Beninea ibecetenensis (4 of 4).



595,337,3 (113.51) (420 : 162.003.55 + 411 : 162.004.56) : 551.351

ON GLYPTOLICHVINELLA SPIRALIS (JONES & KIRKBY)

by Robert F. Lundin

(Arizona State University, Tempe, U.S.A.)

Genus GLYPTOLICHVINELLA Pozner, 1966

Type-species (by original designation): Kirkbya spiralis Jones & Kirkby, 1884

Cytherellacean genus the lateral surfaces of which are ornamented with two ridges, one which is Diagnosis:

subparallel to the lateral outline and may or may not continue to form a marginal ridge, and another which is median and bends below the adductorial sulcus. Straguloid process variably

developed. Domatium with variable number of separate egg compartments.

Remarks: The earliest valid publication of this genus known to me is that of K. Ya. Gurevich (in Fossil Ostracoda, O. S. Vyalov, ed., Acad. Sci. Ukr. SSR, Inst. Geol. & Geochem. Fossil Fuels, 1966 =

Israel Program for Scientific Translations, 1971 English translation of Russian original). In that publication, Pozner is credited with authorship of the genus and the generic name is spelled Glyptolichvinella rather than Glyptolichwinella as it appears in various other literature.

Accordingly, the former spelling is used here.

Lichvinella scopinensis Pozner (op. cit.) is the type-species for Lichvinella. M. N. Gramm (Vladivostok) has provided me with two photographs of L. scopinensis, one of which shows that the females of that species have egg compartments and a limen. The discovery of egg compartments in Glyptolichvinella indicates, therefore, that this genus differs from Lichvinella only by the presence of a separate longitudinal ridge on the lateral surface of each valve. I judge this to be only a species-level difference but do not formally synonymize the two genera until more and better material of each can be studied.

Explanation of Plate 14, 140

Fig. 1, ♂ car., ext. lt. lat (BMNH I 1719, [pars], 1030μm long); fig. 2, ♀ car., ext. rt. lat., light photograph to show egg compartments (BMNH I 1719, [pars], 880 μ m long); fig. 3. Q car., ext. lt. lat. (BMNH OS 7384, 1240 μ m long). Scale A (200 μ m; × 75), fig. 1; scale B (200 μ m; × 84), fig. 2; scale C (200 μ m; × 61), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 141 Glyptolichvinella spiralis (Jones & Kirkby, 1884) Glyptolichvinella spiralis (3 of 4)

1884 Kirkbya spiralis sp. nov.; T. R. Jones & J. W. Kirkby, Berwickshire Nat. Club, Hist., 10, (1882–1884), 323, pl. 2, figs. 12, 13.

1885 Kirkbya spiralis, Jones & Kirkby; T. R. Jones & J. W. Kirkby, Ann. Mag. nat. Hist., ser. 5, 15, 184, pl. 3, fig. 11. 1978 Glyptolichvinella spiralis (Jones & Kirkby, 1884); E. Robinson, in Bate, R. H. & Robinson, J. E. (eds.), A Stratigraphical

Index of British Ostracoda, Geol. J. Spec. Issue, 8, 138, pl. 5, fig. 4, table 2.

Type specimens: Apparently are lost. British Museum (Nat. Hist.) I 2554, identified (slide information) as primary

types of Kirkbya spiralis Jones & Kirkby and "Leperditia subrecta, Portlock", consist of two rock chips with many leperditiid specimens but none of Kirkbya spiralis. Under present knowledge of

the species, it is premature to designate a neotype.

Type locality: Lower Carboniferous (Dinantian) limestone on the coast near Randerstone, Fifeshire, Scotland. Figured specimens: British Museum (Nat. Hist.), OS 7384 (♀ car.: Pl. 14, 140, fig. 3), I 1719 (pars) (♂ car.: Pl. 14,

140, fig. 1), **I 1719** (*pars*) (♀ car.: Pl. **14**, 140, fig. 2; Pl. **14**, 142, figs. 1, 2). **OS 7384** is from Megg's Linn (Lower Asbian, Dinantian), Lewisburn, North Tyne, Northumberland, England; approx. lat. 55° 10'N, long. 2° 20'W. Specimens I 1719 are from Lower Carboniferous (Dinantian)

"Calcareous Sandstone Series, at Linnhouse Water, Linlithgowshire", Scotland; approx. lat. 56° N, long. 3° 40′ W. I 1719 contains six carapaces (two figured herein).

Diagnosis: Glyptolichvinella species with spiral ridge along entire margin which at midlength of dorsum runs

anteroventrally and then parallels anterior, ventral, posterior and posterodorsal margins, terminating just behind the adductorial sulcus. Separate longitudinal ridge on lateral surface bends below adductorial sulcus. Anterior straguloid process weak. Adult females with five (perhaps

more or less) oval to circular egg compartments in each valve. Surface granulose.

Remarks: Along with the report of Lundin & Visintainer (Stereo-Atlas Ostracod Shells, 14 (33), 143–148,

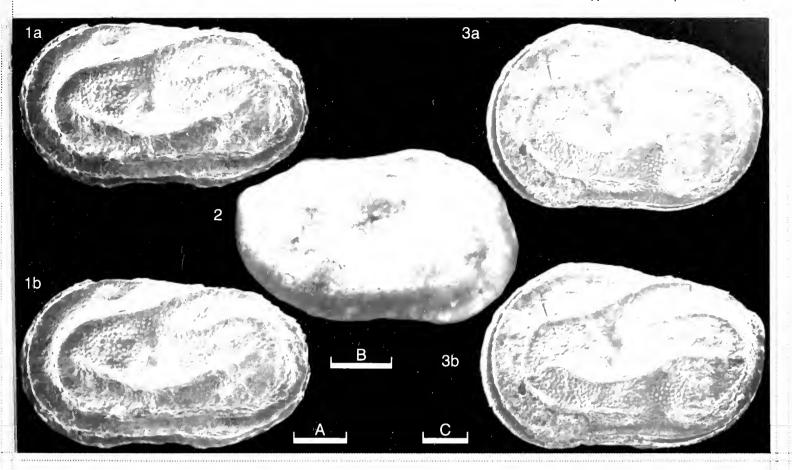
1987) on G. ovicella, this is the first report of egg compartments in the domatium of females of this genus. Only 7 specimens (carapaces) of G. spiralis have been available to me. All are damaged and the 3 illustrated here provide impressions of the species only in the lateral views shown.

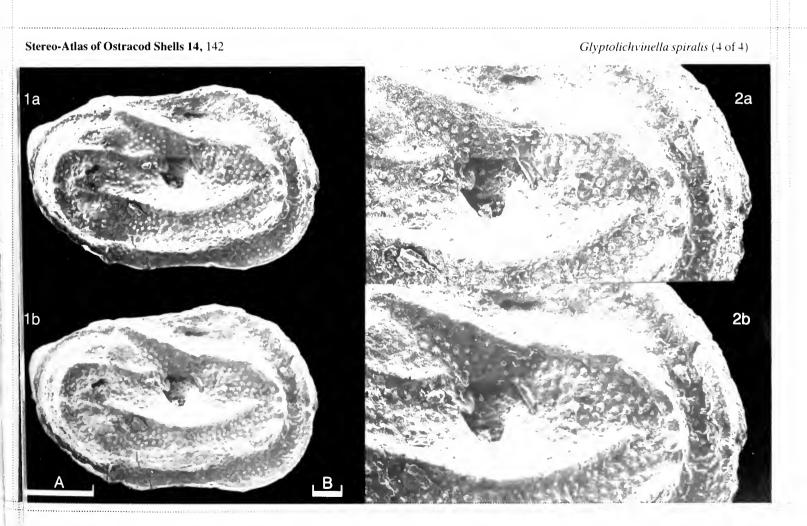
Distribution: Known from Lower Carboniferous (Viséan) of Scotland and England (see Robinson, 1978, op.

cit.) Reported also from the Lower Carboniferous of the USSR (see Gurevich, 1966, op. cit.). Acknowledgement: Support from College Liberal Arts and Sciences, Arizona State Univ. is gratefully acknowledged.

Explanation of Plate 14, 142

Fig. 1, 2, Q car. (BMNH I 1719, [pars], 800μ m long); fig. 1, ext. rt. lat.; fig. 2 ext. rt. lat. (median and mid-anterior areas). Scale A (200 μ m; × 88), fig. 1; scale B (50 μ m; × 164), fig. 2.





Stereo-Atlas of Ostracod Shells 14 (33) 143-148 (**1987**) 595.337.3 (113.45) (941 : 163.127.21) : 551.351

ON GLYPTOLICHVINELLA OVICELLA LUNDIN & VISINTAINER sp. nov.

by Robert F. Lundin & Linda M. Visintainer (Arizona State University, Tempe, U.S.A.)

Glyptolichvinella ovicella sp. nov.

Holotype:

Department of Geology, Arizona State University (ASU), no. ASU X-91; Q car.

Type locality:

White Hill no. 1 borehole, Canning Basin, Western Australia; latitude 21° 9′ 20.35″S, longitude 127° 35′ 14.98″E. Holotype from interval 1520–30m below top of borehole in rocks of probable Famennian age, Devonian. Other figured and studied specimens from samples ranging from

1080-2890m below top of borehole.

Derivation of name: Diagnosis:

Latin ovum, egg, and cella, chamber; referring to the presence of egg compartments.

Glyptolichvinella species with one ridge paralleling the margin and a longitudinal ridge approximately at midheight which bends below S_2 . Females with distinct domatium having three to

six separate egg compartments in each valve.

Figured specimens:

Explanation of Plate 14, 144

Fig. 1, \bigcirc car., ext. lt. lat. (holotype, ASU X–91, 600μ m long); fig. 2, \bigcirc car., ext. lt. lat. (ASU X–97, 730μ m long); fig. 3, \bigcirc car., ext. rt. lat. (ASU X–93, 660μ m long).

Scale A (200 μ m; × 86), fig. 1; scale B (200 μ m; × 76), fig. 2; scale C (200 μ m; × 84), fig. 3.

Stereo-Atlas of Ostracod Shells 14, 145

Glyptolichvinella ovicella (3 of 6)

Remarks:

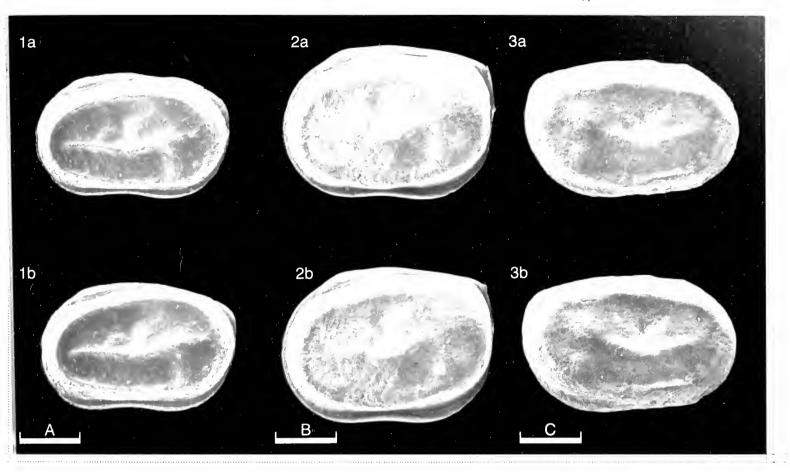
Glyptolichvinella ovicella is readily distinguished from G. spiralis (= Kirkbya spiralis Jones & Kirkby, 1884; see Jones & Kirkby, Ann. Mag. nat. Hist., 5, 15, 184, 1885) by differences in the lateral ridges, and from G. nodosovidera Crasquin, 1983 (see Crasquin, Ann Soc. Géol. Nord. CII, 191–204, 1983) by differences in the lateral ridges and by the absence of eye tubercles.

This is the first report of separate compartments to house eggs for this genus. Lundin (see Stereo-Atlas of Ostracod Shells, 14 (32), 139–142, 1987) reports similar egg compartments in the type-species, G. spiralis (Jones & Kirkby, 1884) and we conclude, therefore, that this is a generic character which needs to be verified in other species of the genus. The number of egg compartments per valve varies from three to six and no carapace studied has more than eleven or fewer than six. Normally the number of egg compartments in each valve of a carapace is equal, but in some cases the right valve has one more compartment than the left valve. There is no systematic change in the number of egg compartments per specimen through the 1810m-interval from which the studied specimens were derived.

Except for one adult tecnomorphic left valve and one juvenile tecnomorphic right valve, all specimens studied are complete carapaces. We cannot, therefore, definitively demonstrate the existence of a limen in the females. An exterior depression at the anteroventral edge of the domatium suggests a limen is present. The presence of a well-developed anterior straguloid process and the morphology of the contact margin and hinge of the two isolated valves available for study further indicate that *Glyptolichvinella* is a typical platycope ostracode.

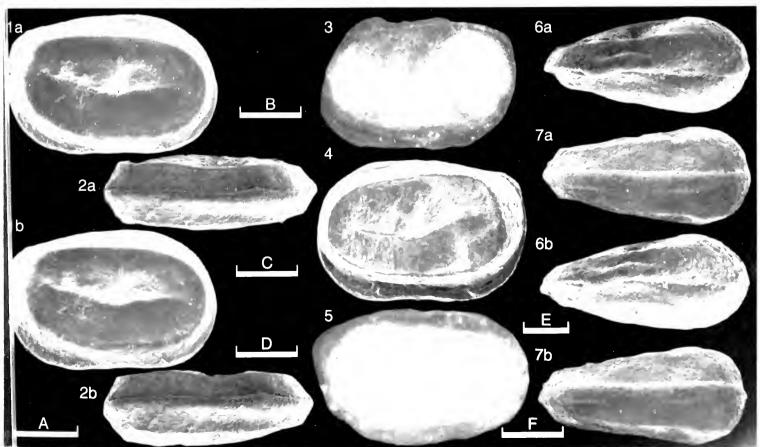
Explanation of Plate 14, 146

Scale A (200 μ m; × 96), figs. 1, 2; scale B (200 μ m; × 81), fig. 3; scale C (200 μ m × 87), fig. 4; scale D (200 μ m; × 86), fig. 5; scale E (200 μ m; × 64), fig. 6; scale F (200 μ m; × 91), fig. 7.



Stereo-Atlas of Ostracod Shells 14, 146

Glyptolichvinella ovicella (4 of 6)



-	
1	
12	
-	

Stereo-Atlas of Ostracod Shells 14, 147

Glyptolichvinella ovicella (5 of 6)

Distribution: Known only from the type locality. The stratigraphic interval containing this species is certainly, in

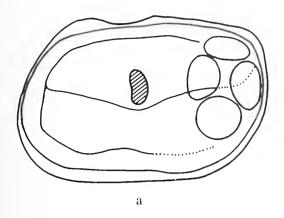
part (if not entirely), late Devonian (Frasnian and/or Famennian) but it possibly ranges into the

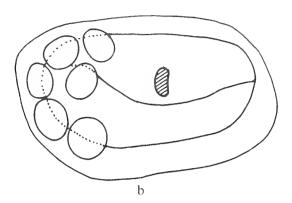
early Carboniferous.

Acknowledgments: We gratefully acknowledge the help of Lee B. Gibson, David Ford, Mobil Exploration &

Producing Services, Inc. and the support of the College of Liberal Arts and Sciences, Arizona

State University.

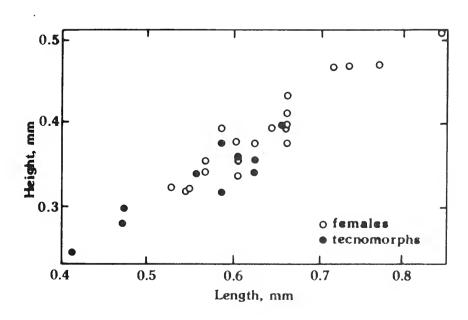




Text-fig. 1 Drawings to show ornamentation and position of egg compartments in *G.ovicella*: a, specimen ASU X-100 (Pl. 14, 146, fig. 3); b, specimen ASU X-99 (Pl. 14, 146, fig. 5).

Stereo-Atlas of Ostracod Shells 14, 148

Glyptolichvinella ovicella (6 of 6)



Text-fig. 2 Size dispersion diagram of thirty specimens of *G.ovicella* from nine stratigraphic intervals in White Hill no. 1 borehole. Western Australia.



reticulata, Cathaycythere; 1–4 robusta, Kuiperiana; 73–76

General Index

Abrotocythere ovata Zhao sp. nov.; 115–118 Abrotocythere quinquicornis Zhao gen. et sp. nov.; 111–114 alaefortis alaefortis, Sagmatocythere; 85–88 alaefortis gallica, Sagmatocythere; 89-92 Albileberis sinensis Hou; 9-12 aphthosa, Metacypris; 131–134 Apostolescu, V., On Beninea ibecetenensis; 135–138 aremorica, Healdianella?; 25–28 Athersuch, J. & Whittaker, J. E., On *Carinocythereis carinata* (Roemer); 97–102 Athersuch, J. & Whittaker, J. E., On *Carinocythereis whitei* (Baird); 103–110 Babinot, J. F. & Colin, J. P., On Spinoleberis eximia (Bosquet); 37-40 Beninea ibecetenensis Apostolescu gen. et sp. nov.; 135-138 Beyrichia (Sagenabeyrichia) siveteri Pollicott subgen. et sp. nov.; 57-64 Brouwers, E. M., On *Pterygocythereis vannieuwenhuisei* Brouwers sp. nov.; 17–20 *Bythocythere intermedia* Elofson; 65–68 Bythocythere zetlandica Athersuch, Horne & Whittaker; 69–72 Calocaria maurae Vannier gen. et sp. nov.; 45-48 camptocytheroidea, Howeina; 33–36 Carbonel, P., Colin, J. P. & Londeix, L., On Kovalevskiella caudata (Lutz); 41–44 carinata, Carinocythereis; 97–102 Carinocythereis carinata (Roemer); 97–102 Carinocythereis whitei (Baird); 103–110 Cathaycythere reticulatà Whatley & Zhao gen. et sp. nov.; 1-4 Cathaycythere reticulata Whatley & Zhao gen. et sp. nov.; 1–4
caudata, Kovalevskiella; 41–44
Coles, G. P. & Cronin, T. M., On Muellerina hazeli Coles & Cronin sp. nov.; 21–24
Colin, J. P. & Babinot, J. F., On Spinoleberis eximia (Bosquet); 37–40
Colin, J. P., Carbonel, P. & Londeix, L., On Kovalevskiella caudata (Lutz); 41–44
Compton-Gooding, E. & Ikeya, N., On Howeina camptocytheroidea Hanai; 33–36 Crasquin, S., On *Healdianella? aremorica* Crasquin sp. nov.; 25–28 Cronin, T. M. & Coles, G. P., On *Muellerina hazeli* Coles & Cronin sp. nov.; 21–24 esurialis, Spinohippula; 49-56 eximia, Spinoleberis; 37-40 Glyptolichvinella ovicella Lundin & Visintainer sp. nov.; 43-148 Glyptolichvinella spiralis (Jones & Kirkby); 139-142 hazeli, Muellerina; 21-24 Healdianella? aremorica Crasquin sp. nov.; 25–28 Horne, D. J., On Bythocythere intermedia Elofson; 65–68 Horne, D. J., On Bythocythere zetlandica Athersuch, Horne & Whittaker; 69-72 Howeina camptocytheroidea Hanai; 33-36 ibecetenensis, Beninea; 135–138 Ikeya, N. & Compton-Gooding, E., On Howeina camptocytheroidea Hanai; 33–36 impressa, Sinocytheridea; 13-16 intermedia, Bythocythere; 65-68 Kovalevskiella caudata (Lutz); 41–44 Krüta, M., Vannier, J. & Marek, L., On Spinohippula esurialis Vannier, Krüta & Marek gen. et sp. nov.; 49–56 Kuiperiana robusta Whatley & Maybury sp. nov.; 73–76 Leucocythere plena Zhao sp. nov.; 123–126
Leucocythere weiningensis Zhao sp. nov.; 119–122
Limnocythere xinanensis Zhao sp. nov.; 127–130
Londeix, L., Carbonel, P. & Colin, J. P., On Kovalevskiella caudata (Lutz); 41–44
Loxocauda subquadrata Maybury & Whatley sp. nov; 77–80
Lundin, R. F., On Glyptolichvinella spiralis (Jones & Kirkby); 139–142
Lundin, R. F. & Visintainer, L. M., On Glyptolichvinella ovicella Lundin & Visintainer sp. nov.; 43–148 Maghrebeis tuberculata Majoran gen. et sp. nov.; 29–32 Majoran, S., On Maghrebeis tuberculata Majoran gen. et sp. nov.; 29–32 Marek, L., Vannier, J. & Krůta, M., On Spinohippula esurialis Vannier, Krůta & Marek gen. et sp. nov.; 49–56 maurae, Calocaria; 45–48 Maybury, C. & Whatley, R. C., On Kuiperiana robusta Whatley & Maybury sp. nov.; 73–76
Maybury, C. & Whatley, R. C., On Loxocauda subquadrata Maybury & Whatley sp. nov.; 77–80
Maybury, C. & Whatley, R. C., On Sagnatocythere alaefortis alaefortis Whatley & Maybury sp. nov.; 85–88
Maybury, C. & Whatley, R. C., On Sagnatocythere alaefortis gallica Whatley & Maybury sp. nov.; 89–92
Maybury, C. & Whatley, R. C., On Sagnatocythere minuta Maybury & Whatley sp. nov.; 81–84
Maybury, C. & Whatley, R. C., On Sagnatocythere wyatti Maybury & Whatley sp. nov.; 93–96
Metacypris aphthosa Zhao sp. nov.; 131–134
minuta Sagnatocythere: 81–84 minuta, Sagmatocythere; 81–84 Muellerina hazeli Coles & Cronin sp. nov.; 21-24 ovata, Abrotocythere; 115-118 plena, Leucocythere; 123–126 Pollicott, P. D., On Beyrichia (Sagenabeyrichia) siveteri Pollicott subgen. et sp. nov.; 57-64 Pterygocythereis vannieuwenhuisei Brouwers sp. nov.; 17-20 quinquicornis, Abrotocythere; 111-114

Sagmatocythere alaefortis alaefortis Whatley & Maybury sp. nov.; 85–88 Sagmatocythere alaefortis gallica Whatley & Maybury sp. nov.; 89–92 Sagmatocythere minuta Maybury & Whatley sp. nov.; 81–84 Sagmatocythere wyatti Maybury & Whatley sp. nov.; 93-96 sinensis, Albileberis; 9-12 sinensis, Sinocythere; 5-8 Sinocythere sinensis Hou; 5-8 Sinocytheridea impressa (Brady); 13-16 Sinotymertaea impressa (Blady), 13–16 siveteri, Beyrichia (Sagenabeyrichia); 57–64 Spinolippula esurialis Vannier, Kruta & Marek gen. et sp. nov.; 49–56 Spinoleberis eximia (Bosquet); 37–40 spiralis, Glyptolichvinella; 139–142 subquadrata, Loxocauda; 77–80 tuberculata, Maghrebeis; 29-32 Vannier, J., On Calocaria maurae Vannier gen. et sp. nov.; 45-48 Vannier, J., Krůta, M. & Marek, L., On *Spinohippula esurialis* Vannier, Krůta & Marek gen. et sp. nov.; 49–56 Visintainer, L. M. & Lundin, R. F., On *Glyptolichvinella ovicella* Lundin & Visintainer sp. nov.; 43–148 *vannieuwenheuisei*, *Pterygocythereis*; 17–20 weiningensis, Leucocythere; 119-122 Weiningensis, Leucocythere; 119–122
Whatley, R. C. & Maybury, C., On Kuiperiana robusta Whatley & Maybury sp. nov.; 73–76
Whatley, R. C. & Maybury, C., On Loxocauda subquadrata Maybury & Whatley sp. nov.; 77–80
Whatley, R. C. & Maybury, C., On Sagmatocythere alaefortis alaefortis Whatley & Maybury sp. nov.; 85–88
Whatley, R. C. & Maybury, C., On Sagmatocythere alaefortis gallica Whatley & Maybury sp. nov.; 89–92
Whatley, R. C. & Maybury, C., On Sagmatocythere minuta Maybury & Whatley sp. nov.; 81–84
Whatley, R. C. & Maybury, C., On Sagmatocythere wyatti Maybury & Whatley sp. nov.; 93–95
Whatley, R. C. & Zhao, Q., On Albileberis sinensis Hou; 9–12
Whatley, R. C. & Zhao, Q., On Cathaycythere reticulata Whatley & Zhao gen. et sp. nov.; 1–4
Whatley, R. C. & Zhao, Q., On Sinocythere sinensis Hou; 5–8
Whatley, R. C. & Zhao, Q., On Sinocytheridea impressa (Brady); 13–16
whitei. Carinocythereis: 103–110 whitei, Carinocythereis; 103-110 Whittaker, J. E. & Athersuch, J., On Carinocythereis carinata (Roemer); 97-102 Whittaker, J. E. & Athersuch, J., On Carinocythereis whitei (Baird); 103-110 wyatti, Sagmatocythere; 93-96 xinanensis, Limnocythere; 127–130 zetlandica, Bythocythere; 69-72 Zhao, Q. & Whatley, R. C., On Albileberis sinensis Hou; 9-12 Zhao, Q. & Whatley, R. C., On Cathaycythere reticulata Whatley & Zhao gen. et sp. nov.; 1-4 Zhao, Q. & Whatley, R. C., On Sinocythere sinensis Hou; 5-8 Zhao, Q. & Whatley, R. C., On Sinocytheridea impressa (Brady); 13-16 Zhao, Y., On Abrotocythere ovata Zhao sp. nov.; 115–118
Zhao, Y., On Abrotocythere quinquicornis Zhao gen. et sp. nov.; 111–114
Zhao, Y., On Leucocythere plena Zhao sp. nov.; 123–126
Zhao, Y., On Leucocythere weiningensis Zhao sp. nov.; 119–122
Zhao, Y., On Limnocythere xinanensis; 127–130 Zhao, Y., On Metacypris aphthosa; 131-134

Index; Geological Horizon

	See 1 (2) 5-22 (1973) for explanation of the	Schedules in the	Universal Decimal Classification
(113.312)	Middle Ordovician:	(118.22)	Pliocene:
	Spinohippula esurialis; 49–56		Carinocythereis carinata; 97–102
(113.331)	Lower Silurian:		Howeina camptocytheroidea; 33–36
(442.222)	Beyrichia (Sagenabeyrichia) siveteri; 57–64		Kuiperiana robusta; 73–76
(113.333)	Upper Silurian:		Loxocauda subquadrata; 77–80
(112.45)	Calocaria maurae; 45–48 Devonian:		Pterygocythereis vannieuwenhuisei; 17–20
(113.45)	Glyptolichvinella ovicella; 43–148		Sagmatocythere alaefortis alaefortis; 85–88 Sagmatocythere alaefortis gallica; 89–92
(113.51)	Lower Carboniferous:		Sagmatocythere minuta; 81–84
(113.31)	Glyptolichvinella spiralis; 139–142		Saginatocythere wyatti; 93–96
	Healdianella? aremorica; 25–28		Sinocytheridea impressa; 13–16
(116.331)	Cenomanian:	(119)	Quaternary:
,	Maghrebeis tuberculata; 29–32		Albileberis sinensis; 9–12
(116.333)	Senonian:	(119.1)	Pleistocene:
(44 (222 2)	Beninea ibecetenensis; 135–138		Carinocythereis whitei; 103–110
(116.333.3)	Maastrichtian:		Leucocythere plena; 123-126
(110.15)	Spinoleberis eximia; 37–40		Leucocythere weiningensis; 119–122
(118.15)	Oligocene:		Limnocythere xinanensis; 127–130
	Abrotocythere ovata; 115–118 Abrotocythere quinquicornis; 111–114		Metacypris apluthosa; 131–134 Muellerina hazeli; 21–24
(118.21)	Miocene:	(119.9)	Recent:
(110.21)	Abrotocytliere ovata; 115–118	(117.7)	Albileberis sinensis; 9–12
	Abrotocythere quinquicornis; 111–114		Bythocythere intermedia; 65–68
	Kovalevskiella caudata; 41–44		Bythocythere zetlandica; 69-72
			Carinocythereis carinata; 97–102
			Carinocythereis whitei; 103-110
			Cathaycythere reticulata; 1–4
			Howeina camptocytheroidea; 33–36
			Muellerina hazeli; 21–24
			Sinocythere sinensis; 5–8
			Sinocytheridea impressa; 13–16

Index; Geographical Location

	See 1 (2) 5–22 (1973) for explanation of the	Schedules in the	Universal Decimal Classification
(261.268)	English Channel:	(481)	Norway:
,	Čarinocythereis carinata; 97–102	,	Beyrichia (Sagenabeyrichia) siveteri; 57–64
(261.4)	North-West Atlantic:	(492)	Netherlands:
	Muellerina lıazeli; 21–24	44	_ Spinoleberis eximia; 37–40
(265.72)	South China Sea:	(496.1)	Turkey:
(111)	Cathaycythere reticulata; 1–4	(540)	Carinocythereis carinata; 97–102
(411)	Scotland:	(510)	China:
	Bythocythere zetlandica; 69–72		Abrotocythere ovata; 115–118
	Carinocythereis carinata; 97–102		Abrotocythere quinquicornis; 111–114
(415)	Glyptolichvinella spiralis; 139–142		Albileberis sinensis; 9–12
(415)	Ireland:		Leucocythere plena; 123–126
	Bythocythere intermedia; 65–68		Leucocythere weiningensis; 119–122
(420)	Bythocythere zetlandica; 69–72 England:		Limnocythere xinanensis; 127–130
(420)	Carinocythereis whitei; 103–110		Metacypris aphthosa; 131–134 Sinocythere sinensis; 5–8
	Glyptoliclivinella spiralis; 139–142		Sinocytheridea impressa; 13–16
	Kuiperiana robusta; 73–76	(520)	Japan:
	Loxocauda subquadrata; 77–80	(320)	Howeina camptocytheroidea; 33–36
	Sagmatocythere alaefortis alaefortis; 85–88	(564.3)	Cyprus:
	Sagmatocythere minuta; 81–84	(501.5)	Carinocythereis whitei; 103–110
	Sagmatocythere wyatti; 93–96		Morocco:
(429)	Wales:		Calocaria maurae; 45–48
	Carinocythereis whitei; 103-110	(65)	Algeria:
(437)	Czechoslovakia:	` /	Maghrebeis tuberculata; 29–32
	Spinohippula esurialis; 49–56	(669)	Nigeria:
(44)	France:		Beninea ibecetenensis; 135–138
	Healdianella? aremorica; 25–28	(798)	Alaska:
	Kovalevskiella caudata; 41–44		Pterygocythereis vannieuwenhuisei; 17–20
	Sagmatocythere.alaefortis gallica; 89–92	(941)	Australia:
(45)	Italy:		Glyptolichvinella ovicella; 43–148
	Carinocythereis carinata; 97–102		
	Carinocythereis whitei; 103–110		

Stereo-Atlas of Ostracod Shells: Vol. 14, Part 2

CONTENTS

- 14 (17) 73- 76 On *Kuiperiana robusta* Whatley & Maybury sp. nov.; by R. C. Whatley & C. Maybury
- 14 (18) 77-80 On Loxocauda subquadrata Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley
- 14 (19) 81– 84 On Sagmatocythere minuta Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley
- 14 (20) 85– 88 On Sagmatocythere alaefortis alaefortis Whatley & Maybury sp. nov.; by R. C. Whatley & C. Maybury
- 14 (21) 89– 92 On Sagmatocythere alaefortis gallica Whatley & Maybury subsp. nov.; by R. C. Whatley & C. Maybury
- 14 (22) 93- 96 On Sagmatocythere wyatti Maybury & Whatley sp. nov.; by C. Maybury & R. C. Whatley
- 14 (23) 97-102 On Carinocythereis carinata (Roemer); by J. Athersuch & J. E. Whittaker
- 14 (24) 103-110 On Carinocythereis whitei (Baird); by J. Athersuch &. J. E. Whittaker
- 14 (25) 111-114 On Abrotocythere quinquicornis Zhao gen. et sp. nov.; by Zhao Yuhong
- 14 (26) 115-118 On Abrotocythere ovata Zhao sp. nov.; by Zhao Yuhong
- 14 (27) 119-122 On Leucocythere weiningensis Zhao sp. nov.; by Zhao Yuhong
- 14 (28) 123-126 On Leucocythere plena Zhao sp. nov.; by Zhao Yuhong
- 14 (29) 127-130 On Limnocythere xinanensis Zhao sp. nov.; by Zhao Yuhong
- 14 (30) 131-134 On Metacypris aphthosa Zhao sp. nov.; by Zhao Yuhong
- 14 (31) 135-138 On Beninea ibecetenensis Apostolescu gen. et sp. nov.; by V. Apostolescu
- 14 (32) 139-142 On Glyptolichvinella spiralis (Jones & Kirkby); by R. F. Lundin
- 14 (33) 143–148 On Glyptolichvinella ovicella Lundin & Visintainer sp. nov.; by R. F. Lundin & L. M. Visintainer
- 14 (34) 149-151 Index for Volume 14, 1987

Prepaid annual subscription (valid for Volume 15, 1988)

Individual subscription £22.00 or US \$50.00 for 2 parts (post free)

Price per Part: £22.00 or US \$50.00

Institutional subscription £45.00 or US \$80.00 for 2 parts (post free)
Price per Part: £45.00 or US \$80.00

Back volumes: Vol. 1 (4 Parts): £20.00; price per Part: £5.00

Vol. 2 (4 Parts): £28.00; price per Part: £7.00

Vol. 3 (2 Parts): £24.00; price per Part: £12.00

Vol. 4 (2 Parts): £30.00; price per Part: £15.00

Vol. 5 (2 Parts): £32.00; price per Part: £16.00

Vol. 6 (2 Parts): £40.00; price per Part: £20.00

Vol. 7 (2 Parts): £40.00; price per Part: £20.00

Vol. 8 (2 Parts): £60.00; price per Part: £30.00

Vol. 9 (2 Parts): £60.00; price per Part: £30.00

Vol. 10 (2 Parts): £60.00; price per Part: £30.00

Vol. 11 (2 Parts): £60.00; price per Part: £30.00

Vol. 12 (2 Parts): £60.00; price per Part: £30.00

Vol. 13 (2 Parts): £60.00; price per Part: £30.00

Vol. 14 (2 Parts): £60.00; price per Part: £30.00

Postage extra in sales of all back Parts
No trade discount is allowed on the subscription rate

Orders should be addressed to: Dr J. E. Whittaker,

Department of Palaeontology, British Museum (Natural History), Cromwell Road, South Kensington,

London SW7 5BD.

Cheques should be made payable to B.M.S. (Stereo-Atlas Account)

SPECIAL OFFER

50% off all back part prices if you become a subscriber to the Atlas ISSN 0952-7451

A Stereo-Atlas of Ostracod Shells

edited by J. Athersuch, D. J. Horne, J. W. Neale, and David J. Siveter



Published by the British Micropalaeontological Society, London 1988

A Stereo-Atlas of Ostracod Shells

Published by the British Micropalaeontological Society, London

OUR AIMS

Volume 1 of A Stereo-Atlas of Ostracod Shells was published in 1973. Our policy is to describe ostracod species from all geological ages from Cambrian to Recent and from all parts of the globe.

We adopt a publishing philosophy which was introduced in Volume 1 under the title "The New Palaeontography". The format of the Atlas gives flexibility. We publish on a page ruled for cutting into standard sized cards that can be sorted and arranged at the whim of the reader. Each taxon is lavishly illustrated in three dimensional representation and printed in high resolution format. Every taxon is classified with a code prescribed by the Universal Decimal system that can be fed into a computer and render possible the storage and retrieval of taxonomic, geographical, stratigraphical, and ecological data.

SUBSCRIPTION and ORDERS

Prepaid annual subscription (valid for Volume 15, 1988)
Individual subscription £22.00 or US \$50.00 for 2 parts (post free)
Price per Part: £22.00 or US \$50.00
Institutional subscription £45.00 or US \$80.00 for 2 parts (post free)
Price per Part: £45.00 or US \$80.00

Back volumes: Vol. 1 (4 Parts): £20.00; price per Part: £5.00

Vol. 2 (4 Parts): £28.00; price per Part: £7.00

Vol. 3 (2 Parts): £24.00; price per Part: £12.00

Vol. 4 (2 Parts): £30.00; price per Part: £15.00

Vol. 5 (2 Parts): £32.00; price per Part: £16.00

Vol. 6 (2 Parts): £40.00; price per Part: £20.00

Vol. 7 (2 Parts): £40.00; price per Part: £20.00

Vol. 8 (2 Parts): £60.00; price per Part: £30.00

Vol. 9 (2 Parts): £60.00; price per Part: £30.00

Vol. 10 (2 Parts): £60.00; price per Part: £30.00

Vol. 11 (2 Parts): £60.00; price per Part: £30.00

Vol. 12 (2 Parts): £60.00; price per Part: £30.00

Vol. 13 (2 Parts): £60.00; price per Part: £30.00 Vol. 14 (2 Parts): £60.00; price per Part: £30.00

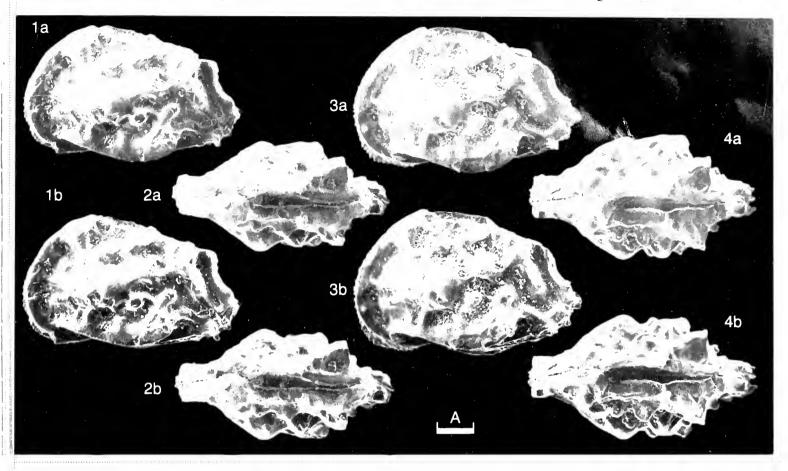
Postage extra in sales of all back Parts
No trade discount is allowed on the subscription rate

Orders should be addressed to: Dr J. E. Whittaker,
Department of Palaeontology, British Museum (Natural History),
Cromwell Road, South Kensington,
London SW7 5BD.
Cheques should be made payable to B.M.S. (Stereo-Atlas Account)

INSTRUCTIONS TO AUTHORS

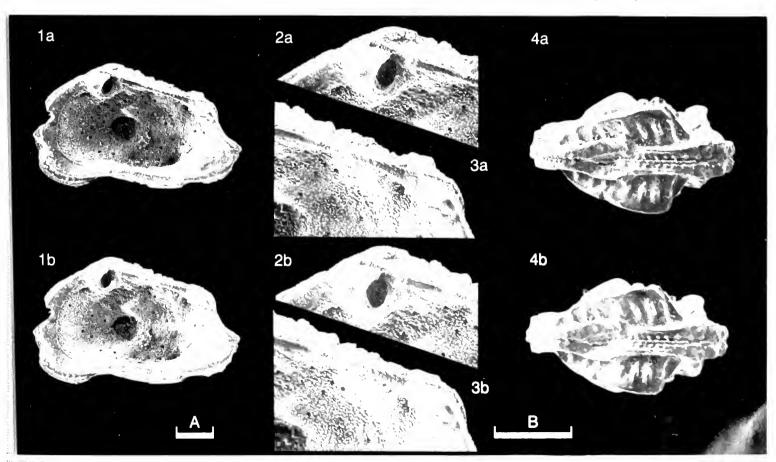
Contributions illustrated by scanning electron micrographs of Ostracoda in stereo-pairs are invited. Full instructions may be obtained on request from any one of the Editors. Format should follow the style set by the papers in recent issues. Descriptive matter apart from illustrations should be cut to a minimum; preferably each plate should be accompanied by one page of text only. Blanks to aid in mounting figures for plates may be obtained from the Editors to whom completed manuscripts should be sent:

Dr J. Athersuch, Stratigraphy Branch, British Petroleum, Chertsey Road, Sunbury-on-Thames, Middlesex TW16 7LN. Dr D. J. Horne, School of Earth Sciences, Thames Polytechnic, Walburgh House, Bigland Street, London E1 2NG. Prof J. W. Neale, Department of Geology, The University, Hull HU6 7RH. Dr David J. Siveter, Department of Geology, The University, Leicester LE1 7RH.



Stereo-Atlas of Ostracod Shells 15, 44

Quadracythere keeni (4 of 4)



Species published in Volumes 1-14

Ordovician

Airina amabilis (Neckaja) Antiaechmina pseudovelata Schallreuter Bilobatia serralobata Schallreuter Bolbihithis abdominalis Schallreuter Bollia delgadoi Vannier Braderupia asymmetrica (Neckaja) Brephocharieis complicata (Salter) Brevibolbina dornbuschi Schallreuter Bromidella sarvi Schallreuter Bulbosclerites unicornis (Neckaja) Byrsolopsina manca Schallreuter Caprabolbina capra Schallreuter Concayhithis latosulcatus Schallreuter Cryptophyllus gutta Schallreuter Distobolbina bispinata Schallreuter Distobolbina grekoffi Schallreuter Disulcina syltensis Schallreuter Duplicristatia asymmetrica Schallreuter Duringia spinosa (Knüpfer) Duringia triformosa Jones Eographiodactylus sulcatus Schallreuter Eolomattella bicuspidata Schallreuter Femerensia gealbertii Schallreuter Foramenella parkis (Neckaja) Gellensia nodoreticulata Schallreuter Gotula gotlandica (Schallreuter) Harperopsis scripta (Harper) Hastatellina normandiensis Pribyl Henningsmoenia costa Orr Henningsmoenia gunnari (Thorslund) Hippula (Cetona) turris (Schallreuter)

Hithis colonus Schallreuter & Siveter

Homeokiesowia frigida (Sarv)
Homeokiesowia epicopa Siveter
Karimutatia crux Schallreuter
Kiesowia (Kiesowia) dissecta (Krause)
Klimphores planus Schallreuter
Kroemmelbeinia valensis Schallreuter
Lomatobolbina vonhachtorum Schallreuter
Miehlkella cribroporata Schallreuter
Naevhithis naevus Schallreuter
Pachydomelloides braderupensis Schallreuter

Piretopsis (Cerninella) bohemica (Barrande)
Platybolbina (Reticulobolbina)spongiosoreticulata
Schallreuter

Platybolbina runica Schallreuter & Krůta
Pyxion posterobicarinatum Schallreuter
Quadritia (Krutatia) iunior Schallreuter
Raimbautina hammanni Vannier
Reginea reginae Schallreuter
Retinoda sulcata (Knüpfer)
Schallreuteria (Lippea) lippensis Schallreuter
Schallreuteria (Schallreuteria) superciliata (Reed)
Spinohippula esurialis Vannier, Krůta & Marek
Tallinnellina dissita Schallreuter & Siveter
Tetradella egorowi Neckaja
Tetradella separata Sidaraviciene
Tetradella pentaloculata Schallreuter

Tetradella pentaloculata Schallreuter
Tetradella? triloculata Schallreuter
Thibautina rorei Vannier
Uscopria memoria Schallreuter
Vittella fecunda Siveter
Wehrlia olbertzae Schallreuter

Silurian

Beyrichia (Altibeyrichia) kiaeri Henningsmoen Beyrichia (Sagenabeyrichia) Pollicot Calocaria maurea Vannier Craspedobolbina (Mitrobeyrichia) impendens (Haswell) Sleia troglodytophila Martinsson Slependia armata (Henningsmoen) Strepula concentrica Jones and Holl Xystista auricularis (Jones) Xystista graffhami (Lundin)

Devonian

Glyptolichvinella ovicella Lundin & Visintainer

Renibeyrichia mulciber Siveter

Carboniferous

Carbonita corrugata Gregory
Glyptolichvinella spiralis (Jones & Kirkby)

Healdianella? aremorica Crasquin Kellettina carnica Ruggieri & Siveter

Triassic

Triadocypris spitzbergensis Weitschat

Jurassic

Angliaecytheridea calvata Sheppard Bairdia aselfingenensis Lord and Moorley Bairdia hahni Lord and Moorley Cytherelloidea longicostata Sheppard Marslatourella dorsispinata Bate & Stephens Micropneumatocythere brendae Sheppard Micropneumatocythere crassa (Peterson) Micropneumatocythere falcata Sheppard Eocytheridea kirtlingtonensis Bate & Mellish

Galliaecytheridea elegans (Sharapova)

Galliaecytheridea gorodischensis Fuller & Lord

Galliaecytheridea miranda (Lyubimova)

Galliaecytheridea volgaensis (Lyubimova)

Gammacythere ubiquita Malz & Lord

Glyptocythere costata Bate

Glyptocythere oscillum Jones & Sherborn

Glyptocythere penni Bate & Mayes

Glyptocythere polita Bate

Glyptocythere raasavensis Stevens

Glyptocythere scitula Bate

Kinkelinella malzi (Dépêche)

Lesleva bathonica Bate

Lophocythere (Lophocythere) ostreata (Jones & Sherborn)

Lophocythere (Neurocythere) bradiana (Jones) Lophocythere (Neurocythere) minuta (Peterson)

Mandelstamia ventrocornuta (Sharapova)

Marslatourella bullata Bate

Micropneumatocythere tumida Stephens & Ware Monoceratina scrobiculata Triebel & Bartenstein

Ogmoconcha ambo Lord and Moorley

Ogmoconcha eocontractula Park

Oligocythereis kostytschevkaensis (Lyubimova)

Procytheridea exempla Peterson

Procytheridea fraudator Sherrington & Lord Procytheropteron prolongatum (Sharapova)

Progonocythere reticulata Bate

Progonocythere stilla Sylvester-Bradley

Ptychobairdia limbata Sheppard

Terquemula robusta Sheppard

Theriosynoecum bathonicum Sylvester-Bradley

Theriosynoecum kirtlingtonense Bate

Theriosynoecum wyomingense (Branson)

Timiriasevia mackerrowi Bate Timiriasevia punctata Clements

Trachycythere munita Sylvester-Bradley

Cretaceous

Acrocythere hauteriviana (Bartenstein)

Apatocythere spinosa Neale

Apateloschizocythere geniculata Bate

Archeocosta alkazwinii Al-Bashir & Keen

Beninea ibecetenensis Apostolescu

Cythereis lindiensis Bate

Donmacythere damottae (Colin)

Iberocypris cornutus Babinot

Kefiella maresi Donze & Said

Maghrebeis tuberculata Majoran

Navarracythere estellensis Colin & Rodriguez-Lazaro

Nigeroloxoconcha oyesesei Reyment

Nigeroloxoconcha oniseguni Reyment

Paracandona occitanica Babinot & Tambareau

Paranotacythere speetonensis (Neale)

Paranotacythere (Paranotacythere) magnifica Lomax

Parapokornyella taxyae (Babinot)

Pennyella pennyi Neale

Pattersoncypris micropapillosa Bate

Philomedes donzei Neale

Sarlatina merlensis Babinot & Colin

Schuleridea bilobata (Triebel)

Schuleridea hammi (Triebel)

Schuleridea juddi Neale Schuleridea lamplughi Neale

Schuleridea praethoerenensis Bartenstein & Brand

Schuleridea rhomboidalis Neale

Spinoleberis eximia (Bosquet)

Stravia crossata Neale

Theriosynoecum fittoni (Mantell)

Veenia (Nigeria) nigeriensis Reyment

Tertiary

Abrotocythere quinquicornis Zhao

Abrotocythere ovata Zhao

Acanthocythereis decoris Siddiqui

Acanthocythereis dohukensis Khalaf

Acanthocythereis hystrix (Reuss)

Actinocythereis iragensis Khalaf

Ambostracon costaforma Whatley & Maybury

Ambostracon delicata Whatley & Maybury

Ambostracon europea Maybury & Whatley

Ambostracon perfecta Maybury & Whatley

Argenticytheretta (Argenticytheretta) fuegoensis Rose

Argenticytheretta (Argenticytheretta) gonzalezi Rose

Argenticytheretta (Argenticytheretta) patagoniensis Rose

Argenticytheretta (Argenticytheretta) punctata Rose

Argenticytheretta (Argenticytheretta) riescoensis Rose

Argenticytheretta (Chiliella) brunswickensis Rose

Argenticytheretta (Magallanella) chileana Rose Bairdia beraguaensis Singh & Tewari

Bairdia jammuensis Singh & Tewari

Bairdia kalakotensis Singh & Tewari

Bairdoppilata kalakotensis Singh & Tewari

Callistocythere mediterranea (Müller)

Kovałevskielła caudata (Lutz)

Kuiperiana robusta Whatley & Maybury

Leguminocythereis chittagongensis Neale & Ahmed

Leocytheridea polleti Keen

Leptocythere multipunctata (Seguenza)

Loculicytheretta (Heptaloculites) cavernosa (Apostolescu

& Magne)

Loculicytheretta (Heptaloculites) semirugosa (Apostolescu

& Magne)

Loculicytheretta (Heptaloculites) sp. A

Loculicytheretta payonia (Brady)

Loxocauda subquadrata Maybury & Whatley

Mutihus albicans Ruggieri

Mutilus cimbaeformis (Seguenza)

Mutilus convexus (Baird)

Mutilus elegantulus Ruggieri & Sylvester-Bradley

Mutilus freudenthali (Sissingh)

Mutilus keiji Ruggieri

Mutilus retiformis (Terquem)

Mutilus speyeri (Brady)

Nigeroloxoconcha oniseguni Reyment

Nigeroloxoconcha ovesesei Revment

Callistocythere montana Doruk Carinocythereis antiquata (Baird) Carinocythereis carinata (Roemer) Chrysocythere cataphracta Ruggieri Chrysocythere nagibi Khalaf

Chrysocythere naqibi Khalaf Chrysocythere paradisus Doruk Cistacythereis equivalis Doruk Cistacythereis pokornyi (Ruggieri)

Costa batei (Brady)
Costa edwardsii (Roemer)
Costa himchariensis Ahmed
Costa obliquifossa Ahmed
Costa punctatissima Ruggieri

Costa trudis Ahmad

Cyamocytheridea contracta Doruk Cyamocytheridea meniscus Doruk Cyamocytheridea obstipa Doruk Cyamocytheridea polygona Doruk Cyprideis exuberans van Harten Cyprideis undosa van Harten

Cytherella (Cytherelloidea) chosta Doruk
Cytherella (Cytherelloidea) glypta Doruk
Cytherella (Cytherelloidea) obolus Doruk
Cytherella (Cytherelloidea) ochthodes Doruk
Cytherella (Cytherelloidea) petrosa Doruk
Cytherella (Cytherelloidea) scarabaeus Doruk

Cytherella postdenticulata Oertli

Cytheretta (Cytheretta) jurinei (V. Münster) Cytheretta (Cytheretta) semipunctata (Bornemann)

Cytheretta (Flexus) plicata (V. Münster)

Cytheridea (Cytheridea) muelleri muelleri (V. Münster) Cytheridea (Cytheridea) muelleri toenisbergensis Weiss

Cytheridea (Cytheridea) pernota Oertli & Keij

Hamanella implexa Finger
Haplocytheridea debilis (Jones)
Haplocytheridea mantelli Keen
Hermanites transversicostata Khalaf
Hornibrookella anna (Lienenklaus)
Howeina camptocytheroidea Hanai

Keijella clauda Doruk Keijella dolabrata Doruk Keijella hodgii (Brady) Keijella procera Doruk Ommatokrithe prolata Ahmad Orionina bireticulata Doruk Orionina tegminata Doruk

Paijenborchella (Eopaijenborchella) malaiensis cymbula Ruggieri

Paijenborchella (Eopaijenborchella) mouliana (Sissingh)

Palmoconcha laevata (Norman) Paracypris jhingrani Singh & Tewari Paracytheridea anapetes Ahmad Paracytheridea inscita Doruk

Paracytheromorpha rimafossa Maybury & Whatley

Paragrenocythere biclavata Al-Furaih Phalcocythere horrescens (Bosquet) Pokornyella mersondaviesi (Latham) Ptervgocythereis vannieuwenhuisei Brouwers

Radimella confragosa (Edwards)

Roundstonia magna Maybury & Whatley Roundstonia minima Whatley & Maybury

Sagmatocythere alaefortis alaefortis Whatley & Maybury Sagmatocythere alaefortis gallica Whatley & Maybury

Sagmatocythere minuta Maybury & Whatley Sagmatocythere paracercinata Whatley & Maybury

Sagmatocythere pseudomultifora Maybury & Whatley

Sagmatocythere wyatti Maybury & Whatley

Sarsicytheridea kempfi Weiss Sarsicytheridea lienenklausi (Kuiper) Schizocythere buendensis Triebel

Schuleridea (Aequacytheridea) oculata Moos Semicytherura incongruens (G. W. Müller)

Semicytherura ruggierii (Pucci) Sinocytheridea impressa (Brady) Sylvestra posterobursa Doruk Tanzanicythere pterota Ahmad

Thaerocythere anisomorphica Whatley & Maybury Thaerocythere irregulare Whatley & Maybury Thaerocythere pieta Maybury & Whatley Thaerocythere regulare Maybury & Whatley

Togoina attitogonensis Apostolescu Triebelina raripila (G. W. Müller) Urocythereis favosa (Roemer) Urocythereis labyrinthica Uliczny Urocythereis seminulum (Seguenza) Waiparacythereis joanae Swanson

Quaternary & Recent

Abrotocythere quinquicornis Zhao Acanthocythereis hystrix (Reuss)

Albileberis sinensis Hou Atjehella kingmai Keij Aurila woodwardii (Brady) Aurila woutersi Horne

Australicythere polylyca (Müller) Baffinicythere howei Hazel Bathycythere vanstraateni Sissingh Bennelongia tunta De Deckker

Bennerong a tuntal De Deckker
Bonnyanne la robertsoni (Brady)
Bythoceratine scaberrima (Brady)
Bythocythere intermedia Elofson

Bythocythere zetlandica Athersuch, Horne & Whittaker

Caboncypris nunkeri De Deckker Callistocythere badia (Norman)

Loxoconcha affinis (Brady) Loxoconcha alata Brady

Loxoconcha amygdalanux Bate & Gurney Loxoconcha conjugalis Athersuch

Loxoconcha conjugalis Athersuch Loxoconcha dimorpha Hartmann Loxoconcha elliptica Brady Loxoconcha linleyi Horne

Loxoconcha malcomsoni Horne & Robinson Loxoconcha multiornata Bate & Gurney Loxoconcha nea Barbeito-Gonzalez

Loxoconcha ovulata (Costa)
Loxoconcha pontica Klie
Loxoconcha rhomboidea (Fischer)

Loxoconcha rubritincta Ruggieri Loxoconcha stellifera G. W. Müller Loxoconcha undulata Al-Furaih Callistocythere crispata (Brady) Callistocythere littoralis (Müller) Callistocythere mediterranea (Müller) Callistocythere murrayi Whittaker Carinocytliereis antiquata (Baird) Carinocythereis carinata (Roemer) Carinocythereis whitei (Baird)

Cathaycythere reticulata Whatley & Zhao Cativella bensoni Neale

Celtia quadridentata (Baird) Centrocypris viridis Neale Cluthia keiji Neale Costa batei (Brady) Costa edwardsii (Roemer) Costa punctatissima Ruggieri Cypretta yapinga De Deckker Cyprideis torosa (Jones) Cypris decaryi Gauthier

Cypris latissima (G. W. Müller) Cypris subglobosa J. de C. Sowerby Cytherelloidea bonanzaensis Keij Cytheretta adriatica Ruggieri Cytheretta judaea (Brady) Çytlıeretta teslıekpukensis Swain

Cytheropteron brastadensis Lord Cytheropteron elofsoni Lord Cytherura gibba (O. F. Müller) Elofsonia baltica (Hirschmann)

Elofsonia pusilla (Brady & Robertson) Eucypris fontana (Graf)

Eucythere anglica Brady Eucythere argus (Sars) Eucythere declivis (Norman)

Eucytliere prava Brady & Robertson

Gambiella caelata Witte Gliardaglaia anibigua Neale

Hemicypris bairdi Martens & Wouters Hemicypris dentatomarginata (Baird)

Hemicythere villosa (Sars)

Hemicytherura aegyptica Hartmann Hemicytherura cellulosa (Norman) Hemicytherura hoskini Horne Hemicytherura videns (Müller)

Heterocyprideis macrotuberculata Masson & Whatley

Heterocypris luzonensis Neale Heterocythereis albomaculata (Baird) Heterocythereis voraginosa Athersuch Hiltermannicythere emaciata (Brady) Hiltermannicytliere rubra (Müller) Hirschmannia viridis (O. F. Müller) Howeina camptocytlieroidea Hanai *Hyocypris monstrifica* (Norman) Ilyocypris schwarzbachi Kempf Ilyocypris taprobanensis Neale

Ilyocypris quinculminata Sylvester-Bradley

Leptocytliere baltica Klie

Leptocythere multipunctata (Seguenza)

Leptocythere porcellanea (Brady) Leucocythere plena Zhao Leucocythere weiningensis Zhao Limnocythere shixiaensis (Wang) Limnocythere xinanensis Zhao Lindisfarnia guttata (Norman) Loculicytheretta pavonia (Brady)

Loxoreticulatum fallax (G. W. Müller) Metacypris aphthosa Zhao Muellerina hazeli Coles & Cronin Mutilus cimbaeformis (Seguenza) Mutilus convexus (Baird) Mutilus speyeri (Brady) Nannocythere pavo (Malcomson) Notiocypridopsis frigogena (Graf)

Oncocypris pustulosa Gurney Paijenborchellina alata Gurney Paijenborchellina venosa Gurney Pallmoconcha laevata (Norman) Paracytheridea cuneiformis (Brady)

Paracytheridea hexalpha Doruk Paralimnocythere vulgaris McKenzie & Swanson

Patagonacythere devexa (Müller)

Pelecocythere sylvesterbradleyi Athersuch

Polycope choane Hasan Polycope foraminosa Hasan Polycope frigida Neale Polycope regina Hasan

Procythereis iganderssoni (Skogsberg) Procythereis torquata (Skogsberg) Propontocypris pirifera (G. W. Müller) Propontocypris trigonella (Sars)

Pterygocythereis jonesii (Baird) Pterygocythereis siveteri Athersuch Puriana fissispinata Benson & Coleman Puriana pacifica Benson

Quadraleberis exquisita Bate & Sheppard Radimella? aurita (Skogsberg) Radimella darwini Pokorný Radimella dictyon Pokorný

Radimella? floridana (Benson & Coleman)

Reticypris pinguis De Deckker Robertsonites tuberculatus (Sars)

Rockallia enigmatica Whatley, Frame & Whittaker

Sagmatocythere napoliana (Puri) Scottia audax (Chapman) Semicytherura cornuta (Brady) Semicytherura exudata Doruk

Semicytherura incongruens (G. W. Müller)

Semicytherura nigrescens (Baird) Semicytherura ruggierii (Pucci) Semicytherura sella (Sars)

Semicytherura sulcata (G. W. Müller) Semicytherura tela Horne & Whittaker Sinocytheridea impressa (Brady) Sinocytheridea sinensis Hou Stenocypris fernandoi Neale

Sulcostocythere knysnaensis Benson & Maddocks

Tanganyikacypris matthesi Kiss Trachyleberis scabrocuneata (Brady) Triebelina raripila (G. W. Müller) Tyrrhenocythere amnicola (Sars) *Urocythereis favosa* (Roemer) Urocythereis labyrinthica Uliczny

Urocythereis phantastica Athersuch & Ruggieri

Xestoleberis aurantia (Baird) Xestoleberis nitida (Liljeborg)

Xestoleberis postangulata Bate & Sheppard

Xestoleberis rubens Whittaker

Zabythocypris redunca Athersuch & Gooday

Type-species of genera and subgenera

Abrotocythere quinquicornis Zhao Acrocythere hauteriviana (Bartenstein)

Albileberis sinensis Hou

Angliaecytheridea calvata Sheppard Apateloschizocythere geniculata Bate Archeocosta alkazwinii Al-Bashir & Keen

Argenticytheretta (Chiliella) brunswickensis Rose

Argenticytheretta (Magallanella) chileana Rose

Australicythere polylyca (Müller) Baffinicythere howei Hazel Bathycythere vanstraateni Sissingh Beninea ibecetenensis Apostolescu

Beyrichia (Sagenabeyrichia) siveteri Pollicot

Bilobatia serralobata Schallreuter Bonnyannella robertsoni (Brady) Braderupia asymmetrica (Neckaja) Brephocharieis complicata (Salter) Caboncypris nunkeri De Deckker Callistocythere littoralis (Müller) Calocaria maurae Vannier Caprabolbina capra Schallreuter

Carinocythereis carinata (Roemer) Cathaycythere reticulata Whatley & Zhao

Celtia quadridentata (Baird) Chrysocythere cataphracta Ruggieri Concavhithis latosulcatus Schallreuter

Costa edwardsii (Roemer) Cyprideis torosa (Jones) Cytheretta judaea (Brady)

Cytheretta (Flexus) plicata (V. Münster)

Cytheridea (Cytheridea) muelleri (V. Münster)

Cytherura gibba (O. F. Müller) Donniacythere damottae (Colin) Duplicristatia asymmetrica Schallreuter

Duringia spinosa (Knüpfer) Elofsonia baltica (Hirschmann) Eucythere declivis (Norman) Femerensia gealbertii Schallreuter Foramenella parkis (Neckaja) Gambiella caelata Witte

Gammacythere ubiquita Malz & Lord Glyptolichvinella spiralis (Jones & Kirkby)

Gotula gotlandica (Schallreuter) Hamanella implexa Finger Harperopsis scripta (Harper) Hemicythere villosa (Sars)

Hemicytherura cellulosa (Norman) Henningsmoenia gunnari (Thorsland) Heterocythereis albomaculata (Baird) Hirschmannia viridis (O. F. Müller) Homeokiesowia frigida (Sarv) Hornibrookella anna (Lienenklaus) Howeina camptocytheroidea Hanai

Iberocypris cornutus Babinot Karinutatia crux Schallreuter Kefiella maresi Donze & Said Keijella hodgii (Brady)

Kiesowia (Kiesowia) dissecta (Krause)

Klimphores planus Schallreuter Leocytheridea polleti Keen Lesleya hathmica Bate Lindisfarnia le evata (Norman) Loculicytheretta pavonia (Brady)

Lophocythere (Lophocythere) ostreata (Jones & Sherborn)

Lophocythere (Neurocythere) bradiana (Jones)

Loxoconcha rhomboidea (Fischer) Loxoreticulatum fallax (G. W. Müller) Maghrebeis tuberculata Majoran Miehlkella cribroporata Schallreuter Mutilus retiformis (Terquem) Naevhithis naevus Schallreuter Nannocythere payo (Malcomson)

Navarracythere estellensis Colin & Rodriguez-Lazaro

Nigeroloxoconcha oniseguni Reyment Notiocypridopsis frigogena (Graf) Ommatokrithe prolata Ahmad

Paracytheromorpha rimafossa Maybury & Whatley

Paragrenocythere biclavata Al-Furaih Parapokornyella taxyae (Babinot) Pattersoncypris micropapillosa Bate Pelecocythere sylvesterbradleyi Athersuch

Pennyella pennyi Neale

Phalcocythere horrescens (Bosquet)

Piretopsis (Cerninella) bohemica (Barrande)

Procythereis torquata (Skogsberg) Procytheridea exempla Peterson Progonocythere stilla Sylvester-Bradley Propontocypris trigonella (Sars) Pterygocythereis jonesii (Baird)

Quadraleberis exquisita Bate & Sheppard Quadritia (Krutatia) iunior Schallreuter

Radimella dictyon Pokorný Raimbautina hammanni Vannier Reginea reginae Schallreuter Renibeyrichia mulciber Siveter Retinoda sulcata (Knüpfer) Robertsonites tuberculatus (Sars)

Rockallia enigmatica Whatley, Frame & Whittaker

Sagmatocythere napoliana (Puri) Sarlatina merlensis Babinot & Colin

Schallreuteria (Lippea) lippensis Schallreuter Schallreuteria (Schallreuteria) superciliata (Reed)

Semicytherura nigescens (Baird) Sinocythere sinensis Hou Sinocytheridea impressa (Brady) Slependia armata (Henningsmoen)

Spinohippula esurialis Vannier, Krůta & Marek

Spinoleheris eximia (Bosquet) Stravia crossata Neale

Strepula concentrica Jones & Holl

Sulcostocythere knysnaensis Benson & Maddocks

Sylvestra posterobursa Doruk Tanganyikacypris matthesi Kiss *Tanzanicythere pterota* (Ahmad)

Theriosynoecum wyomingense (Branson)

Thibautina rorei Vannier

Togoina attitogonensis Apostolescu Trachyleberis scabrocuneata (Brady) Triadocypris spitzbergensis Weitschat Tyrrhenocythere amnicola (Sars) *Urocythereis favosa* (Roemer) Uscopria memoria Schallreuter Veenia (Nigeria) nigeriensis Reyment Waiparacythereis joanae Swanson Wehrlia olbertzae Schallreuter Xestoleheris nitida (Liljeborg)

Xystista auricularis (Jones)

